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(54) Title: MORPHOGEN-INDUCED PERIODONTAL TISSUE REGENERATION

(57) Abstract

Disclosed are methods and compositions for inducing periodontal tissue morphogenesis in a mammal which include a therapeutically effective concentration of a morphogen. The methods and compositions are useful for integrating an implanted tooth in a tooth socket and for inhibiting tissue loss associated with periodontal disease or injury.

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MORPHOGEN-INDUCED PERIODONTAL TISSUE REGENERATION Background of the Invention

This invention relates generally to the dental arts and 5 more specifically to methods and compositions for treating and regenerating periodontal tissue.

The peridontium is the cushioning tissue which anchors the tooth root to the mandibular or maxillar jawbone tissue by suspending the tooth in the tooth socket ("alveolus"). Periodontal tissue includes both the periodontal ligament, a collagen-containing tissue that is in contact with the bone tissue, and cementum, a mineralized tissue that covers the dental root surface. These two hard tissues are connected through the periodontal ligament fibers that run in a perpendicular direction to the two surfaces and thereby serve to anchor and suspend the tooth in the tooth socket, providing a shock-absorptive cushion between the tooth and the jawbone that accommodates the pressure applied to teeth when food is being chewed.

Periodontal tissue loss may occur as a result of disease, including infectious diseases (e.g., gingivitis, caused by bacteria), nutritional diseases, e.g., scurvy,

25 resulting from a vitamin deficiency, and a number of neoplastic diseases, including acute leukemia and lymphomas. The diseases are characterized by inflammation, bleeding and ulceration. Periodontal disease also may result from an opportunistic infection, e.g., in an immune-compromised

30 individual. Left untreated, these diseases can cause significant periodontal tissue loss which loosen the tooth and ultimately can result in loss of the tooth and the alveolar bone tissue (periodontitis.) Chronic periodontitis is the primary cause of tooth loss in adults. Current

35 treatments include professional cleaning to remove plaque

and tartar, use of oral antiseptics, local and/or systemic antibiotic therapies, and/or surgical procedures to remove periodontal pockets formed from periodontal tissue lesions and necrosis. Typically, where a tooth has been lost as a result of periodontitis, a prosthetic tooth or removable bridge is substituted for the natural tooth.

Periodontal tissue loss also may occur as a result of mechanical injury to the tissue or to the tooth itself,

10 particularly one causing tooth loss. Tooth loss also may occur as a result of any of a number of dental diseases,

e.g., dental caries, pulpitis, or osteomyelitis.

A viable tooth can be reimplanted if implantation occurs quickly after loss, e.g., within thirty minutes, and if the periodontal tissue within the tooth socket is still healthy. However, if a significant period of time is allowed to elapse, the living periodontal tissue lining the tooth socket will be resorbed. In addition, the tooth itself begins to degenerate and a prosthetic tooth or removable bridge must be implanted. In the absence of healthy periodontal tissue the prosthetic implant is integrated directly into the jaw bone tissue in a condition called ankylosis (bone tissue in direct contact with dentin tissue.) The life of such prosthetic tooth implants often is limited due to the absence of viable periodontal tissue to enhance tooth anchoring and to absorb the impact of mastication on the prosthesis.

It is an object of this invention to provide a means for inhibiting periodontal tissue loss, as well as means for inducing regeneration of damaged periodontal tissue.

Another object is to provide means for inhibiting the periodontal tissue damage and tooth loss associated with periodontal and other gum diseases. Yet another object is

to enhance integration of an implanted tooth, including a reimplanted natural tooth or tooth prosthesis, in the tooth socket. Still another object is to promote periodontal tissue growth around an implanted tooth. Another object is to inhibit ankylosis of an implanted tooth or tooth prosthesis.

These and other objects and features of the invention will be obvious from the specification, drawings and claims, which follow.

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Summary of the Invention

The invention provides methods and compositions for inhibiting periodontal tissue loss in a mammal, particularly humans, including regenerating damaged tissue and/or inhibiting additional damage thereto. The methods and compositions of this invention may be used to prevent and/or inhibit tooth loss, as well as to enhance integration of an implanted tooth.

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As used herein, "implanted tooth" includes a natural tooth which has grown naturally in the tooth socket, a natural tooth which is reimplanted in a tooth socket, and a prosthetic tooth, which includes both natural teeth from 15 which the root has been removed and replaced with an inert, biocompatible material, and "complete" prostheses made of natural or synthetic, non dentin-containing materials. all cases, "tooth" refers to a natural or synthetic composition essentially defining the shape of a natural 20 tooth, having a solid tooth body, including a crown and tooth root. "Reimplanted natural tooth" includes both an allogenic tooth, e.g., selected from a tooth bank; and a tooth autologous to the mammal, such as a tooth which has fallen out, been knocked out, or otherwise removed from the 25 individual into which it is now being reimplanted. "Integrated tooth" means an implanted tooth with a living, substantially healthy periodontal tissue, including periodontal ligament and cementum, anchoring the tooth to the jaw bone. "Viable" tissue means living, substantially "Viable tooth" refers to an implanted 30 healthy tissue. natural tooth with a living tooth root. "Periodontium" defines the tissues which surround the tooth in the tooth socket and includes both periodontal ligament and cementum. "Inhibit loss" of periodontal tissue, as used herein, means 35 inhibiting damage to, and/or loss of, periodontal tissue,

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including periodontal ligament and/or cementum, and includes regenerating lost or damaged tissue and/or inhibiting additional damage thereto. "Symptom alleviating cofactor" refers to one or more pharmaceuticals which may be

5 administered together with the therapeutic agents of this invention and which alleviate or mitigate one or more of the symptoms typically associated with periodontal tissue loss. Exemplary cofactors include antibiotics, antiseptics, non-steroidal antiinflammatory agents, anaesthetics and analgesics.

The methods and compositions of this invention include a morphogenic protein ("morphogen"), as described herein, which, when provided to the tooth and/or jawbone surfaces in a tooth socket is capable of inducing periodontal tissue formation where periodontal tissue has been lost or damaged, and enhancing integration of an implanted tooth thereby.

In one aspect, the invention features therapeutic

treatment methods and compositions for inhibiting
periodontal tissue loss in a mammal which include
administering to the individual a therapeutically effective
morphogen at a concentration and for a time sufficient to
regenerate damaged periodontal tissue and/or to inhibit

additional damage thereto.

In another aspect, the invention features therapeutic treatment methods and compositions for inhibiting periodontal tissue loss in a mammal which include

30 administering to the individual a compound that stimulates in vivo a therapeutically effective concentration of an endogenous morphogen in the body of the mammal sufficient to regenerate damaged periodontal tissue and/or to inhibit additional damage thereto. These compounds are referred to herein as morphogen-stimulating agents, and are understood

to include substances which, when administered to a mammal, act on cells of tissue(s) or organ(s) that normally are responsible for, or capable of, producing a morphogen and/or secreting a morphogen, and which cause the endogenous level of the morphogen to be altered. The agent may act, for example, by stimulating expression and/or secretion of an endogenous morphogen. In preferred embodiments, the agent stimulates expression and/or secretion of an endogenous morphogen so as to increase amounts of the morphogen in the alveolar bone, periodontium or cementum tissue cells.

In another aspect, the invention provides methods and compositions for enhancing the integration of an implanted tooth, particularly where the tooth socket is substantially 15 reduced in viable periodontal tissue. In fact, the processes and compositions of the invention work well when a tooth socket has lost 30-50% of the periodontal ligament, and as much as 50-100% of the periodontal ligament. methods and compositions include providing to the tooth or 20 tooth socket surface a therapeutically effective concentration of a morphogen or morphogen-stimulating agent sufficient to induce morphogenesis of periodontal tissue. The implanted tooth may be an implanted tooth which has grown naturally in the socket and which is loose as a result 25 of, for example, mechanical injury or due to a dental or periodontal disease. Alternatively, the implanted tooth may be a lost tooth or a tooth prosthesis which has been reimplanted in a vacant tooth socket. The tooth prosthesis may include a natural tooth from which a damaged or diseased 30 root has been removed and replaced with a biocompatible, biologically inert material, as is created in a root canal procedure. The prosthetic tooth also may be composed of synthetic, non dentin-containing materials.

The morphogen may be provided directly to the tooth surface to be implanted, and/or to the tooth socket to which the tooth is to be implanted. Where the morphogen is to be provided to the tissue socket, it may be provided by topical 5 administration to the tooth socket surface or by local injection to periodontal or alveolar bone tissue associated with the socket. Alternatively, an agent capable of stimulating the production and/or secretion of a therapeutically effective concentration of an endogenous 10 morphogen also may be provided to the tooth or tooth socket. Where the morphogen or morphogen stimulating agent (referred to herein collectively as "therapeutic agent") is provided to the tooth surface, it preferably is dispersed in a biocompatible, bioresorbable carrier, most preferably a 15 carrier capable of retaining the therapeutic agent at the tissue surface and/or providing a controlled delivery of the agent to the tooth socket. The therapeutic agent also may be provided to the tooth socket itself, also preferably in association with a carrier capable of maintaining the agent 20 in the tooth socket, and/or capable of enhancing the controlled delivery of the agent to the socket. carriers include compositions having a high viscosity, such as that provided by glycerol and the like, as well as carrier materials formulated from extracellular matrices 25 and/or which contain laminin, collagen, and/or biocompatible synthetic polymers, such as polybutyric, polylactic, polyglycolic acids and copolymers thereof. In addition, or alternatively, an acellular carrier material may be formulated from bone, dentin, cementum or periodontal tissue 30 by demineralizing and guanidine-extracting the tissue essentially as described herein and/or in international application US92/01968 (WO92/15323). Particularly useful acellular matrices include dentin-derived, periodontal ligament-derived and cementum-derived matrices.

In addition, the tooth to be implanted preferably comprises a porous exterior surface onto which the therapeutic agent may be adsorbed, and into which progenitor and differentiating cementoblasts can infiltrate and proliferate. Useful surfaces include natural tooth root surfaces, and porous prosthetic surfaces, including surfaces composed of matrix materials such as collagen, laminin, biocompatible polymers or metals such as titanium oxide. Where a natural tooth or dentin-containing prosthesis is to be implanted, the surface to be implanted first may be partially demineralized, e.g., by transient exposure to an acid to enhance the porosity of the tooth root surface.

Preferably, where the tooth is to be implanted into a
tooth socket, the socket has been freed of fibrous tissue
which may have formed following tooth loss and periodontal
tissue resorption. For example, the tooth socket may have
undergone a healing period of several months after loss or
removal of the tooth such that scar tissue has formed over
the wound. In this case the healed socket preferably is
surgically prepared for tooth implantation by removing the
scar and other undesired tissue to expose the alveolar bone
surface.

25 Preferably, where the therapeutic agent is to be provided to enhance periodontal tissue viability surrounding an implanted tooth, the therapeutic agent is provided topically to the tissue surfaces between the tooth and gingiva. Alternatively, the agent may be injected locally, 30 e.g., into the gingiva itself.

The morphogens described herein may be used to inhibit periodontal tissue loss and/or to enhance viability of periodontal tissue at risk of damage due to a periodontal disease. The periodontal disease may be caused by an

infectious agent, such as a bacterial, fungal or viral agent, or by a nutritional deficiency, including a vitamin deficiency. The morphogens also may be used to regenerate periodontal tissue lost as a result of a neoplastic disease, including squamous cell carcinomas, acute leukemias, lymphomas and metastatic tumors. A detailed description of diseases which damage or destroy periodontal tissue can be found, for example, in Harrison's Principles of Internal Medicine, 243-248, (McGraw-Hill 12th ed. 1991), the disclosure of which is incorporated herein by reference. The efficacy of the morphogens described herein in modulating an inflammatory response are described in detail in international application US92/07358 (WO93/04692).

at risk for periodontal tissue damage due to periodontal disease, a population most particularly at risk are immune-compromised individuals, such as individuals suffering from autoimmune diseases and/or whose immune system has been suppressed as part of a clinical procedure or therapy. Thus, in another aspect, the invention provides methods and compositions for inhibiting periodontal tissue loss in immune-compromised individuals.

As described in international application W092/15323, and Example 2, below, the morphogens described herein also can induce formation of damaged or lost dentin tissue.

Accordingly, where a natural tooth or dentin-containing prosthesis is to be implanted, a morphogen or morphogenstimulating agent also may be provided to damaged areas of the tooth to induce dentin regeneration of damaged or lost dentin tissue. The morphogen may be provided topically or otherwise administered to the tooth tissue. For example, the morphogen may be dispersed in a biocompatible, porous carrier material that then is provided topically to the

damaged dentin tissue. A useful carrier may be formulated from dentin by demineralizing and guanidine-extracting the tissue to create an acellular matrix.

5 The morphogens and morphogen-stimulating agents also may be provided to the periodontium together with other molecules ("cofactors") known to have a beneficial effect in treating damaged periodontal tissue, particularly cofactors capable of mitigating or alleviating symptoms typically associated with periodontal tissue damage and/or loss. Examples of such cofactors include antiseptics such as chlorohexidine and tibezonium iodide, antibiotics, including tetracycline, aminoglycosides, macrolides, penicillins and cephalosporins, anaesthetics and analgesics, and other non-steroidal anti-inflammatory agents.

Among the morphogens useful in this invention are proteins originally identified as osteogenic proteins (see U.S. Patent 5,011,691, incorporated herein by reference), 20 such as the OP-1, OP-2 and CBMP2 proteins, as well as amino acid sequence-related proteins such as DPP (from Drosophila), Vgl (from Xenopus), Vgr-1 (from mouse), GDF-1 (from mouse, see Lee (1991) PNAS 88:4250-4254), all of which are presented in Table II and Seq. ID Nos. 5-14, and the 25 recently identified 60A protein (from Drosophila, Seq. ID No. 24, see Wharton et al. (1991) PNAS 88:9214-9218.) members of this family, which include members of the TGF-\$\beta\$ super-family of proteins, share substantial amino acid sequence homology in their C-terminal regions. The proteins are translated as a precursor, having an N-terminal signal peptide sequence, typically less than about 30 residues, followed by a "pro" domain that is cleaved to yield the mature sequence. The "pro" form of the protein includes the pro domain and the mature domain, and forms a soluble species that appears to be the primary form secreted from

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cultured mammalian cells. The signal peptide is cleaved rapidly upon translation, at a cleavage site that can be predicted in a given sequence using the method of Von Heijne ((1986) Nucleic Acids Research 14:4683-4691.) Table I, below, describes the various morphogens identified to date, including their nomenclature as used herein, their Seq. ID references, and publication sources for the amino acid sequences for the full length proteins not included in the Seq. Listing. The disclosure of these publications is incorporated herein by reference.

TABLE I

"OP-1" Refers generically to the group of morphogenically active proteins expressed from .15 part or all of a DNA sequence encoding OP-1 protein, including allelic and species variants thereof, e.g., human OP-1 ("hOP-1", Seq. ID No. 5, mature protein amino acid sequence), or mouse OP-1 ("mOP-1", Seq. ID No. 6, mature 20 protein amino acid sequence.) The conserved seven cysteine skeleton is defined by residues 38 to 139 of Seq. ID Nos. 5 and 6. The cDNA sequences and the amino acids encoding the full length proteins are provided in Seq. Id Nos. 16 25 and 17 (hOP1) and Seq. ID Nos. 18 and 19 (mOP1.) The mature proteins are defined by residues 293-431 (hOP1) and 292-430 (mOP1). The "pro" regions of the proteins, cleaved to yield the mature, morphogenically active 30 proteins are defined essentially by residues 30-292 (hOP1) and residues 30-291 (mOP1).

Refers generically to the group of active "OP-2" proteins expressed from part or all of a DNA sequence encoding OP-2 protein, including allelic and species variants thereof, e.g., human OP-2 ("hOP-2", Seq. ID No. 7, mature 5 protein amino acid sequence) or mouse OP-2 ("mOP-2", Seq. ID No. 8, mature protein amino acid sequence). The conserved seven cysteine skeleton is defined by residues 38 to 139 of Seq. ID Nos. 7 and 8. The cDNA sequences and 10 the amino acids encoding the full length proteins are provided in Seq. ID Nos. 20 and 21 (hOP2) and Seq. ID Nos. 22 and 23 (mOP2.) mature proteins are defined essentially by residues 264-402 (hOP2) and 261-399 (mOP2). 15 The "pro" regions of the proteins, cleaved to yield the mature, morphogenically active proteins are defined essentially by residues 18-263 (hOP2) and residues 18-260 (mOP1). 20 Refers generically to the morphogenically "CBMP2" active proteins expressed from a DNA sequence encoding the CBMP2 proteins, including allelic and species variants thereof, e.g., human CBMP2A ("CBMP2A(fx)", Seq ID No. 9) or human 25 CBMP2B DNA ("CBMP2B(fx)", Seq. ID No. 10). amino acid sequence for the full length proteins, referred to in the literature as BMP2A and BMP2B, or BMP2 and BMP4, appear in Wozney, et al. (1988) Science 242:1528-1534. 30 The pro domain for BMP2 (BMP2A) likely includes residues 25-248; the mature protein, residues 249-396. The pro domain for BMP4 (BMP2B) likely includes residues 25-256; the mature protein, residues 257-408. 35

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refers to protein sequences encoded by the "DPP(fx)" Drosophila DPP gene and defining the conserved seven cysteine skeleton (Seq. ID No. 11). amino acid sequence for the full length protein appears in Padgett, et al (1987) Nature 325: 5 The pro domain likely extends from the signal peptide cleavage site to residue 456; the mature protein likely is defined by residues 457-588. 10 refers to protein sequences encoded by the "Vql(fx)"

Xenopus Vgl gene and defining the conserved seven cysteine skeleton (Seq. ID No. 12). amino acid sequence for the full length protein appears in Weeks (1987) Cell 51: 861-867. prodomain likely extends from the signal peptide cleavage site to residue 246; the mature protein likely is defined by residues 247-360.

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refers to protein sequences encoded by the "Vgr-1(fx)" murine Vgr-1 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 13). amino acid sequence for the full length protein appears in Lyons, et al, (1989) PNAS 86: 4554-The prodomain likely extends from the signal peptide cleavage site to residue 299; the mature protein likely is defined by residues 300-438.

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refers to protein sequences encoded by the "GDF-1(fx)" human GDF-1 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 14). The cDNA and encoded amino sequence for the full length protein is provided in Seq. ID. No. 32.

The prodomain likely extends from the signal peptide cleavage site to residue 214; the mature protein likely is defined by residues 215-372.

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"60A"

refers generically to the morphogenically active proteins expressed from part or all of a DNA sequence (from the Drosophila 60A gene) encoding the 60A proteins (see Seq. ID No. 24 wherein the cDNA and encoded amino acid sequence for the full length protein is provided). "60A(fx)" refers to the protein sequences defining the conserved seven cysteine skeleton (residues 354 to 455 of Seq. ID No. 24.) The prodomain likely extends from the signal peptide cleavage site to residue 324; the mature protein likely is defined by residues 325-455.

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20 "BMP3(fx)"

refers to protein sequences encoded by the human BMP3 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 26). The amino acid sequence for the full length protein appears in Wozney et al. (1988) Science 242: 1528-1534. The pro domain likely extends from the signal peptide cleavage site to residue 290; the mature protein likely is defined by residues 291-472.

30 "BMP5(fx)"

refers to protein sequences encoded by the human BMP5 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 27). The amino acid sequence for the full length protein appears in Celeste, et al. (1991) PNAS 87: 9843-9847. The pro domain likely extends from

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the signal peptide cleavage site to residue 316; the mature protein likely is defined by residues 317-454.

refers to protein sequences encoded by the "BMP6(fx)" human BMP6 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 28). amino acid sequence for the full length protein appears in Celeste, et al. (1990) PNAS 87: The pro domain likely includes 9843-5847. 10 extends from the signal peptide cleavage site to residue 374; the mature sequence likely includes residues 375-513.

The OP-2 proteins have an additional cysteine residue in 15 the conserved region (e.g., see residue 41 of Seq. ID Nos. 7 and 8), in addition to the conserved cysteine skeleton in common with the other proteins in this family. The GDF-1 protein has a four amino acid insert within the conserved 20 skeleton (residues 44-47 of Seq. ID No. 14) but this insert likely does not interfere with the relationship of the cysteines in the folded structure. In addition, the CBMP2 proteins are missing one amino acid residue within the cysteine skeleton.

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The morphogens are inactive when reduced, but are active as oxidized homodimers and when oxidized in combination with Thus, as defined other morphogens of this invention. herein, a morphogen is a dimeric protein comprising a pair 30 of polypeptide chains, wherein each polypeptide chain comprises at least the C-terminal six cysteine skeleton defined by residues 43-139 of Seq. ID No. 5, including functionally equivalent arrangements of these cysteines (e.g., amino acid insertions or deletions which alter the linear arrangement of the cysteines in the sequence but not

th ir relationship in the folded structure), such that, when the polypeptide chains are folded, the dimeric protein species comprising the pair of polypeptide chains has the appropriate three-dimensional structure, including the 5 appropriate intra- and/or inter-chain disulfide bonds such that the protein is capable of acting as a morphogen as defined herein. Specifically, the morphogens generally are capable of all of the following biological functions in a morphogenically permissive environment: stimulating 10 proliferation of progenitor cells; stimulating the differentiation of progenitor cells; stimulating the proliferation of differentiated cells; and supporting the growth and maintenance of differentiated cells. addition, it is also anticipated that these morphogens are 15 capable of inducing redifferentiation of committed cells under appropriate environmental conditions.

In one preferred aspect, the morphogens of this invention comprise one of two species of generic amino acid sequences: Generic Sequence 1 (Seq. ID No. 1) or Generic Sequence 2 (Seq. ID No. 2); where each Xaa indicates one of the 20 naturally-occurring L-isomer, α-amino acids or a derivative thereof. Generic Sequence 1 comprises the conserved six cysteine skeleton and Generic Sequence 2 comprises the conserved six cysteine skeleton plus the additional cysteine identified in OP-2 (see residue 36, Seq. ID No. 2). In another preferred aspect, these sequences further comprise the following additional sequence at their N-terminus:

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Cys Xaa Xaa Xaa Xaa (Seq. ID No. 15)

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Preferred amino acid sequences within the foregoing generic sequences include: Generic Sequence 3 (Seq. ID No. 3), Generic Sequence 4 (Seq. ID No. 4), Generic Sequence 5 (Seq. ID No. 30) and Generic Sequence 6 (Seq. ID No. 31), 5 listed below. These Generic Sequences accommodate the homologies shared among the various preferred members of this morphogen family identified in Table II, as well as the amino acid sequence variation among them. Generic Sequences 3 and 4 are composite amino acid sequences of the proteins 10 presented in Table II and identified in Seq. ID Nos. 5-14, specifically: human OP-1 (hOP-1, Seq. ID Nos. 5 and 16-17), mouse OP-1 (mOP-1, Seq. ID Nos. 6 and 18-19), human and mouse OP-2 (Seq. ID Nos. 7, 8, and 20-22), CBMP2A (Seq. ID No. 9), CBMP2B (Seq. ID No. 10), DPP (from Drosophila, Seq. ID No. 11), Vgl, (from Xenopus, Seq. ID No. 12), Vgr-1 (from mouse, Seq. ID No. 13), and GDF-1 (from mouse, Seq. ID No. 14.) The generic sequences include both the amino acid identity shared by the sequences in Table II, as well as alternative residues for the variable positions within the 20 sequence. Note that these generic sequences allow for an additional cysteine at position 41 or 46 in Generic Sequences 3 or 4, respectively, providing an appropriate cysteine skeleton where inter- or intramolecular disulfide bonds can form, and contain certain critical amino acids 25 which influence the tertiary structure of the proteins.

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Generic Sequence 3

Leu Tyr Val Xaa Phe

Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa

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Xaa Ala Pro Xaa Gly Xaa Xaa Ala

15 20

Xaa Tyr Cys Xaa Gly Xaa Cys Xaa

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10 Xaa Pro Xaa Xaa Xaa Xaa

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Xaa Xaa Xaa Asn His Ala Xaa Xaa

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Xaa Xaa Leu Xaa Xaa Xaa Xaa

15 50

Xaa Xaa Xaa Xaa Xaa Xaa Cys

55 60

Cys Xaa Pro Xaa Xaa Xaa Xaa

65

20 Xaa Xaa Xaa Leu Xaa Xaa Xaa

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Xaa Xaa Xaa Xaa Val Xaa Leu Xaa

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25 85 90

Xaa Cys Gly Cys Xaa

95

wherein each Xaa is independently selected from a group of one or more specified amino acids defined as follows: "Res." means "residue" and Xaa at res.4 = (Ser, Asp or Glu); Xaa at res.6 = (Arg, Gln, Ser or 5 Lys); Xaa at res.7 = (Asp or Glu); Xaa at res.8 = (Leu or Val); Xaa at res.11 = (Gln, Leu, Asp, His or Asn); Xaa at res.12 = (Asp, Arg or Asn); Xaa at res.14 = (Ile or Val); Xaa at res.15 = (Ile or Val); Xaa at res.18 = (Glu, Gln, Leu, Lys, Pro or Arg); Xaa at res.20 = (Tyr 10 or Phe); Xaa at res.21 = (Ala, Ser, Asp, Met, His, Leu or Gln); Xaa at res.23 = (Tyr, Asn or Phe); Xaa at res.26 = (Glu, His, Tyr, Asp or Gln); Xaa at res.28 = (Glu, Lys, Asp or Gln); Xaa at res.30 = (Ala, Ser, Pro or Gln); Xaa at res.31 = (Phe, Leu or Tyr); Xaa at 15 res.33 = (Leu or Val); Xaa at res.34 = (Asn, Asp, Ala or Thr); Xaa at res.35 = (Ser, Asp, Glu, Leu or Ala); Xaa at res.36 = (Tyr, Cys, His, Ser or Ile); Xaa at res.37 = (Met, Phe, Gly or Leu); Xaa at res.38 = (Asn or Ser); Xaa at res.39 = (Ala, Ser or Gly); Xaa at 20 res.40 = (Thr, Leu or Ser); Xaa at res.44 = (Ile or Val); Xaa at res.45 = (Val or Leu); Xaa at res.46 = (Gln or Arg); Xaa at res.47 = (Thr, Ala or Ser); Xaa at res.49 = (Val or Met); Xaa at res.50 = (His or Asn); Xaa at res.51 = (Phe, Leu, Asn, Ser, Ala or Val); Xaa 25 at res.52 = (Ile, Met, Asn, Ala or Val); Xaa at res.53 = (Asn, Lys, Ala or Glu); Xaa at res.54 = (Pro or Ser); Xaa at res.55 = (Glu, Asp, Asn, or Gly); Xaa at res.56 = (Thr, Ala, Val, Lys, Asp, Tyr, Ser or Ala); Xaa at res.57 = (Val, Ala or Ile); Xaa at res.58 = (Pro or 30 Asp); Xaa at res.59 = (Lys or Leu); Xaa at res.60 = (Pro or Ala); Xaa at res.63 = (Ala or Val); Xaa at res.65 = (Thr or Ala); Xaa at res.66 = (Gln, Lys, Arg or Glu); Xaa at res.67 = (Leu, Met or Val); Xaa at res.68 = (Asn, Ser or Asp); Xaa at res.69 = (Ala, Pro or Ser); Xaa at res.70 = (Ile, Thr or Val); Xaa at

res.71 = (Ser or Ala); Xaa at res.72 = (Val or Met);
Xaa at res.74 = (Tyr or Phe); Xaa at res.75 = (Phe, Tyr or Leu); Xaa at res.76 = (Asp or Asn); Xaa at res.77 = (Asp, Glu, Asn or Ser); Xaa at res.78 = (Ser, Gln, Asn or Tyr); Xaa at res.79 = (Ser, Asn, Asp or Glu); Xaa at res.80 = (Asn, Thr or Lys); Xaa at res.82 = (Ile or Val); Xaa at res.84 = (Lys or Arg); Xaa at res.85 = (Lys, Asn, Gln or His); Xaa at res.86 = (Tyr or His); Xaa at res.87 = (Arg, Gln or Glu); Xaa at res.88 = (Asn, Glu or Asp); Xaa at res.90 = (Val, Thr or Ala); Xaa at res.92 = (Arg, Lys, Val, Asp or Glu); Xaa at res.93 = (Ala, Gly or Glu); and Xaa at res.97 = (His or Arg);

15 Generic Sequence 4

Cys Xaa Xaa Xaa Xaa Leu Tyr Val Xaa Phe 10 1 Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa 15 20 Xaa Ala Pro Xaa Gly Xaa Xaa Ala 20 25 Xaa Tyr Cys Xaa Gly Xaa Cys Xaa 30 35 25 Xaa Pro Xaa Xaa Xaa Xaa Xaa Xaa Xaa Asn His Ala Xaa Xaa 50 Xaa Xaa Leu Xaa Xaa Xaa Xaa 55 30 Xaa Xaa Xaa Xaa Xaa Xaa Cys 60 65 Cys Xaa Pro Xaa Xaa Xaa Xaa 70 35 Xaa Xaa Xaa Leu Xaa Xaa Xaa 75 80

Xaa Xaa Xaa Val Xaa Leu Xaa 85

Xaa Xaa Xaa Met Xaa Val Xaa 90 95

5 Xaa Cys Gly Cys Xaa 100

wherein each Xaa is independently selected from a group of one or more specified amino acids as defined by the following: "Res." means "residue" and Xaa at res.2 = 10 (Lys or Arg); Xaa at res.3 = (Lys or Arg); Xaa at res.4 = (His or Arg); Xaa at res.5 = (Glu, Ser, His, Gly, Arg or Pro); Xaa at res.9 = (Ser, Asp or Glu); Xaa at res.11 = (Arg, Gln, Ser or Lys); Xaa at res.12 = (Asp or Glu); Xaa at res.13 = (Leu or Val); Xaa at res.16 = (Gln, Leu, Asp, His or Asn); Xaa at res.17 = (Asp, Arg, 15 or Asn); Xaa at res.19 = (Ile or Val); Xaa at res.20 = (Ile or Val); Xaa at res.23 = (Glu, Gln, Leu, Lys, Pro or Arg); Xaa at res.25 = (Tyr or Phe); Xaa at res.26 = (Ala, Ser, Asp, Met, His, Leu, or Gln); Xaa at res.28 = (Tyr, Asn or Phe); Xaa at res.31 = (Glu, His, Tyr, Asp 20 or Gln); Xaa at res.33 = Glu, Lys, Asp or Gln); Xaa at res.35 = (Ala, Ser or Pro); Xaa at res.36 = (Phe, Leu or Tyr); Xaa at res.38 = (Leu or Val); Xaa at res.39 = (Asn, Asp, Ala or Thr); Xaa at res.40 = (Ser, Asp, Glu, 25 Leu or Ala); Xaa at res.41 = (Tyr, Cys, His, Ser or Ile); Xaa at res.42 = (Met, Phe, Gly or Leu); Xaa at res.44 = (Ala, Ser or Gly); Xaa at res.45 = (Thr, Leu or Ser); Xaa at res.49 = (Ile or Val); Xaa at res.50 = (Val or Leu); Xaa at res.51 = (Gln or Arg); Xaa at 30 res.52 = (Thr, Ala or Ser); Xaa at res.54 = (Val or Met); Xaa at res.55 = (His or Asn); Xaa at res.56 = (Phe, Leu, Asn, Ser, Ala or Val); Xaa at res.57 = (Ile, Met, Asn, Ala or Val); Xaa at res.58 = (Asn, Lys, Ala or Glu); Xaa at res.59 = (Pro or Ser); Xaa at res.60 = (Glu, Asp, or Gly); Xaa at res.61 = (Thr, Ala, Val,

Lys, Asp, Tyr, Ser or Ala); Xaa at res.62 = (Val, Ala or Ile); Xaa at res.63 = (Pro or Asp); Xaa at res.64 = (Lys or Leu); Xaa at res.65 = (Pro or Ala); Xaa at res.68 = (Ala or Val); Xaa at res.70 = (Thr or Ala); 5 Xaa at res.71 = (Gln, Lys, Arg or Glu); Xaa at res.72 = (Leu, Met or Val); Xaa at res.73 = (Asn, Ser or Asp); Xaa at res.74 = (Ala, Pro or Ser); Xaa at res.75 = (Ile, Thr or Val); Xaa at res.76 = (Ser or Ala); Xaa at res.77 = (Val or Met); Xaa at res.79 = (Tyr or Phe); 10 Xaa at res.80 = (Phe, Tyr or Leu); Xaa at res.81 = (Asp or Asn); Xaa at res.82 = (Asp, Glu, Asn or Ser); Xaa at res.83 = (Ser, Gln, Asn or Tyr); Xaa at res.84 = (Ser, Asn, Asp or Glu); Xaa at res.85 = (Asn, Thr or Lys); Xaa at res.87 = (Ile or Val); Xaa at res.89 = (Lys or Arg); Xaa at res.90 = (Lys, Asn, Gln or His); Xaa at 15 res.91 = (Tyr or His); Xaa at res.92 = (Arg, Gln or Glu); Xaa at res.93 = (Asn, Glu or Asp); Xaa at res.95 = (Val, Thr or Ala); Xaa at res.97 = (Arg, Lys, Val, Asp or Glu); Xaa at res.98 = (Ala, Gly or Glu); and Xaa 20 at res. 102 = (His or Arg).

Similarly, Generic Sequence 5 (Seq. ID No. 30) and Generic Sequence 6 (Seq. ID No. 31) accommodate the homologies shared among all the morphogen protein 25 family members identified in Table II. Specifically, Generic Sequences 5 and 6 are composite amino acid sequences of human OP-1 (hOP-1, Seq. ID Nos. 5 and 16-17), mouse OP-1 (mOP-1, Seq. ID Nos. 6 and 18-19), human and mouse OP-2 (Seq. ID Nos. 7, 8, and 20-22), CBMP2A (Seq. ID No. 9), CBMP2B (Seq. ID No. 10), DPP (from Drosophila, Seq. ID No. 11), Vgl, (from Xenopus, Seq. ID No. 12), Vgr-1 (from mouse, Seq. ID No. 13), and GDF-1 (from mouse, Seq. ID No. 14), human BMP3 (Seq. ID No. 26), human BMP5 (Seq. ID No. 27), human BMP6 (Seq. ID No. 28) and 60(A) (from Drosophila, Seq. 35

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ID Nos. 24-25). The generic sequences include both the amino acid identity shared by these sequences in the C-terminal domain, defined by the six and seven cysteine skeletons (Generic Sequences 5 and 6,

5 respectively), as well as alternative residues for the variable positions within the sequence. As for Generic Sequences 3 and 4, Generic Sequences 5 and 6 allow for an additional cysteine at position 41 (Generic Sequence 5) or position 46 (Generic Sequence 6), providing an appropriate cysteine skeleton where inter- or intramolecular disulfide bonds can form, and containing certain critical amino acids which influence the tertiary structure of the proteins.

15 <u>Generic Sequence 5</u>

Leu Xaa Xaa Xaa Phe

Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa

5

20 10

1

Xaa Xaa Pro Xaa Xaa Xaa Ala

15 20

Xaa Tyr Cys Xaa Gly Xaa Cys Xaa

25 30

25 Xaa Pro Xaa Xaa Xaa Xaa

35

Xaa Xaa Xaa Asn His Ala Xaa Xaa

40 45

Xaa Xaa Xaa Xaa Xaa Xaa Xaa

30 50

Xaa Xaa Xaa Xaa Xaa Xaa Cys

55 60

Cys Xaa Pro Xaa Xaa Xaa Xaa Xaa

65

5 Xaa Xaa Xaa Leu Xaa Xaa Xaa

70 75

Xaa Xaa Xaa Xaa Val Xaa Leu Xaa

80

Xaa Xaa Xaa Xaa Met Xaa Val Xaa

10 85 90

Xaa Cys Xaa Cys Xaa

95

wherein each Xaa is independently selected from a group of one or more specified amino acids defined as 15 follows: "Res." means "residue" and Xaa at res.2 = (Tyr or Lys); Xaa at res.3 = Val or Ile); Xaa at res.4 = (Ser, Asp or Glu); Xaa at res.6 = (Arg, Gln, Ser, Lys or Ala); Xaa at res.7 = (Asp, Glu or Lys); Xaa at res.8 = (Leu, Val or Ile); Xaa at res.11 = (Gln, Leu, Asp, 20 His, Asn or Ser); Xaa at res.12 = (Asp, Arg, Asn or Glu); Xaa at res.14 = (Ile or Val); Xaa at res.15 = (Ile or Val); Xaa at res.16 (Ala or Ser); Xaa at res.18 = (Glu, Gln, Leu, Lys, Pro or Arg); Xaa at res.19 = (Gly or Ser); Xaa at res.20 = (Tyr or Phe); Xaa at 25 res.21 = (Ala, Ser, Asp, Met, His, Gln, Leu or Gly); Xaa at res.23 = (Tyr, Asn or Phe); Xaa at res.26 = (Glu, His, Tyr, Asp, Gln or Ser); Xaa at res.28 = (Glu, Lys, Asp, Gln or Ala); Xaa at res.30 = (Ala, Ser, Pro, Gln or Asn); Xaa at res.31 = (Phe, Leu or Tyr); Xaa at 30 res.33 = (Leu, Val or Met); Xaa at res.34 = (Asn, Asp,

Ala, Thr or Pro); Xaa at res.35 = (Ser, Asp, Glu, Leu,

Ala or Lys); Xaa at res.36 = (Tyr, Cys, His, Ser or Ile); Xaa at res.37 = (Met, Phe, Gly or Leu); Xaa at res.38 = (Asn, Ser or Lys); Xaa at res.39 = (Ala, Ser, Gly or Pro); Xaa at res.40 = (Thr, Leu or Ser); Xaa at 5 res.44 = (Ile, Val or Thr); Xaa at res.45 = (Val, Leu or Ile); Xaa at res.46 = (Gln or Arg); Xaa at res.47 = (Thr, Ala or Ser); Xaa at res.48 = (Leu or Ile); Xaa at res.49 = (Val or Met); Xaa at res.50 = (His, Asn or Arg); Xaa at res.51 = (Phe, Leu, Asn, Ser, Ala or Val); Xaa at res.52 = (Ile, Met, Asn, Ala, Val or Leu); Xaa at res.53 = (Asn, Lys, Ala, Glu, Gly or Phe); Xaa at res.54 = (Pro, Ser or Val); Xaa at res.55 = (Glu, Asp, Asn, Gly, Val or Lys); Xaa at res.56 = (Thr, Ala, Val, Lys, Asp, Tyr, Ser, Ala, Pro or His); Xaa at res.57 = (Val, Ala or Ile); Xaa at res.58 = (Pro or Asp); Xaa at 15 res.59 = (Lys, Leu or Glu); Xaa at res.60 = (Pro or Ala); Xaa at res.63 = (Ala or Val); Xaa at res.65 = (Thr, Ala or Glu); Xaa at res.66 = (Gln, Lys, Arg or Glu); Xaa at res.67 = (Leu, Met or Val); Xaa at res.68 20 = (Asn, Ser, Asp or Gly); Xaa at res.69 = (Ala, Pro or Ser); Xaa at res.70 = (Ile, Thr, Val or Leu); Xaa at res.71 = (Ser, Ala or Pro); Xaa at res.72 = (Val, Met or Ile); Xaa at res.74 = (Tyr or Phe); Xaa at res.75 = (Phe, Tyr, Leu or His); Xaa at res.76 = (Asp, Asn or 25 Leu); Xaa at res.77 = (Asp, Glu, Asn or Ser); Xaa at res.78 = (Ser, Gln, Asn, Tyr or Asp); Xaa at res.79 = (Ser, Asn, Asp, Glu or Lys); Xaa at res.80 = (Asn, Thr or Lys); Xaa at res.82 = (Ile, Val or Asn); Xaa at res.84 = (Lys or Arg); Xaa at res.85 = (Lys, Asn, Gln, 30 His or Val); Xaa at res.86 = (Tyr or His); Xaa at res.87 = (Arg, Gln, Glu or Pro); Xaa at res.88 = (Asn, Glu or Asp); Xaa at res.90 = (Val, Thr, Ala or Ile); Xaa at res.92 = (Arg, Lys, Val, Asp or Glu); Xaa at res.93 = (Ala, Gly, Glu or Ser); Xaa at res.95 = (Gly 35 or Ala) and Xaa at res.97 = (His or Arg).

Generic Sequence 6

Cys Xaa Xaa Xaa Leu Xaa Xaa Phe 1 5 10 5 Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa 15 Xaa Xaa Pro Xaa Xaa Xaa Ala 25 20 Xaa Tyr Cys Xaa Gly Xaa Cys Xaa 10 30 Xaa Pro Xaa Xaa Xaa Xaa 40 Xaa Xaa Xaa Asn His Ala Xaa Xaa 15 Xaa Xaa Xaa Xaa Xaa Xaa Xaa 55 Xaa Xaa Xaa Xaa Xaa Xaa Cys Cys Xaa Pro Xaa Xaa Xaa Xaa 70 20 Xaa Xaa Xaa Leu Xaa Xaa Xaa 75 Xaa Xaa Xaa Xaa Val Xaa Leu Xaa 85 25 Xaa Xaa Xaa Met Xaa Val Xaa 90 Xaa Cys Xaa Cys Xaa 100

wherein each Xaa is independently selected from a group of one or more specified amino acids as defined by the following: "Res." means "residue" and Xaa at res.2 = (Lys, Arg, Ala or Gln); Xaa at res.3 = (Lys, Arg or Met); Xaa at res.4 = (His, Arg or Gln); Xaa at res.5 = (Glu, Ser, His, Gly, Arg, Pro, Thr, or Tyr); Xaa at

res.7 = (Tyr or Lys); Xaa at res.8 = (Val or Ile); Xaa at res.9 = (Ser, Asp or Glu); Xaa at res.11 = (Arg, Gln, Ser, Lys or Ala); Xaa at res.12 = (Asp, Glu, or Lys); Xaa at res.13 = (Leu, Val or Ile); Xaa at res.16 5 = (Gln, Leu, Asp, His, Asn or Ser); Xaa at res.17 = (Asp, Arg, Asn or Glu); Xaa at res.19 = (Ile or Val); Xaa at res.20 = (Ile or Val); Xaa at res.21 = (Ala or Ser); Xaa at res.23 = (Glu, Gln, Leu, Lys, Pro or Arg); Xaa at res.24 = (Gly or Ser); Xaa at res.25 = (Tyr or 10 Phe); Xaa at res.26 = (Ala, Ser, Asp, Met, His, Gln, Leu, or Gly); Xaa at res.28 = (Tyr, Asn or Phe); Xaa at res.31 = (Glu, His, Tyr, Asp, Gln or Ser); Xaa at res.33 = Glu, Lys, Asp, Gln or Ala); Xaa at res.35 = (Ala, Ser, Pro, Gln or Asn); Xaa at res.36 = (Phe, Leu 15 or Tyr); Xaa at res.38 = (Leu, Val or Met); Xaa at res.39 = (Asn, Asp, Ala, Thr or Pro); Xaa at res.40 = (Ser, Asp, Glu, Leu, Ala or Lys); Xaa at res.41 = (Tyr, Cys, His, Ser or Ile); Xaa at res.42 = (Met, Phe, Gly or Leu); Xaa at res.43 = (Asn, Ser or Lys); Xaa at 20 res.44 = (Ala, Ser, Gly or Pro); Xaa at res.45 = (Thr, Leu or Ser); Xaa at res.49 = (Ile, Val or Thr); Xaa at res.50 = (Val, Leu or Ile); Xaa at res.51 = (Gln or Arg); Xaa at res.52 = (Thr, Ala or Ser); Xaa at res.53 = (Leu or Ile); Xaa at res.54 = (Val or Met); Xaa at 25 res.55 = (His, Asn or Arg); Xaa at res.56 = (Phe, Leu, Asn, Ser, Ala or Val); Xaa at res.57 = (Ile, Met, Asn, Ala, Val or Leu); Xaa at res.58 = (Asn, Lys, Ala, Glu, Gly or Phe); Xaa at res.59 = (Pro, Ser or Val); Xaa at res.60 = (Glu, Asp, Gly, Val or Lys); Xaa at res.61 = 30 (Thr, Ala, Val, Lys, Asp, Tyr, Ser, Ala, Pro or His); Xaa at res.62 = (Val, Ala or Ile); Xaa at res.63 = (Pro or Asp); Xaa at res.64 = (Lys, Leu or Glu); Xaa at res.65 = (Pro or Ala); Xaa at res.68 = (Ala or Val); Xaa at res.70 = (Thr, Ala or Glu); Xaa at res.71 = 35 (Gln, Lys, Arg or Glu); Xaa at res.72 = (Leu, Met or

Val); Xaa at res.73 = (Asn, Ser, Asp or Gly); Xaa at res.74 = (Ala, Pro or Ser); Xaa at res.75 = (Ile, Thr, Val or Leu); Xaa at res.76 = (Ser, Ala or Pro); Xaa at res.77 = (Val, Met or Ile); Xaa at res.79 = (Tyr or 5 Phe); Xaa at res.80 = (Phe, Tyr, Leu or His); Xaa at res.81 = (Asp, Asn or Leu); Xaa at res.82 = (Asp, Glu, Asn or Ser); Xaa at res.83 = (Ser, Gln, Asn, Tyr or Asp); Xaa at res.84 = (Ser, Asn, Asp, Glu or Lys); Xaa at res.85 = (Asn, Thr or Lys); Xaa at res.87 = (Ile, 10 Val or Asn); Xaa at res.89 = (Lys or Arg); Xaa at res.90 = (Lys, Asn, Gln, His or Val); Xaa at res.91 = (Tyr or His); Xaa at res.92 = (Arg, Gln, Glu or Pro); Xaa at res.93 = (Asn, Glu or Asp); Xaa at res.95 = (Val, Thr, Ala or Ile); Xaa at res.97 = (Arg, Lys, Val, 15 Asp or Glu); Xaa at res.98 = (Ala, Gly, Glu or Ser); Xaa at res.100 = (Gly or Ala); and Xaa at res.102 = (His or Arg).

Particularly useful sequences for use as morphogens in 20 this invention include the C-terminal domains, e.g., the Cterminal 96-102 amino acid residues of Vgl, Vgr-1, DPP, OP-1, OP-2, CBMP-2A, CBMP-2B, GDF-1 (see Table II, below, and Seq. ID Nos. 5-14), as well as proteins comprising the C-terminal domains of 60A, BMP3, BMP5 and BMP6 (see Seq. ID 25 Nos. 24-28), all of which include at least the conserved six or seven cysteine skeleton. In addition, biosynthetic constructs designed from the generic sequences, such as COP-1, 3-5, 7, 16, disclosed in U.S. Pat. No. 5,011,691, also are useful. Other sequences include the 30 inhibins/activin proteins (see, for example, U.S. Pat. Nos. 4,968,590 and 5,011,691). Accordingly, other useful sequences are those sharing at least 70% amino acid sequence homology or "similarity", and preferably 80% homology or similarity with any of the sequences above. 35 anticipated to include allelic, species variants and other

sequence variants (e.g., including "muteins" or "mutant proteins"), whether naturally-occurring or biosynthetically produced, as well as novel members of this morphogenic family of proteins. As used herein, "amino acid sequence 5 homology" is understood to mean amino acid sequence similarity, and homologous sequences share identical or similar amino acids, where similar amino acids are conserved amino acids as defined by Dayoff et al., Atlas of Protein Sequence and Structure; vol.5, Suppl.3, pp.345-362 (M.O. 10 Dayoff, ed., Nat'l BioMed. Research Fdn., Washington D.C. Thus, a candidate sequence sharing 70% amino acid homology with a reference sequence requires that, following alignment of the candidate sequence with the reference sequence, 70% of the amino acids in the candidate sequence 15 are identical to the corresponding amino acid in the reference sequence, or constitute a conserved amino acid change thereto. "Amino acid sequence identity" is understood to require identical amino acids between two aligned sequences. Thus, a candidate sequence sharing 60% 20 amino acid identity with a reference sequence requires that, following alignment of the candidate sequence with the reference sequence, 60% of the amino acids in the candidate sequence are identical to the corresponding amino acid in the reference sequence.

25

As used herein, all homologies and identities calculated use OP-1 as the reference sequence. Also as used herein, sequences are aligned for homology and identity calculations using the method of Needleman et al. (1970) J.Mol. Biol.

30 48:443-453 and identities calculated by the Align program (DNAstar, Inc.) In all cases, internal gaps and amino acid insertions in the candidate sequence as aligned are ignored when making the homology/identity calculation.

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The currently most preferred protein sequences useful as morphogens in this invention include those having greater than 60% identity, preferably greater than 65% identity, with the amino acid sequence defining the conserved six 5 cysteine skeleton of hOP1 (e.g., residues 43-139 of Seq. ID No. 5). These most preferred sequences include both allelic and species variants of the OP-1 and OP-2 proteins, including the Drosophila 60A protein. Accordingly, in another preferred aspect of the invention, useful morphogens 10 include active proteins comprising species of polypeptide chains having the generic amino acid sequence herein referred to as "OPX" (Seq. ID No. 29), which defines the seven cysteine skeleton and accommodates the homologies between the various identified species of OP1 and OP2. As 15 described therein, each Xaa at a given position independently is selected from the residues occurring at the corresponding position in the C-terminal sequence of mouse or human OP1 or OP2 (see Seq. ID Nos. 5-8 and/or Seq. ID Nos. 16-23).

20

In still another preferred aspect of the invention, useful morphogens include dimeric proteins comprising amino acid sequences encoded by nucleic acids that hybridize to DNA or RNA sequences encoding the C-terminal sequences defining the conserved seven cysteine domain of OP1 or OP2, e.g., nucleotides 1036-1341 and nucleotides 1390-1695 of Seq. ID No. 16 and 20, respectively, under stringent hybridization conditions. As used herein, stringent hybridization conditions are defined as hybridization in 40% formamide, 5 X SSPE, 5 X Denhardt's Solution, and 0.1% SDS at 37°C overnight, and washing in 0.1 X SSPE, 0.1% SDS at 50°C.

The morphogens useful in the methods, compositions and devices of this invention include proteins comprising any of

the polypeptide chains described above, whether isolated from naturally-occurring sources, or produced by recombinant DNA or other synthetic techniques, and includes allelic and species variants of these proteins, naturally-occurring or 5 biosynthetic mutants thereof, as well as various truncated and fusion constructs. Deletion or addition mutants also are envisioned to be active, including those which may alter the conserved C-terminal cysteine skeleton, provided that the alteration does not functionally disrupt the 10 relationship of these cysteines in the folded structure. Accordingly, such active forms are considered the equivalent of the specifically described constructs disclosed herein. The proteins may include forms having varying glycosylation patterns, varying N-termini, a family of related proteins 15 having regions of amino acid sequence homology, and active truncated or mutated forms of native or biosynthetic proteins, produced by expression of recombinant DNA in host cells.

The morphogenic proteins can be expressed from intact or truncated cDNA or from synthetic DNAs in procaryotic or eucaryotic host cells, and purified, cleaved, refolded, and dimerized to form morphogenically active compositions. Currently preferred host cells include E. coli or mammalian cells, such as CHO, COS or BSC cells. A detailed description of the morphogens useful in the methods, compositions and devices of this invention is disclosed in copending US patent application Serial Nos. 752,764, filed August 30, 1991, and 667,724, filed March 11, 1991, the disclosures of which are incorporated herein by reference.

Thus, in view of this disclosure, skilled genetic engineers can isolate genes from cDNA or genomic libraries of various different species which encode appropriate amino acid sequences, or construct DNAs from oligonucleotides, and

then can express them in various types of host cells, including both procaryotes and eucaryotes, to produce large quantities of active proteins capable of stimulating the morphogenesis of, and/or inhibiting damage to, periodontal tissue.

Other features and advantages of the invention will be apparent from the following description of the preferred embodiments, and from the claims.

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Brief Description of the Drawings

The foregoing and other objects and features of this invention, as well as the invention itself, may be more fully understood from the following description, when read together with the accompanying drawings, in which:

Fig. 1 is a schematic illustration of a healthy tooth in the tooth socket; and

10

Fig. 2 (A and B) are photomicrographs demonstrating the effect of morphogen (2A) or carrier alone (2B) on periodontal tissue regeneration in a surgically prepared canine tooth socket.

15

Fig.3 (A and B) are photomicrographs demonstrating the effect of morphogen (3A) or carrier alone (3B) on dentine tissue regeneration in a surgically exposed dental pulp experiment.

Detailed Description

It has been discovered that the morphogens described herein can stimulate periodontal tissue formation, including regenerating lost or damaged periodontal ligament and/or cementum. The invention may be used for tooth implant integration as well as to inhibit and/or repair periodontal tissue loss due to disease or mechanical injury. The invention is practiced using a morphogen or morphogen—
10 stimulating agent, as defined herein, and according to the procedures described herein.

Provided below is a description of tooth anatomy and useful morphogens, including methods for their production and formulation, as well as exemplary, non-limiting examples which (1) demonstrate the suitability of the morphogens described herein in the methods of the invention, and (2) provide assays with which to test candidate morphogens for their efficacy.

20

I. Tooth Anatomy

A vertical section of a tooth in the tooth socket is shown schematically in Fig. 1. The crown 6 of the tooth is composed of enamel 8 and dentin 22. The pulp chamber 12 is seen in the interior of the crown 6 and the center of the root 10; it extends downward into the bony area 14, 16, 18 and opens by a minute orifice, the apical foramen 20, at the extremity of the root 10. The pulp chamber 12 contains dental pulp, a loose connective tissue richly supplied with vessels and nerves, which enter the cavity through the apical foramen 20. Some of the cells of the pulp, i.e., odontoblasts, the precursors of dentin 22, are arranged as a layer on the wall of the pulp chamber 12. During development of the tooth, odontoblasts are columnar, but

later, after the dentin 22 is fully formed, they become flattened and resemble osteoblasts.

The solid portion of the mature tooth includes dentin 5 22, enamel 8, and a thin layer of cementum 24, which is disposed on the surface of the root 25. Enamel 8 is formed during development of the tooth from amyloblasts, and cementum 24 is formed from cementoblasts. In a fully developed tooth, the principal mass of the tooth comprises dentin 22, which is made up of hydroxyapatite crystals embedded in a strong meshwork of collagen fibers. dentin includes a number of minute wavy and branching tubes called dental canaliculi, embedded in a dense homogeneous substance, the matrix. The dental canaliculi are parallel 15 with one another and open at their inner ends into the pulp chamber 12. The dentin matrix is translucent and comprises the majority of the inorganic mass of the dentin. includes a number of fine fibrils, which are continuous with the fibrils of the dental pulp. After the organic matter 20 has been removed by steeping a tooth in weak acid, the remaining organic matter may be torn into laminae that run parallel with the pulp chamber 12 across the direction of the tubes.

The cementum 24 is disposed as a thin mineralized layer covering the tooth root. It extends from where the enamel terminates to the apex of each root, where it is usually very thick. Cementum resembles bone in structure and chemical composition in that it contains, sparingly, the lacunae and canaliculi that characterize true bone; in the thicker portions of the cementum, the lamellae and Haversian canals peculiar to bone are also found. As a result of aging, the cementum increases in thickness and the pulp chamber also becomes partially filled with a hard substance that is intermediate in structure between dentin and bone.

It appears to be formed by a slow conversion of the dental pulp, which shrinks or even disappears.

The periodontal ligament, or periodontal membrane 26, is
the layer of periodontal tissue which forms a cushion
between the cementum 24 and the bone 14, 16, 18; it holds
the tooth in position by suspending it in the socket
(alveolus) of the jawbone. The periodontal ligament is a
highly organized tissue which is formed from periodontal
fibroblasts. It organizes the collagen fibers which pass
directly from the bone of the jaw into the cementum.

II. Useful Morphogens

As defined herein a protein is morphogenic if it is 15 capable of inducing the developmental cascade of cellular and molecular events that culminate in the formation of new, organ-specific tissue and comprises at least the conserved C-terminal six cysteine skeleton or its functional equivalent (see supra). Specifically, the morphogens generally are capable of all of the following biological functions in a morphogenically permissive environment: stimulating proliferation of progenitor cells; stimulating the differentiation of progenitor cells; stimulating the 25 proliferation of differentiated cells; and supporting the growth and maintenance of differentiated cells. how the morphogens useful in the method of this invention first were identified, as well as a description on how to make, use and test them for morphogenic activity are 30 disclosed in international application (US92/01968 (WO92/15323), the disclosure of which is incorporated hereinabove by reference. As disclosed therein, the morphogens may be purified from naturally-sourced material or recombinantly produced from procaryotic or eucaryotic 35 host cells, using the genetic sequences disclosed therein.

Alternatively, novel morphogenic sequences may be identified following the procedures disclosed therein.

Particularly useful proteins include those which

5 comprise the naturally derived sequences disclosed in Table

II. Other useful sequences include biosynthetic constructs

such as those disclosed in U.S. Pat. 5,011,691, the

disclosure of which is incorporated herein by reference

(e.g., COP-1, COP-3, COP-4, COP-5, COP-7, and COP-16).

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Accordingly, the morphogens useful in the methods and compositions of this invention also may be described by morphogenically active proteins having amino acid sequences sharing 70% or, preferably, 80% homology (similarity) with any of the sequences described above, where "homology" is as defined herein above.

The morphogens useful in the method of this invention also can be described by any of the 6 generic sequences 20 described herein (Generic Sequences 1, 2, 3, 4, 5 and 6). Generic sequences 1 and 2 also may include, at their N-terminus, the sequence

Cys Xaa Xaa Xaa Xaa (Seq. ID No. 15)

25

Table II, set forth below, compares the amino acid sequences of the active regions of native proteins that have been identified as morphogens, including human OP-1 (hOP-1, Seq. ID Nos. 5 and 16-17), mouse OP-1 (mOP-1, Seq. ID Nos. 6 and 18-19), human and mouse OP-2 (Seq. ID Nos. 7, 8, and 20-23), CBMP2A (Seq. ID No. 9), CBMP2B (Seq. ID No. 10), BMP3 (Seq. ID No. 26), DPP (from Drosophila, Seq. ID No. 11), Vgl, (from Xenopus, Seq. ID No. 12), Vgr-1 (from mouse, Seq. 35 ID No. 13), GDF-1 (from mouse, Seq. ID Nos. 14, 32 and 33),

60A protein (from Drosophila, Seq. ID Nos. 24 and 25), BMP5 (Seq. ID No. 27) and BMP6 (Seq. ID No. 28). The sequences are aligned essentially following the method of Needleman et al. (1970) J. Mol. Biol., 48:443-453, calculated using the Align Program (DNAstar, Inc.) In the table, three dots indicates that the amino acid in that position is the same as the amino acid in hOP-1. Three dashes indicates that no amino acid is present in that position, and are included for purposes of illustrating homologies. For example, amino acid residue 60 of CBMP-2A and CBMP-2B is "missing". Of course, both these amino acid sequences in this region comprise Asn-Ser (residues 58, 59), with CBMP-2A then comprising Lys and Ile, whereas CBMP-2B comprises Ser and Ile.

15

TABLE II

	hOP-1	Cys	Lys	Lys	His	Glu	Leu	Tyr	Val
20	mOP-1	•••	• • •	• • •	• • •	• • •	•••	•••	• • •
	hOP-2		Arg	Arg	• • •	•••	• • •	•••	• • •
	mOP-2	•••	Arg	Arg	•••	• • •	•••	•••	• • •
	DPP	• • •	Arg	Arg	• • •	Ser	• • •	• • •	• • •
	Vgl	•••	• • •	Lys	Arg	His	•••	• • •	• • •
25	Vgr-1	• • •	•••	• • •	• • •	Gly	•••	• • •	• • •
	CBMP-2A	• • •	• • •	Arg	•••	Pro	•••	• • •	• • •
	CBMP-2B	• • •	Arg	Arg	• • •	Ser	•••	• • •	• • •
	BMP3	• • •	Ala	Arg	Arg	Tyr	•••	Lys	
	GDF-1	• • •	Arg	Ala	Arg	Arg	•••	• • •	• • •
30	60A	• • •	Gln	Met	Glu	Thr	•••	• • •	• • •
	BMP5	• • •	•••	• • •	• • •	• • •	• • •	• • •	• • •
	BMP6	• • •	Arg	• • •	• • •	• • •	•••	• • •	• • •
		1				5			

	hOP-1	Ser	Phe	Arg	Asp	Leu	Gly	Trp	Gln	Asp
	mOP-1	•••	• • •	•••	• • •	•••	•••	•••	• • •	
	hOP-2		• • •	Gln	•••	• • •	• • •	• • •	Leu	• • •
	mOP-2	Ser	• • •	•••	• • •	•••	• • •	•••	Leu	• • •
5	DPP	Asp	• • •	Ser	• • •	Val	• • •	•••	Asp	• • •
	Vgl	Glu	• • •	Lys	• • •	Val	• • •	•••	•••	Asn
	Vgr-1	•••	• • •	Gln	• • •	Val	• • •	•••	• • •	• • •
	CBMP-2A	Asp	• • •	Ser	•••	Val	• • •	•••	Asn	•••
	CBMP-2B	Asp	• • •	Ser	• • •	Val	• • •	•••	Asn	• • •
10	вир3	Asp	• • •	Ala	•••	Ile	• • •	•••	Ser	Glu
	GDF-1	• • •	• • •	•••	Glu	Val	• • •	• • •	His	Arg
	60A	Asp	• • •	Lys	• • •	•••	• • •	• • •	His	• • •
	BMP5	• • •	• • •	•••	• • •	• • •	• • •	•••	• • •	• • •
	BMP6	•••	• • •	Gln	• • •	• • •	• • •	•••	•••	•••
15			10					15		
	hOP-1	Trp	Ile	Ile	Ala	Pro	Glu	Gly	Tyr	Ala
	mOP-1	• • •	•••	•••	•••	•••	•••	•••	• • •	•••
	hOP-2	• • •	Val	•••	•••	•••	Gln	•••	•••	Ser
20	mOP-2	• • •	Val	•••		•••	Gln	• • •	• • •	Ser
	DPP	•••	•••	Val	• • •	• • •	Leu	•••	•••	Asp
	Vgl	•••	Val	•••	• • •		Gln	•••	•••	Met
	Vgr-1	•••	• • •	•••	•••		Lys	• • •		•••
	CBMP-2A	•••	• • •	Val	• • •	• • •	Pro	•••	•••	His
25	CBMP-2B	•••	• • •	Val	•••	• • •	Pro	•••	• • •	Gln
	вир3	•••	• • •	•••	Ser	• • •	Lys	Ser	Phe	Asp
	GDF-1	•••	Val	•••	•••	• • •	Arg	•••	Phe	Leu
	60A	• • •	• • •	•••	•••	• • •	• • •	•••	• • •	Gly
	BMP5	• • •	• • •	• • •	• • •	• • •	• • •	•••	•••	•••
30	вир6	•••	• • •	• • •	•••	•••	Lys	•••	• • •	•••
				20					25	

	hOP-1	Ala	Tyr	Tyr	Cys	Glu	Gly	Glu	Cys	Ala
	mOP-1	• • •	• • •	•••	•••	•••	• • •	•••	• • •	• • •
	hOP-2	• • •	• • •	•••	• • •	•••	• • •	• • •	•••	Ser
	mOP-2	• • •	• • •	•••	•••	• • •	•••	•••	• • •	• • •
5	DPP	•••	• • •	•••	• • •	His	• • •	Lys	• • •	Pro
	Vgl	•••	Asn	•••	• • •	Tyr	•••	• • •	• • •	Pro
	Vgr-1	•••	Asn	• • •	•••	Asp	• • •	• • •	• • •	Ser
	CBHP-2A	•••	Phe	• • •	• • •	His	• • •	Glu	• • •	Pro
	CBMP-2B	•••	Phe	• • •		His	• • •	Asp	•••	Pro
10	BMP3	•••	•••		•••	Ser	•••	Ala	•••	Gln
	GDF-1	•••	Asn		•••	Gln	• • •	Gln	•••	• • •
	60A	•••	Phe	•••	•••	Ser	•••		• • •	Asn
	BMP5	•••	Phe	•••	•••	Asp		• • •	• • •	Ser
	BMP6	•••	Asn	• • •	• • •	Asp		• • •	• • •	Ser
15					30	•				35
	h0P-1	Phe	Pro	Leu	Asn	Ser	Tyr	Het	Asn	Ala
	mOP-1		• • •	•••		•••	•••	•••	• • •	• • •
	h0P-2	•••		• • •	Asp	•••	Cys	• • •		• • •
20	mOP-2	•••		• • •	Asp	•••	Cys	• • •	• • •	•••
	DPP	•••	• • •	• • •	Ala	Asp	His	Phe	•••	Ser
	Vgl	Tyr	• • •	• • •	Thr	Glu	Ile	Leu	•••	Gly
	Vgr-1	-,-	• • •	• • •	•••	Ala	His	•••	•••	•••
	CBHP-2A	•••		• • •	Ala	Asp	His	Leu	•••	Ser
25	CBMP-2B	•••	•••	•••	Ala	Asp	His	Leu	• • •	Ser
	GDF-1	Leu	• • •	Val	Ala	Leu	Ser	Gly	Ser**	• • •
	вирз	•••		Het	Pro	Lys	Ser	Leu	Lys	Pro
	60A	•••	• • •		•••	Ala	His	•••	•••	
	BMP5	•••	• • •	•••	•••	Ala	His	Het	•••	•••
30	BMP6	•••	• • •	•••	•••	Ala	His	Het	•••	•••
,,,		•••				40		-		

	h0P-1	Thr	Asn	His	Ala	Ile	Val	Gln	Thr	Leu
	mOP-1	•••	• • •	•••	• • •	• • •	• • •	•••	•••	•••
	hOP-2	•••	• • •	• • •	• • •	• • •	Leu	•••	Ser	• • •
	mOP-2	•••	• • •	••••	• • •	• • •	Leu		Ser	• • •
5	DPP	•••	• • •	•••	• • •	Val	• • •	•••	•••	• • •
	Vgl	Ser		• • •	• • •	• • •	Leu	•••	• • •	• • •
	Vgr-1	•••	•••	• • •	• • •	• • •	•••	•••	• • •	• • •
	CBHP-2A	• • •	• • •	• • •	• • •	• • •	• • •	•••	•••	• • •
	CBMP-2B	•••	•••	• • •	• • •	•••		•••		• • •
10	BMP3	Ser	•••		•••	Thr	Ile	• • •	Ser	Ile
	GDF-1	Leu		• • •	• • •	Val	Leu	Arg	Ala	•••
	60A	•••	• • •		• • •	• • •		•••	• • •	•••
	BMP5	•••	•••	• • •	•••	• • •		•••	•••	•••
	BMP6	•••			•••		•••	•••	• • •	• • •
15		45					50			
	hOP-1	Val	His	Phe	Ile	Asn	Pro	Glu	Thr	Val
	mOP-1	•••	•••	•••	• • •	• • •	•••	Asp	• • •	•••
20	hOP-2	•••	His	Leu	Met	Lys	•••	Asn	Ala	• • •
	mOP-2	•••	His	Leu	Met	Lys	•••	Asp	Val	•••
•	DPP	• • •	Asn	Asn	Asn	•••	• • •	Gly	Lys	• • •
	Vgl	• • •	• • •	Ser	• • •	Glu	•••	• • •	Asp	Ile
	Vgr-1	• • •	•••	Val	Het	•••	• • •	•••	Tyr	•••
25	CBHP-2A	•••	Asn	Ser	Val	•••	Ser		Lys	Ile
	CBMP-2B	•••	Asn	Ser	Val	• • •	Ser		Ser	Ile
	BMP3	•••	Arg	Ala**	Gly	Val	Val	Pro	Gly	Ile
	GDF-1	Met	•••	Ala	Ala	Ala	• • •	Gly	Ala	Ala
	60A	•••	•••	Leu	Leu	Glu		Lys	Lys	• • •
30	BMP5	•••	•••	Leu	Met	Phe	•••	Asp	His	• • •
	BMP6	• • •	•••	Leu	Het	• • •	• • •	•••	Tyr	• • •
			55					60	•	

	h0P-1	Pro	Lys	Pro	Cys	Cys	Ala	Pro	Thr	Gln
	mOP-1	•••	• • •	•••	•••	•••	• • •	• • •	• • •	• • •
	hOP-2	•••	• • •	Ala	• • •	• • •	• • •	• • •	• • •	Lys
	mOP-2	•••	• • •	Ala	• • •	• • •	• • •	• • •	•••	Lys
5	DPP	•••		Ala	• • •	• • •	Val	•••	•••	• • •
	Vgl	•••	Leu	• • •	• • •	• • •	Val	•••	•••	Lys
	Vgr-1	• • •		• • •			• • •	• • •	• • •	Lys
	CBMP-2A	• • •	•••	Ala	•••	• • •	Val	• • •	•••	Glu
	CBMP-2B	•••	• • •	Ala	• • •	• • •	Val	• • •	• • •	Glu
10	BMP3	• • •	Glu	•••	•••	• • •	Val	• • •	Glu	Lys
	GDF-1	Asp	Leu	• • •	•••	•••	Val	• • •	Ala	Arg
	60A	•••	•••		•••	• • •	• • •	• • •	•••	Arg
	BMP5	•••	• • •	•••			•••	•••	•••	Lys
	BMP6	•••	• • •	• • •		•••	•••	•••	• • •	Lys
15				65					70	•
									•	
	hOP-1	Leu	Asn	Ala	Ile	Ser	Val	Leu	Tyr	Phe
	mOP-1	•••	•••	•••	• • •	•••	•••	• • •	• • •	• • •
	hOP-2	•••	Ser	• • •	Thr	•••	• • •	• • •	• • •	Tyr
20	mOP-2	•••	Ser	• • •	Thr	• • •	•••	•••	• • •	Tyr
	Vgl	Met	Ser	Pro	• • •	•••	Met	•••	Phe	Tyr
	Vgr-1	Val	•••	•••	• • •	• • •	•••	•••	•••	•••
	DPP	•••	Asp	Ser	Val	Ala	Het	• • •	•••	Leu
	CBMP-2A	•••	Ser	•••	• • •	• • •	Het	• • •	•••	Leu
25	CBMP-2B	• • •	Ser	•••	• • •	• • •	Het	• • •	• • •	Leu
	BMP3	Het	Ser	Ser	Leu	•••	Ile	•••	Phe	Tyr
	GDF-1	•••	Ser	Pro	•••	•••	•••	• • •	Phe	• • •
	60A	•••	Gly	• • •	Leu	Pro	•••	• • •	•••	His
	BMP5	• • •	•••	•••	• • •	•••	•••	•••	• • •	• • •
30	BMP6	•••	•••	• • •	• • •	• • •	•••	• • •	•••	• • •
					75					80

	hOP-1	Asp	Asp	Ser	Ser	Asn	Val	Ile	Leu	Lys
	mOP-1	. • • •	•••	• • •	• • •	• • •	• • •	• • •	• • •	• • •
	h0P-2	•••	Ser	• • •	Asn		• • •	•••	• • •	Arg
	mOP-2	• • •	Ser	•••	Asn	• • •	• • •	• • •	• • •	Arg
5	DPP	Asn	•••	Gln	•••	Thr	• • •	Val	• • •	• • •
	Vgl	•••	Asn	Asn	Asp	• • •	• • •	Val	• • •	Arg
	Vgr-1	•••	• • •	Asn			• • •	• • •	• • •	• • •
	CBMP-2A		Glu	Asn	Glu	Lys	• • •	Val	•••	• • •
	CBMP-2B	•••	Glu	Týr	Asp	Lys	•••	Val	• • •	• • •
10	вир3	•••	Glu	Asn	Lys	• • •	• • •	Val	• • •	• • •
	GDF-1	•••	Asn	•••	Asp	•••	• • •	Val	•••	Arg
	60A	Leu	Asn	Asp	Glu	• • •	• • •	Asn	• • •	•••
	BMP5	•••	• • •	• • •	•••	• • •	• • •	• • •	• • •	• • •
	BMP6	•••	•••	Asn	• • •	• • •	•••	• • •	• • •	:
15						85				
	•									
	h0P-1	Lys	Tyr	Arg	Asn	Het	Val	Val	Arg	
	mOP-1	•••	•••	• • •	•••	•••	•••	• • •	• • •	
20	hOP-2	•••	His	• • •	•••	• • •	.* • •	• • •	Lys	
•	mOP-2	. •••	His	• • •	•••	• • •	• • •	• • •	Lys	
	DPP	Asn	• • •	Gln	Glu	•••	Thr	•••	Val	
	Vgl	His	• • •	Glu	•••	• • •	Ala	• • •	Asp	
	Vgr-1	•••	• • •	• • •	• • •	•••	• • •	• • •	• • •	
25	CBMP-2A	A		Gln	Asp		• • •		Glu	
		Asn	• • •		-					
	CBMP-2B	Asn	•••	Gln	Glu	• • •	•••	•••	Glu	
					-	•••	 Thr	•••	Glu	
	CBMP-2B	Asn	•••	Gln	Glu					
	CBMP-2B BMP3	Asn Val	•••	Gln Pro	Glu	•••	Thr	• • •	Glu	
30	CBMP-2B BMP3 GDF-1	Asn Val Gln	•••	Gln Pro Glu	Glu Asp	•••	Thr	•••	Glu Asp	
30	CBMP-2B BMP3 GDF-1 60A	Asn Val Gln	•••	Gln Pro Glu	Glu Asp	•••	Thr Ile	•••	Glu Asp Lys	

	hOP-1	Ala	Cys	Gļy	Cys	His
	mOP-1	• • •	• • •	•••	• • •	• • •
	hOP-2	• • •	• • •	• • •	•••	• • •
	mOP-2	• • •	• • •	• • •	• • •	• • •
5	DPP	Gly	• • •	•••	• • •	Arg
	Vgl	Glu	• • • •	• • •	• • •	Arg
	Vgr-1	• • •	• • •	• • •	• • •	• • •
	CBMP-2A	Gly	• • •	•••	• • •	Arg
	CBMP-2B	Gly	• • •	• • •	•••	Arg
10	BMP3	Ser	• • •	Ala	• • • -	Arg
	GDF-1	Glu	• • •	•••	•••	Arg
	60A	Ser	•••	•••	•••	• • •
	BMP5	Ser	• • •	• • •	•••	• • •
	BMP6	•••	• • •	• • •	•••	• • •
15				100		

**Between residues 56 and 57 of BMP3 is a Val residue; between residues 43 and 44 of GDF-1 lies the amino acid sequence Gly-Gly-Pro-Pro.

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As is apparent from the foregoing amino acid sequence comparisons, significant amino acid changes can be made within the generic sequences while retaining the morphogenic activity. For example, while the GDF-1 protein sequence depicted in Table II shares only about 50% amino acid identity with the hOP1 sequence described therein, the GDF-1 sequence shares greater than 70% amino acid sequence homology (or "similarity") with the hOP1 sequence, where "homology" or "similarity" includes allowed conservative amino acid changes within the sequence as defined by Dayoff, et al., Atlas of Protein Sequence and Structure vol.5, supp.3, pp.345-362, (M.O. Dayoff, ed., Nat'l BioMed. Res. Fd'n, Washington D.C. 1979.)

The currently most preferred protein sequences useful as morphogens in this invention include those having greater than 60% identity, preferably greater than 65% identity, with the amino acid sequence defining the conserved six 5 cysteine skeleton of hOP1 (e.g., residues 43-139 of Seq. ID No. 5). These most preferred sequences include both allelic and species variants of the OP-1 and OP-2 proteins, including the Drosophila 60A protein. Accordingly, in still another preferred aspect, the invention includes morphogens 10 comprising species of polypeptide chains having the generic amino acid sequence referred to herein as "OPX", which defines the seven cysteine skeleton and accommodates the identities between the various identified mouse and human OP1 and OP2 proteins. OPX is presented in Seq. ID No. 29. 15 As described therein, each Xaa at a given position independently is selected from the residues occurring at the corresponding position in the C-terminal sequence of mouse or human OP1 or OP2 (see Seq. ID Nos. 5-8 and/or Seq. ID Nos. 16-23).

20

Alternatively, an effective amount of an agent capable of stimulating endogenous morphogen levels may be administered by any of the routes described herein below. For example, an agent capable of stimulating morphogen production and/or secretion from periodontal tissue cells, alveolar bone tissue cells in the fresh tooth socket, or dentin tissue, may be provided to a mammal, e.g., by direct administration of the morphogen-stimulating agent to the tooth root and/or tooth socket bone surface. Alternatively, the morphogen-stimulating agent may induce morphogen expression and/or secretion at a distant site (e.g., at a tissue locus other than periodontal, dental or alveolar bone tissue), with the expressed morphogen targeting itself to periodontal tissue. A method for identifying and testing agents capable of modulating the levels of endogenous

morphogens in a given tissue is described generally herein in Example 3, and in detail in copending USSN [Atty Docket CRP-058CP], filed August 28, 1992 and USSN 752,859, filed August 30, 1991, the disclosures of which are incorporated berein by reference. Briefly, candidate compounds can be identified and tested by incubating the compound in vitro with a test tissue or cells thereof, for a time sufficient to allow the compound to affect the production, i.e., the expression and/or secretion, of a morphogen produced by the cells of that tissue. Here, suitable tissue, or cultured cells of a tissue, preferably comprise periodontal fibroblasts, cementoblasts, odontoblasts or osteoblasts.

III. Formulations and Methods for Administration

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1. Therapeutic Agent Considerations

The morphogens may be provided to the tooth root and/or tooth socket surface by any suitable means. Preferably, the 20 morphogen, or a morphogen-stimulating agent, (collectively, the therapeutic agent) is provided directly to the tissue surface by topical administration. Alternatively, the therapeutic agent may be provided to the tissue by, for example, local injection. While not currently preferred, 25 systemic injection also may be a viable administration route for certain applications, such as periodontal tissue maintenance in older adults, immuno-suppressed individuals, or others at chronic risk for periodontal tissue loss. A detailed description of considerations for systemic 30 administration, including oral and parenteral administration, is disclosed, for example, in international application US92/07358 (WO93/04692), incorporated hereinabove by reference.

Where the therapeutic agent is provided directly to the tooth socket, the therapeutic agent may be provided to the socket surface as part of a biocompatible formulation that may be a liquid, gel or solid. The therapeutic agent

5 further may be dispersed in and associated with a carrier capable of maintaining the morphogen at the administered locus. Useful formulations include viscous compositions. Biocompatible compositions that increase the viscosity of the formulation include glycerol, polyalkylene glycols such as polyethylene glycol, oils of vegetable origin, hydrogenated naphthalenes, and the like.

The formulation also may include an in vivo bioresorbable carrier material that acts as a controlled 15 release delivery vehicle. Useful carriers may include biocompatible, preferably biodegradable structural components from, e.g., an extracellular matrix, such as collagen, laminin, hyaluronic acid, and the like, or polymeric materials, such as polylactic, polybutyric and 20 polyglycolic acids. The carrier also may comprise an acellular tissue matrix, substantially depleted in nonstructural components, such as a demineralized, guanidine-extracted dentin, periodontal ligament or cementum matrix. Details for preparing such matrices are disclosed 25 in international application US92/01968 (WO93/15323). Other useful controlled release carriers in which the therapeutic agent may be dispersed are described in U.S. Pat. Nos. 4,975,526 and 4,919,939, the disclosures of which are incorporated herein by reference.

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Where the morphogen is to be provided to a tooth root surface, it may be formulated in a composition for controlled delivery as described above and applied topically to the tooth root surface as described below.

35 Alternatively, or in addition, the therapeutic agent may be

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dispersed in a liquid formulation into which at least the tooth root surface is placed and the liquid lyophilized to adsorb the therapeutic agent onto the tooth surface.

5 Where the agent is administered to inhibit periodontal tissue loss and/or to regenerate periodontal tissue surrounding an implanted tooth, the agent may be provided to the area between the tooth and gum (gingiva) by injection or by topical application.

10

Where the morphogen is to be provided directly (e.g., locally, as by injection, e.g., to a periodontal or alveolar tissue site), the morphogen preferably comprises part of an aqueous solution which also may contain a carrier material.

- 15 The solution is physiologically acceptable so that in addition to delivery of the desired morphogen to the patient, the solution does not otherwise adversely affect the patient's electrolyte and volume balance. The aqueous medium for the morphogen thus may comprise normal
- 20 physiologic saline (0.85% NaCl, 0.15M), pH 7-7.4. aqueous solution containing the morphogen can be made, for example, by dissolving the protein in 50% ethanol containing acetonitrile in 0.1% trifluoroacetic acid (TFA) or 0.1% HCl, or equivalent solvents. One volume of the resultant
- solution then is added, for example, to ten volumes of phosphate buffered saline (PBS), which further may include 0.1-0.2% human serum albumin (HSA). The resultant solution preferably is vortexed extensively. If desired, a given morphogen may be made more soluble in the solution by
- association with a suitable molecule. For example, the pro form of the morphogenic protein comprises a species that is soluble in physiological solutions. In fact, the endogenous protein is thought to be transported (e.g., secreted and circulated) in this form. This soluble form of the protein
- may be obtained from the culture medium of

morphogen-secreting mammalian cells. Alternatively, a soluble species may be formulated by complexing the mature dimer (or an active fragment thereof) with part or all of a pro domain. Other components, including various serum proteins, also may be useful. A more detailed description for formulating soluble morphogen complexes appears in Example 4, below.

Finally, the morphogens or morphogen-stimulating agents 10 provided herein may be administered alone or in combination with other molecules, particularly symptom alleviating cofactors. Useful pharmaceutical cofactors include antiseptics, antibiotics, anaesthetics and analgesics. Preferred antiseptics for use in the present system include 15 chlorhexidine and tibezonium iodide; preferred antibiotics include tetracycline, aminoglycosides such as neomycin, gentamycin, kanamycin, tobramycin, netilmicin, sisomicin, amicamycin, their sulfates or other derivatives, macrolides such as erythromycin, its salts and other derivatives, spiramycin, josamicin or miocamicin, penicillins such as 20 ampicillin, amoxicillin and the like, and cephalosporins, for example, cefaclor, cefadroxil, cefazolin, cefoperazone, cefotaxime, cephalothin, cefalexin, ceforanide, cefonicide or ceftriaxone. Preferred anaesthetics/analgesics include 25 amide-type local anaesthetics such as lidocaine, mepivacaine, pyrrocaine, bupivacaine, prilocaine, etidocaine, or other widely used anaesthetics such as procaine.

30 Other cofactors include non-steroidal anti-inflammatory agents. However, the morphogens described herein themselves modulate the body's inflammatory/immune response to an initial tissue injury. Specifically, and as described in detail in international application US92/07358 (WO93/04692), in the presence of a morphogen, progenitor inflammatory

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effector cells induced to migrate to a site of tissue injury do not become significantly activated. Without being limited to any given theory, it is thought that, in the presence of the morphogen, damaged tissue is induced to undergo a recapitulation of tissue morphogenesis, where progenitor cells are induced to proliferate and differentiate in a tissue-specific manner, and new, functional, organized tissue is formed to replace the damaged or lost tissue, rather than disorganized, fibrous scar tissue.

The formulated compositions contain therapeutically effective amounts of the morphogen, e.g., amounts which provide appropriate concentrations of the morphogen to the tooth surface for a time sufficient to stimulate growth and development of periodontal tissues, including morphogenesis of periodontal ligament and/or cementum, and/or to substantially inhibit periodontal tissue loss.

20 As will be appreciated by those skilled in the art, the concentration of the compounds described in a therapeutic composition will vary depending upon a number of factors, including the biological efficacy of the selected morphogen, the chemical characteristics (e.g., hydrophobicity) of the 25 compounds employed, the formulation of the compound excipients, the administration route, and the treatment envisioned. The preferred dosage to be administered also is likely to depend on such variables such as the condition of the tissues within the tooth socket, the size of the tooth 30 or tooth socket, the length of time after tooth loss, extent of periodontal tissue loss and the overall health status of the particular patient. The amount of morphogen applied also will depend on the tooth size. In general, 0.1-1000 μg of morphogen are sufficient with 1-100 μ g being preferable. 35 For example, for a large tooth, e.g., an incisor or large

molar, about 10-100 μg, and preferably 50 μg of morphogen,
may be used to advantage; a medium tooth may be treated with
approximately 5-50 μg , and preferably 25 μg; and a small
tooth, with approximately 1-25, preferably 5-10 μg
5 morphogen. No obvious morphogen induced pathological
lesions are induced when mature morphogen (e.g., OP-1, 20
μg) is administered daily to normal growing rats for
21 consecutive days. Moreover, 10 μg systemic injections of
morphogen (e.g., OP-1) injected daily for 10 days into
10 normal newborn mice does not produce any gross
abnormalities.

2. Tooth Preparation

Tooth loss may be repaired by implanting a viable tooth having a healthy root and pulp system or by implanting a tooth prosthesis. The prosthesis may be a tooth from which the root has been removed and replaced with a biocompatible, biologically inert material, e.g., as typically is replaced in a root canal procedure, or may be a completely synthetic prosthesis coated, for example, with a porous material to enhance tooth integration in the tooth socket. Useful prosthesis coating materials include collagen fibers, ceramics and metals, such as titanium oxide. The root of the implanted tooth first may be partially demineralized as described below. Alternatively, a clean, mineralized natural tooth or dentin-containing prosthetic tooth may be implanted.

A tooth to be implanted first is obtained, e.g., by loss or removal of a natural tooth from the tooth socket, e.g., using standard tooth extraction means well known to one skilled in the dentistry art. Alternatively, an allogenic tooth may be obtained from a tooth bank. The natural,

35 mineralized tooth or tooth root may be coated as is with a

morphogen and implanted as described below. Alternatively, the mineralized, natural tooth root surface first may be scored or scraped to expose dentin tissue beneath the enamel. Natural, mineralized teeth also may be treated 5 briefly with an acidic solution (e.g., sodium citrate, about pH 3.5) to remove a thin external layer, e.g., about 1-5 cells in thickness from at least the root surface. Preferred treatment times are from about 0.5 to 5 minutes. The treated teeth preferably then are washed, dried and 10 coated with morphogen as described below. Alternatively, the tooth root portion may be at least partially demineralized according to any conventional procedure prior to implantation. A currently preferred demineralization method is to soak the tooth in a demineralizing solution for a length of time sufficient to remove at least some mineral components from the tooth. For example, at least the root portion of the tooth may be placed in a volume, e.g., 0.025-1 liter of a demineralizing agent such as hydrochloric acid (HCl) at a cool temperature for a time sufficient to achieve 20 partial demineralization, e.g., 0.5-0.6 M HCl at 4°C for a prescribed number of minutes (e.g., preferably within the range of about 10-200 minutes.) Essentially complete demineralization may be achieved by acid exposure for 1-7 days. If desired, several changes of the demineralizing 25 agent may be performed. The partially demineralized tooth will be of the same shape as prior to demineralization, but will weigh less due to the absence of the mineral content. The tooth then may be dried by lyophilization.

30 The tooth or tooth prosthesis may be treated with morphogenic protein as follows. The morphogen may be applied to the tooth or tooth prosthesis root surface by any means known in the art for adsorbing a protein to a surface. A currently preferred method is to suspend the morphogen in a small volume sufficient to cover the tooth surface, e.g.,

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200-300µl, freeze the tooth in solution, and then lyophilize the frozen liquid. A currently preferred solution is ethanol (e.g. 50%) or acetonitrile/trifluroactic acid (TFA), other solutions include HCL/TFA, buffered saline, and the like. Alternatively, or in addition, the therapeutic agent may be provided to the tooth root surface dispersed in a suitable carrier material as described above. Similarly, and as described above, the therapeutic agent may be provided to the tooth socket surface and the tooth to be implanted embedded in the morphogen composition on the socket surface. Also as described above, the morphogen may be provided to the tooth root surface in admixture with one or more cofactors.

15 The tooth then is implanted into a fresh or surgically prepared tooth socket. A surgically prepared surface is prepared by extracting the tooth and removing any scar or other undesired fibrous tissue built up in the socket by standard mechanical and/or chemical procedures well known on the surgical and dental arts. The tooth then is implanted in the site using standard dental and surgical procedures.

The implanted tooth is allowed to grow in the prepared socket for a time sufficient to allow the periodontium to regenerate, e.g., one to several months. The integrity and health of the integrated tooth then may be assessed by a dentist by radiography and visual examination.

For experimental purposes, the integration of an implanted tooth following morphogen treatment can be assessed for integrity and health by removing the entire mandibular area, including the tooth socket and tooth, and examining cross sections of the mandibular area. 5-10 μm cross sections may be prepared for histological evaluation by standard histology procedures, e.g., fixing tissue with

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formalin, preparing sections for slides and staining with eosin and hematoxylin. The growth and integrity of hard tissues, such as bone, cementum and dentin, also can evaluated radiographically.

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Finally, as described in Example 2 below, the morphogens of this invention also induce dentin tissue morphogenesis when provided to an area of lost or damaged dentin.

Accordingly, using the procedures described herein and in international application (US92/01968 (WO92/15323), the morphogen described herein also may be used to repair and regenerate damaged and/or lost dentin tissue in an implanted tooth.

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IV. Examples

Example 1. Experimental Regeneration of Peridontium in a Dog Model

20

The following experiment demonstrates successful integration of an implanted demineralized, protein-extracted morphogen-treated tooth in a mammal. Premolar teeth were extracted from a dog and divided into three experimental groups: (a) demineralized teeth; (b) demineralized and guanidine extracted teeth; and (c) demineralized, guanidine extracted, and morphogen-treated teeth. Teeth from each group were tested in "fresh" sockets, e.g., tooth sockets from which the teeth had just been removed, as well as surgically prepared sockets, e.g., sockets from which teeth had been extracted 2 months previously and in which scar tissue had formed. These "healed" sockets were surgically prepared for tooth implantation by removing (e.g., by scraping) scar tissue build up to reveal fresh alveolar bone.

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The teeth from all three groups were completely demineralized by placing them in 4 liters of 0.5 M HCl at 4°C for 5 days. The 0.5 M HCl solution was changed every 24 hours during the 5 day period. The teeth then were washed in 4 liters of deionized water at 4°C for 5 days. The water solution was also changed every 24 hours during the 5 day period. Teeth from group (a) then were lyophilized until dry and set aside and maintained at 4°C until ready for use.

10 Teeth from groups (b) and (c) then were protein-extracted by multiple extractions in 6 M quanidine Hcl, followed by washes with distilled water. Specifically, the teeth were placed in in 2-4 liters of 6 M guanidine-HCl/Tris HCl pH 7.0 at 4°C for 72 hours; then washed and 15 further extracted in 200 ml of the guanidine-HCl solution for 4 hours. The teeth were washed again with 4 liters of distilled dH₂O at 4°C for 48 hours, and 4 liters of dH₂O for an additional 12 hours with 3 changes of dH₂O. The teeth were then lyophilized until dry. Teeth from group (b) were 20 then set aside and maintained at 4°C until ready for use.

Teeth from group (c) then were treated with the morphogen OP-1 as follows. 1.15 mg of OP-1 was resuspended in 4 ml of 47.5% ethanol/0.09% trifluoroacetic acid (TFA).

25 The concentration was determined to be 0.273 mg/ml. Approximately 50 µg of OP-1 (183 µl of the OP-1 solution) was dispensed into an eppendorf tube, and the total volume brought to 300 µl of 47.5% ethanol/0.09% TFA. Each tooth then was placed in an eppendorf tube such that the OP-1 solution just covered the tooth. The tube was placed at 70°C until the OP-1 solution was frozen, and lyophilized until dry. During lyophilization, care was taken to keep the tube cold. Approximately 50-70% of the OP-1 can be expected to remain in or on the tooth after lyophilization.

The teeth from each of groups (a), (b), and (c) were then implanted into a freshly prepared tooth socket or surgically prepared socket using standard dental surgery procedures known in the art.

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10

The implanted teeth in all three groups were allowed to remain in the socket for two months. The dog then was sacrificed, the mandible cross-sectioned and x-rayed, and histology performed. The results are described below and follows.

Ankylosis formed in the group (a) implants, where demineralized tooth matrix was implanted alone. Cross-sections of the group (a) mandible revealed that the demineralized tooth was surrounded by bone directly attaching to the root or dentin surface. In addition, there was little new tissue growth between the tooth and the bone. Representative histology is illustrated in the photomicrograph of Fig. 2A where bone tissue 14 grows directly into dental tissue 22 in the implanted tooth.

In the group (b) implants, cross-sections revealed formation of unorganized fibrous tissue around the implanted demineralized, guanidine extracted tooth. The periodontal ligament was loose and disorganized, as was the surrounding bony tissue. Examination of the tooth root surface where cementum matrix normally appears revealed resorption of cementum in the upper coronal surface of the tooth. Histological sections also revealed inflammation as evidenced by the presence of macrophages.

As is evident in Fig. 2b, group (c) implant crosssections revealed formation of newly formed, organized cementum 24 and periodontal ligament tissue 26 around the 35 morphogen-treated tooth matrix, and growth of new bone connecting the newly form d periodontium to the mandible.

The tooth was firmly anchored in the tooth socket. The
tissues surrounding the tooth, i.e., the newly-formed
cementum growing perpendicular to the newly-formed

periodontal ligament, and the alveolar bony tissue, all were
healthy and organized much as the tooth and tooth socket
shown schematically in Fig. 1. The newly-formed cementum
comprised immature columnar cell layers which were beginning
to flatten into mature cementoblasts, and the newly-formed
periodontal ligament comprised a thick layer of tissue to
anchor and cushion the tooth within the tooth socket.

The results of this experiment demonstrate that morphogens promote tooth integration into a tooth socket, and induce morphogenesis of periodontium, including morphogenesis of the regeneration and formation of the periodontium, new cementum and periodontal ligament.

Without being limited to any particular theory, the
morphogens may act in the tooth socket environment by
inducing a differentiation of primary fibroblasts on the
alveolar surface to differentiate into cementoblasts which
then induct other primary fibroblasts to form periodontal
ligament.

25

Example 2. Morphogen-Induced Dentinogenesis

The examples presented below demonstrate the efficacy of morphogens in inducing dentin tissue morphogenesis in an animal model. Further details of the first experiment and the implications of this biological activity of morphogens are disclosed in international application (US92/01968 (WO92/15323).

To date, the unpredictable response of dental pulp tissue to injury is a basic clinical problem in dentistry. Cynomolgus monkeys were chosen as primate models for the reparative dentine/pulp capping examples described below.

5

Using standard dental surgical procedures, small areas (e.g., 2mm) of dental pulps were surgically exposed by removing the enamel and dentin immediately above the pulp (by drilling) of sample teeth, performing a partial

10 amputation of the coronal pulp tissue, inducing hemostasis, application of the pulp treatment, and sealing and filling the cavity by standard procedures.

Pulp treatments used were: OP1 dispersed in a carrier 15 matrix; carrier matrix alone and no treatment. Twelve teeth per animal (four for each treatment) were prepared, and two animals were used. At four weeks, teeth were extracted and processed histologically for analysis of dentin formation, and/or ground to analyze dentin mineralization. Morphogen 20 treatment produced dramatic effects: Control treatments with carrier alone or with no treatment (PBS) showed little or no reparation of the lost tissue. By contrast, morphogen-treated teeth showed significant dentin tissue formation in the area where dentin tissue had been 25 surgically removed. The experimental results show that morphogen treatment reliably induced formation of reparative or osteodentin bridges on surgically exposed healthy dental pulps. See, for example, Fig. 3A, where OP1 dispersed in a carrier (demineralized, guanidine-extracted bone collagen matrix prepared as described in U.S. Patent No. 4,975,526) constituted the pulp treatment. As is evident from the micrograph new dentine formation effectively bridges or "caps" the surgically exposed dental pulp, maintaining the integrity and viability of the pulp tissue. By contrast, 35 pulps treated with carrier matrix alone, or not treated,

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failed to form reparative dentin. See, for example Fig. 3B where carrier alone (demineralized, guanidine-extracted bone collagen matrix prepared as described in U.S. Patent No. 4,975,526) constituted the pulp treatment. As is evident from the micrograph, minimal reparative dentin formed, insufficient to bridge the exposed pulp tissue. Without further treatment such exposed, unprotected pulp tissue will become infected and die.

In a supplemental experiment, a range of morphogen 10 concentrations were tested. In all cases, human OP-1, prepared as described in Sampath et al. (1992) J. Biol. Chem. 267: 20352-20362, was the morphogen tested, and bone collagen matrix, prepared as described in U.S. 15 Patent No. 4,975,526 was the carrier material/delivery vehicle ("CM"). Briefly, cortical bone powder was prepared from freshly obtained bovine femurs. The epiphyses, adherent flesh and marrow were removed and residual lipids extracted with hexane, isopropanol, and ethyl ether. The 20 resulting material was ground and sieved to a described particle size of 75-425 μm . The cortical bone powder then was demineralized in acid, and soluble proteins extracted with quanidine hydrochloride. The demineralized, extracted bone powder then was subjected to a thermal acid treatment, 25 washed with water, and lyophilized. The final dry powder was sieved to remove particles >425 µm and stored at 4°C.

The hOP-1/CM samples were prepared by combining hOP-1 with the CM and drying under vacuum. The batch used in 30 these experiments contained 2.5µg hOP-1/mg CM. Prior to implant the sample was moistened with a sterile aqueous solution, preferably saline, to form a paste-like substance. CM controls were prepared using the same procedure, omitting the morphogen. The samples were stored at -20°C until used.

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The pulp capping experiments were conducted using 4 adult female non-human primates (Macaca fasicularis) of approximately 4 kg each. The animals were sedated using standard procedures, e.g., with ketamine (15 mg/kg body wt.) and acepromazine (0.55 mg/kg body wt.) supplemented with local intraoral infiltration anesthesia.

Thirty premolar and molar teeth in four animals were isolated by rubber dam and the pulps exposed using standard 10 dentistry procedures, e.g., using sterile high speed rotary cutting instruments with water spray coolant. The pulp exposures made were approximately 1-1.5 by 2-2.5 mm. Partial hemostasis was achieved with sterile cotton pellets but the teeth were not dried extensively prior to treatment. 15 The exposed pulps were treated with: hOP-1/CM (2.5 μg hOP-1/mg CM) at 1.5, 3.0 or 6.0 mg/tooth; or one of three controls: Ca(OH), paste, a standard pulp capping agent used in the art ("Dycal", L.D. Caulk, Milford, DE); CM alone, 3.0 mg/tooth; or no treatment material. The teeth then were 20 sealed with a standard adhesive, e.g., Temp-Bond NETM (Kerr U.S.A., Romulus, MI). The teeth were allowed to heal for six weeks. No changes in behavior were noted by any of the animals during the healing period.

The animals were sacrificed six weeks following surgery and prepared for histomorphometric analysis using standard procedures. For example, teeth were fixed by immersion in 10% formalin in phosphate buffered saline (pH 7.2) and decalcified in formic acid/sodium citrate at room temperature for 6-8 days. The specimens were processed, imbedded in paraffin, serial sectioned (5 μ m) and stained.

In all teeth treated with hOP-1/CM and for all OP1 concentrations tested, reparative dentine sufficient to 35 bridge the surgically created gap that exposed the

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underlying pulp tissue was formed. As in the previous experiment, the morphogen/CM device was resorbed in the healed teeth, and replaced with reparative dentine, fully integrated with the cut dentine at the exposure site. Also 5 as in the previous experiment, the pulp tissue beneath the cap appeared normal, with intact odontoblasts lining the pulp chamber. The amount of new dentine tissue increased as the amount of OP-1 provided in a sample was increased, indicating that the amount of reparative dentine formed was 10 related to the mass of OP1/CM administered. Pulp treatments using CM alone or no treatment did not succeed in bridging the exposure site and in several cases resulted in necrotic pulp tissue. Treatments using Ca(OH), succeeded in bridging the gap, but the paste remained and the bridge created lies 15 within the pulp chamber itself.

Example 3. Screening Assay for Candidate Compounds which Alter Endogenous Morphogen Levels

20 Candidate compound(s) which may be administered to affect the level of a given morphogen may be found using the following screening assay, in which the level of morphogen production by a cell type which produces measurable levels of the morphogen is determined with and without incubating the cell in culture with the compound, in order to assess the effects of the compound on the cell. This can be accomplished by detection of the morphogen either at the protein or RNA level. A more detailed description also may be found in international application US92/07359

30 (WO93/05172), incorporated hereinabove by reference.

3.1 Growth of Cells in Culture

Cell cultures of kidney, adrenals, urinary bladder, 35 brain, or other organs, may be prepared as described widely in the literature. For example, kidneys may be explanted from neonatal or new born or young or adult rodents (mouse or rat) and used in organ culture as whole or sliced (1-4 mm) tissues. Primary tissue cultures and established cell lines, also derived from kidney, adrenals, urinary, bladder, brain, mammary, or other tissues may be established in multiwell plates (6 well or 24 well) according to conventional cell culture techniques, and are cultured in the absence or presence of serum for a period of time (1-7 days). Cells may be cultured, for example, in Dulbecco's Modified Eagle medium (Gibco, Long Island, NY) containing serum (e.g., fetal calf serum at 1%-10%, Gibco) or in serum-deprived medium, as desired, or in defined medium (e.g., containing insulin, transferrin, glucose, albumin, or other growth factors).

Samples for testing the level of morphogen production includes culture supernatants or cell lysates, collected periodically and evaluated for OP-1 production by immunoblot analysis (Sambrook et al., eds., 1989, Molecular Cloning, Cold Spring Harbor Press, Cold Spring Harbor, NY), or a portion of the cell culture itself, collected periodically and used to prepare polyA+ RNA for RNA analysis. To monitor de novo OP-1 synthesis, some cultures are labeled according to conventional procedures with an 35 S-methionine/
35 S-cysteine mixture for 6-24 hours and then evaluated for OP-1 synthesis by conventional immunoprecipitation methods.

3.2 Determination of Level of Morphogenic Protein

30

In order to quantitate the production of a morphogenic protein by a cell type, an immunoassay may be performed to detect the morphogen using a polyclonal or monoclonal antibody specific for that protein. For example, OP-1 may

be detected using a polyclonal antibody specific for OP-1 in an ELISA, as follows.

1 μ q/100 μ l of affinity-purified polyclonal rabbit IgG 5 specific for OP-1 is added to each well of a 96-well plate and incubated at 37°C for an hour. The wells are washed four times with 0.167M sodium borate buffer with 0.15 M NaCl (BSB), pH 8.2, containing 0.1% Tween 20. To minimize non-specific binding, the wells are blocked by filling 10 completely with 1% bovine serum albumin (BSA) in BSB and incubating for 1 hour at 37°C. The wells are then washed four times with BSB containing 0.1% Tween 20. A 100 μ l aliquot of an appropriate dilution of each of the test samples of cell culture supernatant is added to each well in 15 triplicate and incubated at 37°C for 30 min. After incubation, 100 µl biotinylated rabbit anti-OP-1 serum (stock solution is about 1 mg/ml and diluted 1:400 in BSB containing 1% BSA before use) is added to each well and incubated at 37°C for 30 min. The wells are then washed 20 four times with BSB containing 0.1% Tween 20. 100 μ l strepavidin-alkaline (Southern Biotechnology Associates, Inc. Birmingham, Alabama, diluted 1:2000 in BSB containing 0.1% Tween 20 before use) is added to each well and incubated at 37°C for 30 min. The plates are washed four 25 times with 0.5M Tris buffered Saline (TBS), pH 7.2. 50μ l substrate (ELISA Amplification System Kit, Life Technologies, Inc., Bethesda, MD) is added to each well incubated at room temperature for 15 min. Then, 50 μ l amplifier (from the same amplification system kit) is added and incubated for another 15 min at room temperature. The reaction is stopped by the addition of 50 μ l 0.3 M sulphuric The OD at 490 nm of the solution in each well is recorded. To quantitate OP-1 in culture media, a OP-1 standard curve is performed in parallel with the test 35 samples.

Polyclonal antibody may be prepared as follows. Each rabbit is given a primary immunization of 100 ug/500 µl E. coli produced OP-1 monomer (amino acids 328-431 in SEQ ID NO:5) in 0.1% SDS mixed with 500 µl Complete Freund's Adjuvant. The antigen is injected subcutaneously at multiple sites on the back and flanks of the animal. The rabbit is boosted after a month in the same manner using incomplete Freund's Adjuvant. Test bleeds are taken from the ear vein seven days later. Additional boosts and test bleeds are performed at monthly intervals until antibody against OP-1 is detected in the serum using an ELISA assay. Then, the rabbit is boosted with 100 µg of antigen and bled (15 ml per bleed) at days seven and ten after boosting.

15

Monoclonal antibody specific for a given morphogen may be prepared as follows. A mouse is given two injections of E. coli produced OP-1 monomer. The first injection contains $100\mu g$ of OP-1 in complete Freund's adjuvant and is given 20 subcutaneously. The second injection contains 50 μ g of OP-1 in incomplete adjuvant and is given intraperitoneally. The mouse then receives a total of 230 μ g of OP-1 (amino acids 307-431 in SEQ ID NO:5) in four intraperitoneal injections at various times over an eight month period. One week prior 25 to fusion, the mouse is boosted intraperitoneally with 100 μg of OP-1 (307-431) and 30 μg of the N-terminal peptide (Ser₂₉₃-Asn₃₀₉-Cys) conjugated through the added cysteine to bovine serum albumin with SMCC crosslinking agent. This boost was repeated five days (IP), four days (IP), three 30 days (IP) and one day (IV) prior to fusion. The mouse spleen cells are then fused to myeloma (e.g., 653) cells at a ratio of 1:1 using PEG 1500 (Boeringer Mannheim), and the cell fusion is plated and screened for OP-1-specific antibodies using OP-1 (307-431) as antigen. The cell fusion 35 and monoclonal screening then are according to standard

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procedures well described in standard texts widely available in the art.

Example 4. Soluble Morphogen Complexes

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A currently preferred form of the morphogen useful in therapeutic formulations for systemic administration, having improved solubility in aqueous solutions and consisting essentially of amino acids, is a dimeric morphogenic protein 10 comprising at least the 100 amino acid peptide sequence having the pattern of seven or more cysteine residues characteristic of the morphogen family complexed with a peptide comprising part or all of a pro region of a member of the morphogen family, or an allelic, species or other 15 sequence variant thereof. Preferably, the dimeric morphogenic protein is complexed with two peptides. Also, the dimeric morphogenic protein preferably is noncovalently complexed with the pro region peptide or peptides. region peptides also preferably comprise at least the 20 N-terminal eighteen amino acids that define a given morphogen pro region. In a most preferred embodiment, peptides defining substantially the full length pro region are used.

Other soluble forms of morphogens include dimers of the uncleaved pro forms of these proteins, as well as "hemi-dimers" wherein one subunit of the dimer is an uncleaved pro form of the protein, and the other subunit comprises the mature form of the protein, including truncated forms thereof, preferably noncovalently associated with a cleaved pro domain peptide.

As described above, useful pro domains include the full length pro regions, as well as various truncated forms

35 hereof, particularly truncated forms cleaved at proteolytic

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Arg-Xaa-Xaa-Arg cleavage sites. For example, in OP-1, possible pro sequences include sequences defined by residues 30-292 (full length form); 48-292; and 158-292. Soluble OP-1 complex stability is enhanced when the pro region comprises the full length form rather than a truncated form, such as the 48-292 truncated form, in that residues 30-47 show sequence homology to the N-terminal portions of other morphogens, and are believed to have particular utility in enhancing complex stability for all morphogens.

10 Accordingly, currently preferred pro sequences are those encoding the full length form of the pro region for a given morphogen. Other pro sequences contemplated to have utility include biosynthetic pro sequences, particularly those that incorporate a sequence derived from the N-terminal portion of one or more morphogen pro sequences.

As will be appreciated by those having ordinary skill in the art, useful sequences encoding the pro region may be obtained from genetic sequences encoding known morphogens.

20 Alternatively, chimeric pro regions can be constructed from the sequences of one or more known morphogens. Still another option is to create a synthetic sequence variant of one or more known pro region sequences.

- In another preferred aspect, useful pro region peptides include polypeptide chains comprising an amino acid sequence encoded by a nucleic acid that hybridizes under stringent conditions with a DNA or RNA sequence encoding at least the N-terminal eighteen amino acids of the pro region sequence for OP1 or OP2, e.g., nucleotides 136-192 and 152-211 of Seq. ID No. 16 and 20, respectively.
 - 4.1 Isolation of Soluble morphogen complex from conditioned media or body fluid

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Morphogens are expressed from mammalian cells as soluble complexes. Typically, however the complex is disassociated during purification, generally by exposure to denaturants often added to the purification solutions, such as

5 detergents, alcohols, organic solvents, chaotropic agents and compounds added to reduce the pH of the solution.

Provided below is a currently preferred protocol for purifying the soluble proteins from conditioned media (or, optionally, a body fluid such as serum, cerebro-spinal or peritoneal fluid), under non-denaturing conditions. The method is rapid, reproducible and yields isolated soluble morphogen complexes in substantially pure form.

Soluble morphogen complexes can be isolated from 15 conditioned media using a simple, three step chromatographic protocol performed in the absence of denaturants. protocol involves running the media (or body fluid) over an affinity column, followed by ion exchange and gel filtration chromatographies. The affinity column described below is a 20 Zn-IMAC column. The present protocol has general applicability to the purification of a variety of morphogens, all of which are anticipated to be isolatable using only minor modifications of the protocol described below. An alternative protocol also envisioned to have 25 utility an immunoaffinity column, created using standard procedures and, for example, using antibody specific for a given morphogen pro domain (complexed, for example, to a protein A-conjugated Sepharose column.) Protocols for developing immunoaffinity columns are well described in the 30 art, (see, for example, Guide to Protein Purification, M. Deutscher, ed., Academic Press, San Diego, 1990, particularly sections VII and XI.)

In this experiment OP-1 was expressed in mammalian CHO (chinese hamster ovary) cells as described in the art (see,

for example, international application US90/05903 (WO91/05802).) The CHO cell conditioned media containing 0.5% FBS was initially purified using Immobilized Metal-Ion Affinity Chromatography (IMAC). The soluble OP-1 complex 5 from conditioned media binds very selectively to the Zn-IMAC resin and a high concentration of imidazole (50 mM imidazole, pH 8.0) is required for the effective elution of The Zn-IMAC step separates the soluble the bound complex. OP-1 from the bulk of the contaminating serum proteins that 10 elute in the flow through and 35 mM imidazole wash fractions. The Zn-IMAC purified soluble OP-1 is next applied to an S-Sepharose cation-exchange column equilibrated in 20 mM NaPO, (pH 7.0) with 50 mM NaCl. This S-Sepharose step serves to further purify and concentrate 15 the soluble OP-1 complex in preparation for the following gel filtration step. The protein was applied to a Sephacryl S-200HR column equilibrated in TBS. Using substantially the same protocol, soluble morphogens also may be isolated from one or more body fluids, including serum, cerebro-spinal 20 fluid or peritoneal fluid.

IMAC was performed using Chelating-Sepharose (Pharmacia) that had been charged with three column volumes of 0.2 M ZnSO4. The conditioned media was titrated to pH 7.0 and applied directly to the ZN-IMAC resin equilibrated in 20 mM HEPES (pH 7.0) with 500 mM NaCl. The Zn-IMAC resin was loaded with 80 mL of starting conditioned media per mL of resin. After loading, the column was washed with equilibration buffer and most of the contaminating proteins were eluted with 35 mM imidazole (pH 7.0) in equilibration buffer. The soluble OP-1 complex then is eluted with 50 mM imidazole (pH 8.0) in 20 mM HEPES and 500 mM NaCl.

The 50 mM imidazole eluate containing the soluble OP-1 35 complex was diluted with nine volumes of 20 mM NaPO₄ (pH

7.0) and applied to an S-Sepharose (Pharmacia) column equilibrated in 20 mM NaPO, (pH 7.0) with 50 mM NaCl. S-Sepharose resin was loaded with an equivalent of 800 mL of starting conditioned media per mL of resin. After loading 5 the S-Sepharose column was washed with equilibration buffer and eluted with 100 mM NaCl followed by 300 mM and 500 mM NaCl in 20 mM NaPO, (pH 7.0). The 300 mM NaCl pool was further purified using gel filtration chromatography. Fifty mls of the 300 mm NaCl eluate was applied to a 5.0 X 90 cm 10 Sephacryl S-200HR (Pharmacia) equilibrated in Tris buffered saline (TBS), 50 mM Tris, 150 mM NaCl (pH 7.4). The column was eluted at a flow rate of 5 mL/minute collecting 10 mL fractions. The apparent molecular of the soluble OP-1 was determined by comparison to protein molecular weight 15 standards (alcohol dehydrogenase (ADH, 150 kDa), bovine serum albumin (BSA, 68 kDa), carbonic anhydrase (CA, 30 kDa) and cytochrome C (cyt C, 12.5 kDa). The purity of the S-200 column fractions was determined by separation on standard 15% polyacrylamide SDS gels stained with coomassie blue. 20 The identity of the mature OP-1 and the pro-domain was determined by N-terminal sequence analysis after separation of the mature OP-1 from the pro-domain using standard reverse phase C18 HPLC.

The soluble OP-1 complex elutes with an apparent molecular weight of 110 kDa. This agrees well with the predicted composition of the soluble OP-1 complex with one mature OP-1 dimer (35-36 kDa) associated with two pro-domains (39 kDa each). Purity of the final complex can be verified by running the appropriate fraction in a reduced 15% polyacrylamide gel.

The complex components can be verified by running the complex-containing fraction from the S-200 or S-200HR

35 columns over a reverse phase C18 HPLC column and eluting in

an acetonitrile gradient (in 0.1% TFA), using standard procedures. The complex is dissociated by this step, and the pro domain and mature species elute as separate species. These separate species then can be subjected to N-terminal 5 sequencing using standard procedures (see, for example, Guide to Protein Purification, M. Deutscher, ed., Academic Press, San Diego, 1990, particularly pp. 602-613), and the identity of the isolated 36kD, 39kDa proteins confirmed as mature morphogen and isolated, cleaved pro domain, 10 respectively. N-terminal sequencing of the isolated pro domain from mammalian cell produced OP-1 revealed 2 forms of the pro region, the intact form (beginning at residue 30 of Seq. ID No. 16) and a truncated form, (beginning at residue 48 of Seq. ID No. 16.) N-terminal sequencing of the 15 polypeptide subunit of the isolated mature species reveals a range of N-termini for the mature sequence, beginning at residues 293, 300, 313, 315, 316, and 318, of Seq. ID No. 16, all of which are active as demonstrated by the standard bone induction assay.

20

4.2 In Vitro Soluble Morphogen Complex Formation

As an alternative to purifying soluble complexes from culture media or a body fluid, soluble complexes may be

25 formulated from purified pro domains and mature dimeric species. Successful complex formation apparently requires association of the components under denaturing conditions sufficient to relax the folded structure of these molecules, without affecting disulfide bonds. Preferably, the

30 denaturing conditions mimic the environment of an intracellular vesicle sufficiently such that the cleaved pro domain has an opportunity to associate with the mature dimeric species under relaxed folding conditions. The concentration of denaturant in the solution then is

35 decreased in a controlled, preferably step-wise manner, so

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as to allow proper refolding of the dimer and pro regions while maintaining the association of the pro domain with the dimer. Useful denaturants include 4-6M urea or guanidine hydrochloride (GuHCl), in buffered solutions of pH 4-10, 5 preferably pH 6-8. The soluble complex then is formed by controlled dialysis or dilution into a solution having a final denaturant concentration of less than 0.1-2M urea or GuHCl, preferably 1-2 M urea of GuHCl, which then preferably can be diluted into a physiological buffer. Protein 10 purification/renaturing procedures and considerations are well described in the art, and details for developing a suitable renaturing protocol readily can be determined by one having ordinary skill in the art. One useful text one the subject is Guide to Protein Purification, M. 15 Deutscher, ed., Academic Press, San Diego, 1990, particularly section V. Complex formation also may be aided by addition of one or more chaperone proteins.

4.3 Stability of Soluble Morphogen Complexes

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The stability of the highly purified soluble morphogen complex in a physiological buffer, e.g., tris-buffered saline (TBS) and phosphate-buffered saline (PBS), can be enhanced by any of a number of means. Currently preferred is by means of a pro region that comprises at least the first 18 amino acids of the pro sequence (e.g., residues 30-47 of Seq. ID NO. 16 for OP-1), and preferably is the full length pro region. Residues 30-47 show sequence homology to the N-terminal portion of other morphogens and are believed to have particular utility in enhancing complex stability for all morphogens. Other useful means for enhancing the stability of soluble morphogen complexes include three classes of additives. These additives include basic amino acids (e.g., L-arginine, lysine and betaine); nonionic detergents (e.g., Tween 80 or NonIdet P-120); and

carrier proteins (e.g., serum albumin and casein). Useful concentrations of these additives include 1-100 mM, preferably 10-70 mM, including 50 mM, basic amino acid;, 0.01-1.0%, preferably 0.05-0.2%, including 0.1% (v/v) nonionic detergent;, and 0.01-1.0%, preferably 0.05-0.2%, including 0.1% (w/v) carrier protein.

The invention may be embodied in other specific forms without departing from the spirit or essential

10 characteristics thereof. The present embodiments are therefore to be considered in all respects as illustrative and not restrictive, the scope of the invention being indicated by the appended claims rather than by the foregoing description, and all changes which come within the meaning and range of equivalency of the claims are therefore intended to be embraced therein.

- 73 -

SEQUENCE LISTING

(1) GENERAL INFORMATION: 5 (i) APPLICANT: (A) NAME: CREATIVE BIOMOLECULES, INC. (B) STREET: 45 SOUTH STREET (C) CITY: HOPKINTON 10 (D) STATE: HA (E) COUNTRY: USA (F) POSTAL CODE (ZIP): 01748 (G) TELEPHONE: 1-508-435-9001 (H) TELEFAX: 1-508-435-0454 15 (I) TELEX: (ii) TITLE OF INVENTION: MORPHOGEN-INDUCED PERIODONTAL TISSUE REGENERATION 20 (iii) NUMBER OF SEQUENCES: 33 (iv) CORRESPONDENCE ADDRESS: (A) ADDRESSEE: CREATIVE BIOHOLECULES, INC. (B) STREET: 45 SOUTH STREET (C) CITY: HOPKINTON 25 (D) STATE: MA (E) COUNTRY: USA (F) ZIP: 01748 (V) COMPUTER READABLE FORM: 30 (A) MEDIUM TYPE: Floppy disk (B) COMPUTER: IBM PC compatible (C) OPERATING SYSTEM: PC-DOS/MS-DOS (D) SOFTWARE: PatentIn Release #1.0, Version #1.25 35 (vi) CURRENT APPLICATION DATA: (A) APPLICATION NUMBER: (B) FILING DATE: (C) CLASSIFICATION: 40 (vii) PRIOR APPLICATION DATA: (A) APPLICATION NUMBER: (B) FILING DATE: (viii) ATTORNEY/AGENT INFORMATION: 45 (A) NAME: KELLEY ESQ, ROBIN D. (B) REGISTRATION NUMBER: 34,637 (C) REFERENCE/DOCKET NUMBER: CRP-067 50 (ix) TELECOMMUNICATION INFORMATION: (A) TELEPHONE: 617/248-7477 (B) TELEFAX: 617/248-7100

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(2) INFORMATION FOR SEQ ID NO:1:

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(A) LENGTH: 97 amino acids

- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- 10 (ii) MOLECULE TYPE: protein
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 - (A) NAME/KEY: Protein
- 15 (B) LOCATION: 1..97
 - (D) OTHER INFORMATION: /label= GENERIC-SEQ1
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 ONE OF THE 20 NATURALLY-OCCURING L-ISOMER, A-AMINO
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- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

Xaa

- 45 (2) INFORMATION FOR SEQ ID NO:2:
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 - (B) TYPE: amino acid
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 - (ii) MOLECULE TYPE: protein

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Asp Thr Val Pro Lys Pro Cys Cys Ala Pro Thr Gln Leu Asn Ala Ile 105 Ser Val Leu Tyr Phe Asp Asp Ser Ser Asn Val Ile Leu Lys Lys Tyr 5 Arg Asn Met Val Val Arg Ala Cys Gly Cys His 10 (2) INFORMATION FOR SEQ ID NO:7: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 139 amino acids (B) TYPE: amino acid 15 (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (vi) ORIGINAL SOURCE: 20 (A) ORGANISM: HOMO SAPIENS (F) TISSUE TYPE: HIPPOCAMPUS (ix) FEATURE: 25 (A) NAME/KEY: Protein (B) LOCATION: 1..139 (D) OTHER INFORMATION: /label= HOP2-MATURE 30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7: Ala Val Arg Pro Leu Arg Arg Gln Pro Lys Lys Ser Asn Glu Leu 35 Pro Gln Ala Asn Arg Leu Pro Gly Ile Phe Asp Asp Val His Gly Ser His Gly Arg Gln Val Cys Arg Arg His Glu Leu Tyr Val Ser Phe Gln 40 Asp Leu Gly Trp Leu Asp Trp Val Ile Ala Pro Gln Gly Tyr Ser Ala Tyr Tyr Cys Glu Gly Glu Cys Ser Phe Pro Leu Asp Ser Cys Met Asn 45 Ala Thr Asn His Ala Ile Leu Gln Ser Leu Val His Leu Met Lys Pro 50 Asn Ala Val Pro Lys Ala Cys Cys Ala Pro Thr Lys Leu Ser Ala Thr

Ser Val Leu Tyr Tyr Asp Ser Ser Asn Asn Val Ile Leu Arg Lys His Arg Asn Met Val Val Lys Ala Cys Gly Cys His 5 135 (2) INFORMATION FOR SEQ ID NO:8: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 139 amino acids
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(C) STRANDEDNESS: single 10 (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein 15 (vi) ORIGINAL SOURCE: (A) ORGANISM: MURIDAE (F) TISSUE TYPE: EMBRYO 20 (ix) FEATURE: (A) NAME/KEY: Protein (B) LOCATION: 1..139 (D) OTHER INFORMATION: /label= MOP2-MATURE 25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8: Ala Ala Arg Pro Leu Lys Arg Arg Gln Pro Lys Lys Thr Asn Glu Leu 30 Pro His Pro Asn Lys Leu Pro Gly Ile Phe Asp Asp Gly His Gly Ser 35 Arg Gly Arg Glu Val Cys Arg Arg His Glu Leu Tyr Val Ser Phe Arg Asp Leu Gly Trp Leu Asp Trp Val Ile Ala Pro Gln Gly Tyr Ser Ala 40 Tyr Tyr Cys Glu Gly Glu Cys Ala Phe Pro Leu Asp Ser Cys Het Asn Ala Thr Asn His Ala Ile Leu Gln Ser Leu Val His Leu Met Lys Pro 45 Asp Val Val Pro Lys Ala Cys Cys Ala Pro Thr Lys Leu Ser Ala Thr 50 Ser Val Leu Tyr Tyr Asp Ser Ser Asn Asn Val Ile Leu Arg Lys His

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- (C) STRANDEDNESS: single
- 10 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (vi) ORIGINAL SOURCE:
- 15 (A) ORGANISM: bovinae
 - (ix) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1..101
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 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:
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- 30
 Glu Cys Pro Phe Pro Leu Ala Asp His Leu Asn Ser Thr Asn His Ala
 35
 40
 45
- Ile Val Gln Thr Leu Val Asn Ser Val Asn Ser Lys Ile Pro Lys Ala
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 - Cys Cys Val Pro Thr Glu Leu Ser Ala Ile Ser Met Leu Tyr Leu Asp 65 70 75 80
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 - (A) LENGTH: 101 amino acids
 - (B) TYPE: amino acid
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 - (D) TOPOLOGY: linear

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	(vi)		INAL ORG				PHIL	A ME	LANO	GAST	ER					
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	Val	Gly	Cys	Gly 100	Cys ·	Arg										
25	(2) INFO	RMAT	ION 1	FOR S	SEQ J	ED NO	:12:	:								
30	(i)	(A (B (C	UENCI) LEI) TYI) STI) TOI	NGTH: PE: & RANDI	: 102 mino EDNES	ami aci S: s	no a d ingl	cids	i							
	(ii)	HOL	ECULE	TYP	e: p	rote	in									
35	(vi)		GINAI) ORC				US									
40	(ix)	(A) (B)	TURE: NAM LOC OTH	E/KE	N: 1	10	2	/lab	el=	VGL-	FX					
45	(xi)	SEQU	JENCE	DES	CRIP	TION	: SE	Q ID	NO:	12:						•
	Cys 1	Lys	Lys	Arg	His 5	Leu	Tyr	Val	Glu	Phe 10	Lys	Asp	Val	Gly	Trp 15	Gln
50	Asn	Trp	Val	Ile 20	Ala	Pro	Gln	_	Tyr 25	Met .	Ala .	Asn	Tyr	Cys 30	Tyr	Gly
	Glu	Cys	Pro 35	Tyr	Pro	Leu		Glu 40	Ile	Leu	Asn	-	Ser 45	Asn	His	Ala

	TTE	50	GIN	inr	reu	Val	55	SET	116	Giu	FIU	60	vsħ	116	110	LDEU
5	Pro 65	Cys	Cys	Val	Pro	Thr 70	Lys	Het	Ser	Pro	Ile 75	Ser	Het	Leu	Phe	Tyr 80
10	Asp	Asn	Asn	Asp	Asn 85	Val	Val	Leu	Arg	His 90	Tyr	Glu	Asn	Het	Ala 95	Val
10	Asp	Glu	Cys	Gly 100	Cys	Arg										
15	(2) INFO	RMAT:	ION I	FOR :	SEQ :	ID NO): 13:	:								
10	(i)	(B) LEI) TY	NGTH PE:	: 102 amin	am:	ino a Id	acid	s		,					
20		•	.	RANDI POLO			_	Le							,	
	(ii)	HOL	ECULI	E TYI	PE: 1	prote	ein									
25	(vi)			L SOU			DAE	•	. •							
30	(ix)	(A) (B)) NAI) LO	HE/KI CATIO HER I	ON: 1	110)2	/lal	el=	VGR-	-1-F2	2				
	(xi)	SEQ	JENCI	E DES	SCRIE	OITS	l: SI	EQ II	NO:	:13:						
35	Cys 1	Lys	Lys	His	Glu 5	Leu	Tyr	Val	Ser	Phe 10	Gln	Asp	Val	Gly	Trp 15	Gln
40	Asp	Trp	Ile	Ile 20	Ala	Pro	Lys	Gly	Tyr 25	Ala	Ala	Asn	Tyr	Cys 30	Asp	Gly
40	Glu	Cys	Ser 35	Phe	Pro	Leu	Asn	Ala 40	His	Het	Asn	Ala	Thr 45	Asn	His	Ala
45	Ile	Val 50	Gln	Thr	Leu	Val	His 55	Val	Met	Asn	Pro	Glu 60	Tyr	Val	Pro	Lys
	Pro 65	Cys	Cys	Ala	Pro	Thr 70	Lys	Val	Asn	Ala	Ile 75	Ser	Val	Leu		Phe 80
50	Asp	Asp	Asn	Ser	Asn 85	Val	Ile	Leu	Lys	Lys 90	Tyr	Arg	Asn	Het	Val 95	Val

Arg Ala Cys Gly Cys His 100

(2) INFORMATION FOR SEQ ID NO:14: 5 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 106 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single 10 (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (iii) HYPOTHETICAL: NO 15 (iv) ANTI-SENSE: NO (vi) ORIGINAL SOURCE: (A) ORGANISM: Homo sapiens 20 (F) TISSUE TYPE: brain (ix) FEATURE: (A) NAME/KEY: Protein (B) LOCATION: 1..106 (D) OTHER INFORMATION: /note= "GDF-1 (fx)" 25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14: Cys Arg Ala Arg Arg Leu Tyr Val Ser Phe Arg Glu Val Gly Trp His 30 Arg Trp Val Ile Ala Pro Arg Gly Phe Leu Ala Asn Tyr Cys Gln Gly 35 Gln Cys Ala Leu Pro Val Ala Leu Ser Gly Ser Gly Gly Pro Pro Ala Leu Asn His Ala Val Leu Arg Ala Leu Met His Ala Ala Ala Pro Gly 40 Ala Ala Asp Leu Pro Cys Cys Val Pro Ala Arg Leu Ser Pro Ile Ser 45 Val Leu Phe Phe Asp Asn Ser Asp Asn Val Val Leu Arg Gln Tyr Glu Asp Met Val Val Asp Glu Cys Gly Cys Arg 105

(2) INFORMATION FOR SEQ ID NO:15:

50

(i) SEQUENCE CHARACTERISTICS:

5		(A) LENGTH: 5 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
3	(ii)	MOLECULE TYPE: peptide	
10	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:15:	•
	Cys 1	Xaa Xaa Xaa 5	
15	(2) INFOR	MATION FOR SEQ ID NO:16:	
20	(i) S	SEQUENCE CHARACTERISTICS: (A) LENGTH: 1822 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(ii) ł	MOLECULE TYPE: cDNA	
25	(iii) H	IYPOTHETICAL: NO	
	(iv) A	NTI-SENSE: NO	
30	(vi) (ORIGINAL SOURCE: (A) ORGANISH: HOHO SAPIENS (F) TISSUE TYPE: HIPPOCAMPUS	
35	(ix) F	PEATURE: (A) NAME/KEY: CDS (B) LOCATION: 491341 (C) IDENTIFICATION HETHOD: experimental (D) OTHER INFORMATION: /function= "OSTEOGENIC PROTEIN" /product= "OP1"	
40		/evidence= EXPERIMENTAL /standard_name= "OP1"	
	(xi) S	EQUENCE DESCRIPTION: SEQ ID NO:16:	
45	GGTGCGGGCC	CGGAGCCCGG AGCCCGGGTA GCGCGTAGAG CCGGCGCG ATG CAC GTG Het His Val	57
50		G CGA GCT GCG GCG CCG CAC AGC TTC GTG GCG CTC TGG GCA u Arg Ala Ala Pro His Ser Phe Val Ala Leu Trp Ala 10	105

	CCC Pro 20	Leu	TTC Phe	CTG Leu	CTG Leu	CGC Arg 25	TCC Ser	GCC Ala	CTG Leu	GCC Ala	GAC Asp 30	TTC Phe	AGC Ser	CTG Leu	GAC Asp	AAC Asn 35	153
5	GAG Glu	GTG Val	CAC His	TCG Ser	AGC Ser 40	TTC Phe	ATC Ile	CAC His	CGG Arg	CGC Arg 45	CTC Leu	CGC Arg	AGC Ser	CAG Gln	GAG Glu 50	CGG Arg	201
10	CGG Arg	GAG Glu	ATG Met	CAG Gln 55	CGC Arg	GAG Glu	ATC Ile	CTC Leu	TCC Ser 60	ATT Ile	TTG Leu	GGC Gly	TTG Leu	CCC Pro 65	CAC His	CGC Arg	249
15	CCG Pro	CGC Arg	CCG Pro 70	CAC His	CTC Leu	CAG Gln	GGC Gly	AAG Lys 75	CAC His	AAC Asn	TCG Ser	GCA Ala	CCC Pro 80	ATG Met	TTC Phe	ATG Het	297
20	CTG Leu	GAC Asp 85	CTG Leu	TAC Tyr	AAC Asn	GCC Ala	ATG Het 90	GCG Ala	GTG Val	GAG Glu	GAG Glu	GGC Gly 95	GGC Gly	GGG Gly	CCC Pro	GGC Gly	345
20	GGC Gly 100	CAG Gln	GGC Gly	TTC Phe	TCC Ser	TAC Tyr 105	CCC Pro	TAC Tyr	AAG Lys	GCC Ala	GTC Val 110	TTC Phe	AGT Ser	ACC	CAG Gln	GGC Gly 115	393
25	CCC Pro	CCT Pro	CTG Leu	GCC Ala	AGC Ser 120	CTG Leu	CAA Gln	GAT Asp	AGC Ser	CAT His 125	TTC Phe	CTC Leu	ACC Thr	GAC Asp	GCC Ala 130	GAC Asp	441
30	ATG Het	GTC Val	ATG Met	AGC Ser 135	TTC Phe	GTC Val	AAC Asn	CTC Leu	GTG Val 140	GAA Glu	CAT His	GAC Asp	AAG Lys	GAA Glu 145	TTC Phe	TTC Phe	489
35	CAC His	CCA Pro	CGC Arg 150	TAC Tyr	CAC His	CAT His	CGA Arg	GAG Glu 155	TTC Phe	CGG Arg	TTT Phe	GAT Asp	CTT Leu 160	TCC Ser	AAG Lys	ATC Ile	537
40				GAA Glu													585
40	Tvr	Ile	Arg	GAA Glu	Arg	Phe	Asp	Asn	Glu	Thr	Phe	Arg	Ile	Ser	Val	Tyr	633
45	CAG Gln	GTG Val	CTC Leu	CAG Gln	GAG Glu 200	CAC His	TTG Leu	GGC Gly	AGG Arg	GAA Glu 205	TCG Ser	GAT Asp	CTC Leu	TTC Phe	CTG Leu 210	CTC Leu	681
50	GAC Asp	AGC Ser	CGT Arg	ACC Thr 215	CTC Leu	TGG Trp	GCC Ala	Ser	GAG Glu 220	GAG Glu	GGC Gly	TGG Trp	Leu	GTG Val 225	TTT Phe	GAC Asp	729

				Thr								CCG Pro					777
5	GGC Gly	CTG Leu 245	CAG Gln	CTC Leu	TCG Ser	GTG Val	GAG Glu 250	ACG Thr	CTG Leu	GAT Asp	GGG Gly	CAG Gln 255	AGC Ser	ATC Ile	AAC Asn	CCC Pro	825
10	AAG Lys 260	TTG Leu	GCG Ala	GGC Gly	CTG Leu	ATT Ile 265	GGG Gly	CGG Arg	CAC His	GGG Gly	CCC Pro 270	CAG Gln	AAC Asn	AAG Lys	CAG Gln	CCC Pro 275	873
15												CAC His					921
20												CGC Arg					969
20												GCA Ala					1017
25	AGC Ser											CTG Leu 335					1065
30												CCT Pro					1113
35												CTG Leu					1161
40												GTC Val					1209
												ACG Thr					1257
45	ATC Ile											GTC Val 415					1305
50						GTC Val 425						CAC His	TAGO	TCCI	CC		1351
	GAGA	ATTC	AG A	CCCI	TTGG	G GC	CAAG	TTTI	TCT	GGA1	CCT	CCAT	TGCI	CG C	CTTG	GCCAG	1411

	GAACCAGCAG ACCAACTGCC TTTTGTGAGA CCTTCCCCTC CCTATCCCCA ACTTTAAAGG	1471
_	TGTGAGAGTA TTAGGAAACA TGAGCAGCAT ATGGCTTTTG ATCAGTTTTT CAGTGGCAGC	1531
5	ATCCAATGAA CAAGATCCTA CAAGCTGTGC AGGCAAAACC TAGCAGGAAA AAAAAACAAC	1591
	GCATAAAGAA AAATGGCCGG GCCAGGTCAT TGGCTGGGAA GTCTCAGCCA TGCACGGACT	1651
10	CGTTTCCAGA GGTAATTATG AGCGCCTACC AGCCAGGCCA CCCAGCCGTG GGAGGAAGGG	1711
	GGCGTGGCAA GGGGTGGGCA CATTGGTGTC TGTGCGAAAG GAAAATTGAC CCGGAAGTTC	1771
15	CTGTAATAAA TGTCACAATA AAACGAATGA ATGAAAAAAA AAAAAAAAA A	1822
	(2) INFORMATION FOR SEQ ID NO:17:	
20	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 431 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear	·
25	(ii) MOLECULE TYPE: protein	
23	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:	
	Met His Val Arg Ser Leu Arg Ala Ala Ala Pro His Ser Phe Val Ala 1 5 10 15	
30	Leu Trp Ala Pro Leu Phe Leu Leu Arg Ser Ala Leu Ala Asp Phe Ser 20 25 30	
35	Leu Asp Asn Glu Val His Ser Ser Phe Ile His Arg Arg Leu Arg Ser 35 40 45	
	Gln Glu Arg Arg Glu Met Gln Arg Glu Ile Leu Ser Ile Leu Gly Leu 50 60	
40	Pro His Arg Pro Arg Pro His Leu Gln Gly Lys His Asn Ser Ala Pro 65 70 75 80	
	Net Phe Net Leu Asp Leu Tyr Asn Ala Net Ala Val Glu Glu Gly Gly 85 90 95	
45	Gly Pro Gly Gly Gln Gly Phe Ser Tyr Pro Tyr Lys Ala Val Phe Ser 100 105 110	
50	Thr Gln Gly Pro Pro Leu Ala Ser Leu Gln Asp Ser His Phe Leu Thr 115 120 125	
	Asp Ala Asp Met Val Met Ser Phe Val Asn Leu Val Glu His Asp Lys 130 140	

	Glu 145		Phe	His	Pro	Arg 150	Tyr	His	His	Arg	Glu 155	Phe	Arg	Phe	Asp	Leu 160
5	Ser	Lys	Ile	Pro	Glu 165	Gly	Glu	Ala	Val	Thr 170	Ala	Ala	Glu	Phe	Arg 175	Ile
10	Tyr	Lys	Asp	Tyr 180	Ile	Arg	Glu	Arg	Phe 185	Asp	Asn	Glu	Thr	Phe 190	Arg	Ile
10	Ser	Val	Tyr 195	Gln	Val	Leu	Gln	Glu 200	His	Leu	Gly	Arg	Glu 205	Ser	Asp	Leu
15	Phe	Leu 210	Leu	Asp	Ser	Arg	Thr 215	Leu	Trp	Ala	Ser	Glu 220	Glu	Gly	Trp	Leu
	Val 225	Phe	Asp	Ile	Thr	Ala 230	Thr	Ser	Asn	His	Trp 235	Val	Val	Asn	Pro	Arg 240
20	His	Asn	Leu	Gly	Leu 245	Gln	Leu	Ser	Val	Glu 250	Thr	Leu	Asp	Gly	Gln 255	Ser
25	Ile	Asn		Lys 260	Leu	Ala	Gly	Leu	Ile 265	Gly	Arg	His	Gly	Pro 270	Gln	Asn
27	Lys	Gln	Pro 275	Phe	Het	Val	Ala	Phe 280	Phe	Lys	Ala	Thr	Glu 285	Val	His	Phe
30	Arg	Ser 290	Ile	Arg	Ser	Thr	Gly 295	Ser	Lys	Gln	Arg	Ser 300	Gln	Asn	Arg	Ser
	Lys 305	Thr	Pro	Lys	Asn	Gln 310	Glu	Ala	Leu	Arg	Met 315	Ala	Asn	Val	Ala	Glu 320
35	Asn	Ser	Ser	Ser	Asp 325	Gln	Arg	Gln	Ala	Cys 330	Lys	Lys	His	Glu	Leu 335	Tyr
40	Val	Ser	Phe	Arg 340	Asp	Leu	Gly	Trp	Gln 345	Asp	Trp	Ile	Ile	Ala 350	Pro	Glu
40	Gly	Tyr	Ala 355	Ala	Tyr	Tyr	Cys	Glu 360	Gly	Glu	Cys	Ala	Phe 365	Pro	Leu	Asn
45	Ser	Tyr 370	Met	Asn	Ala	Thr	Asn 375	His	Ala	Ile	Val	Gln 380	Thr	Leu	Val	His
	Phe 385	Ile	Asn	Pro	Glu	Thr 390	Val	Pro	Lys	Pro	Cys 395	Cys	Ala	Pro	Thr	Gln 400
50	Leu	Asn	Ala	Ile	Ser 405	Val	Leu	Tyr	Phe	Asp 410	Asp	Ser	Ser	Asn	Val 415	Ile

	Leu Lys Lys Tyr Arg Asn Het Val Val Arg Ala Cys Gly Cys His 420 425 430	
5	(2) INFORMATION FOR SEQ ID NO:18:	
	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1873 base pairs (B) TYPE: nucleic acid	
10	(C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: cDNA	
15	(111) HYPOTHETICAL: NO	
13	(iv) ANTI-SENSE: NO	
20	(vi) ORIGINAL SOURCE: (A) ORGANISH: MURIDAE (F) TISSUE TYPE: EMBRYO	
05	(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1041393	
25	(D) OTHER INFORMATION: /function= "OSTEOGENIC PROTEIN" /product= "MOP1" /note= "MOP1 (CDNA)"	
30	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:	
•	CTGCAGCAAG TGACCTCGGG TCGTGGACCG CTGCCCTGCC	60
35	CGGCGCGGGC CCGGTGCCCC GGATCGCGCG TAGAGCCGGC GCG ATG CAC GTG CGC Het His Val Arg 1	115
40	TCG CTG CGC GCT GCG GCG CCA CAC AGC TTC GTG GCG CTC TGG GCG CCT Ser Leu Arg Ala Ala Ala Pro His Ser Phe Val Ala Leu Trp Ala Pro 5 10 15 20	163
15	CTG TTC TTG CTG CGC TCC GCC CTG GCC GAT TTC AGC CTG GAC AAC GAG Leu Phe Leu Leu Arg Ser Ala Leu Ala Asp Phe Ser Leu Asp Asn Glu 25 30 35	211
.	GTG CAC TCC AGC TTC ATC CAC CGG CGC CTC CGC AGC CAG GAG CGG CGG Val His Ser Ser Phe Ile His Arg Arg Leu Arg Ser Gln Glu Arg Arg 40 45 50	259
50	GAG ATG CAG CGG GAG ATC CTG TCC ATC TTA GGG TTG CCC CAT CGC CCG Glu Met Gln Arg Glu Ile Leu Ser Ile Leu Gly Leu Pro His Arg Pro 55 60 65	307

			His												TTG	355
5		Leu													CAG Gln 100	403
10															CCT	451
15															GTC Val	499
20															CCT Pro	547
															GAG Glu	595
25					ACC Thr											643
30					GAC Asp 185											691
35					TCA Ser			Glu								739
10					GCT Ala											787
20			Ser	Asn	CAC His	Trp	Val	Val	Asn	Pro	Arg	His	Asn			835
15	CAG Gln 245				GAG Glu											883
50	GCA Ala				GGA Gly 265				Pro							931

	GTG Val	GCC Ala	TTC Phe	TTC Phe 280	AAG Lys	GCC Ala	ACG Thr	GAA Glu	GTC Val 285	CAT His	CTC Leu	CGT Arg	AGT Ser	ATC Ile 290	CGG Arg	TCC Ser		979
5	ACG Thr	GGG Gly	GGC Gly 295	AAG Lys	CAG Gln	CGC Arg	AGC Ser	CAG Gln 300	AAT Asn	CGC Arg	TCC Ser	AAG Lys	ACG Thr 305	CCA Pro	AAG Lys	AAC Asn		1027
10	CAA Gln	GAG Glu 310	GCC Ala	CTG Leu	AGG Arg	ATG Met	GCC Ala 315	AGT Ser	GTG Val	GCA Ala	GAA Glu	AAC Asn 320	AGC Ser	AGC Ser	AGT Ser	GAC Asp		1075
15	CAG Gln 325	AGG Arg	CAG Gln	GCC Ala	TGC Cys	AAG Lys 330	AAA Lys	CAT His	GAG Glu	CTG Leu	TAC Tyr 335	GTC Val	AGC Ser	TTC Phe	CGA Arg	GAC Asp 340		1123
20			TGG Trp															1171
20	TAC Tyr	TGT Cys	GAG Glu	GGA Gly 360	GAG Glu	TGC Cys	GCC Ala	TTC Phe	CCT Pro 365	CTG Leu	AAC Asn	TCC Ser	TAC Tyr	ATG Het 370	AAC Asn	GCC Ala		1219
25	ACC Thr	AAC Asn	CAC His 375	GCC Ala	ATC Ile	GTC Val	CAG Gln	ACA Thr 380	CTG Leu	GTT Val	CAC His	TTC Phe	ATC Ile 385	AAC Asn	CCA Pro	GAC Asp		1267
30	ACA Thr	GTA Val 390	CCC	AAG Lys	CCC	TGC Cys	TGT Cys 395	GCG Ala	CCC Pro	ACC Thr	CAG Gln	CTC Leu 400	AAC Asn	GCC Ala	ATC Ile	TCT Ser		1315
35			TAC Tyr															1363
			GTG Val								TAGO	CTCTT	CC I	GAGA	CCCI	CG		1413
40	ACCI	TTGC	GG G	GCCA	CACC	T TI	CCAA	ATCI	TCG	ATGI	CTC	ACCA	TCTA	AG I	CTCI	CACT	G	1473
	CCCA	CCTI	GG C	GAGG	AGAA	C AG	ACCA	ACCI	CTC	CTGA	GCC	TTC	CTCA	CC I	CCCA	ACCG	G	1533
45	AAGC	ATGI	AA G	GGTI	CCAG	AA AA	CCTG	AGCG	TGC	AGCA	GCT	GATG	AGCG	cc c	TTTC	CTTC	ľ	1593
	GGCA	.CGTG	AC G	GACA	AGAT	C CI	ACCA	GCTA	CCA	CAGO	AAA:	CGCC	TAAG	AG C	AGGA	AAAA!	r	1653
E0	GTCT	GCCA	GG A	AAGI	GTCC	A GT	GTCC	ACAT	GGC	CCCI	CGC	GCTC	TGAG	TC I	TTGA	.GGAG	r	1713
50	AATC	GCAA	GC C	TCGT	TCAG	C TG	CAGC	AGAA	GGA	AGGG	CTT	AGCC	AGGG	TG G	GCGC	TGGC	G	1773
	TCTG	TGTI	GA A	.GGGA	AACC	A AG	CAGA	AGCC	ACT	'GTAA	TGA	TATG	TCAC	I AA	'AAAA	CCCA'	r	1833

GAATGAAAAA AAAAAAAAA AAAAAAAAA AAAAGAATTC

1873

5 (2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 430 amino acids

(B) TYPE: amino acid

10 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

15

Met His Val Arg Ser Leu Arg Ala Ala Ala Pro His Ser Phe Val Ala

1 5 10 15

Leu Trp Ala Pro Leu Phe Leu Leu Arg Ser Ala Leu Ala Asp Phe Ser 20 25 30

Leu Asp Asn Glu Val His Ser Ser Phe Ile His Arg Arg Leu Arg Ser 35 40 45

25 Gln Glu Arg Glu Het Gln Arg Glu Ile Leu Ser Ile Leu Gly Leu 50 55 . 60

Pro His Arg Pro Arg Pro His Leu Gln Gly Lys His Asn Ser Ala Pro 65 70 75 80

30

Met Phe Met Leu Asp Leu Tyr Asn Ala Met Ala Val Glu Glu Ser Gly
85

90

95

Pro Asp Gly Gln Gly Phe Ser Tyr Pro Tyr Lys Ala Val Phe Ser Thr 35 100 105 110

Gln Gly Pro Pro Leu Ala Ser Leu Gln Asp Ser His Phe Leu Thr Asp 115 120 125

40 Ala Asp Het Val Het Ser Phe Val Asn Leu Val Glu His Asp Lys Glu 130 135 140

Phe Phe His Pro Arg Tyr His His Arg Glu Phe Arg Phe Asp Leu Ser 145 150 155 160

45
Lys Ile Pro Glu Gly Glu Arg Val Thr Ala Ala Glu Phe Arg Ile Tyr
165
170
175

Lys Asp Tyr Ile Arg Glu Arg Phe Asp Asn Glu Thr Phe Gln Ile Thr 50 180 185 190

Val Tyr Gln Val Leu Gln Glu His Ser Gly Arg Glu Ser Asp Leu Phe 195 200 205

	Leu	Leu 210		Ser	Arg	Thr	Ile 215	Trp	Ala	Ser	Glu	Glu 220	Gly	Trp	Leu	Val
5	Phe 225	Asp	Ile	Thr	Ala	Thr 230	Ser	Asn	His	Trp	Val 235	Val	Asn	Pro	Arg	His 240
	Asn	Leu	Gly	Leu	Gln 245	Leu	Ser	Val	Glu	Thr 250	Leu	Asp	Gly	Gln	Ser 255	Ile
10	Asn	Pro	Lys	Leu 260	Ala	Gly	Leu	Ile	Gly 265	Arg	His	Gly	Pro	Gln 270	Asn	Lys
15	Gln	Pro	Phe 275	Met	Val	Ala	Phe	Phe 280	Lys	Ala	Thr	Glu	Val 285	His	Leu	Arg
	Ser	Ile 290	Arg	Ser	Thr	Gly	Gly 295	Lys	Gln	Arg	Ser	Gln 300	Asn	Arg	Ser	Lys
20	Thr. 305	Pro	Lys	Asn	Gln	Glu 310	Ala	Leu	Arg	Met	Ala 315	Ser	Val	Ala	Glu	Asn 320
25	Ser	Ser	Ser	Asp	Gln 325	Arg	Gln	Ala	Cys	Lys 330	Lys	His	Glu	Leu	Tyr 335	Val
23	Ser	Phe	Arg	Asp 340	Leu	Gly	Trp	Gln	Asp 345	Trp	Ile	Ile	Ala	Pro 350	Glu	Gly
30	Tyr	Ala	Ala 355	Tyr	Tyr	Cys	Glu	Gly 360	Glu	Cys	Ala	Phe	Pro 365	Leu	Asn	Ser
	Tyr	Met 370	Asn	Ala	Thr	Asn	His 375	Ala	Ile	Val	Gln	Thr 380	Leu	Val	His	Phe
35	Ile 385	Asn	Pro	Asp	Thr	Val 390	Pro	Lys	Pro	Cys	Cys 395	Ala	Pro	Thr	Gln	Leu 400
10	Asn	Ala	Ile	Ser	Val 405	Leu	Tyr	Phe	Asp	Asp 410	Ser	Ser	Asn	Val	Asp 415	Leu
	Lys	Lys	Tyr	Arg 420	Asn	Het	Val	Val	Arg 425	Ala	Cys	Gly	Cys	His 430		
15	(2)	INFO	RHAT	CION	FOR	SEQ	ID N	0:20):							
		(i)	(A (B) LE	NGTH PE:	ARAC : 17 nucl	23 b eic	ase acid	pair 	s						
50			•	•		EDNE GY:		_	te							

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

	(A) ORGANISM: Homo sapiens (F) TISSUE TYPE: HIPPOCAMPUS	
5 10	<pre>(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 4901696 (D) OTHER INFORMATION: /function= "OSTEOGENIC PROTEIN"</pre>	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:	
15	GGCGCCGGCA GAGCAGGAGT GGCTGGAGGA GCTGTGGTTG GAGCAGGAGG TGGCACGGCA	60
	GGGCTGGAGG GCTCCCTATG AGTGGCGGAG ACGGCCCAGG AGGCGCTGGA GCAACAGCTC	120
20	CCACACCGCA CCAAGCGGTG GCTGCAGGAG CTCGCCCATC GCCCCTGCGC TGCTCGGACC	180
20	GCGGCCACAG CCGGACTGGC GGGTACGGCG GCGACAGAGG CATTGGCCGA GAGTCCCAGT	240
	CCGCAGAGTA GCCCCGGCCT CGAGGCGGTG GCGTCCCGGT CCTCTCCGTC CAGGAGCCAG	300
25	GACAGGTGTC GCGCGGCGGG GCTCCAGGGA CCGCGCCTGA GGCCGGCTGC CCGCCCGTCC	360
	CGCCCCGCCC CGCCGCCGC CGCCCGCCGA GCCCAGCCTC CTTGCCGTCG GGGCGTCCCC	420
30	AGGCCCTGGG TCGGCCGCGG AGCCGATGCG CGCCCGCTGA GCGCCCCAGC TGAGCGCCCC	480
	CGGCCTGCC ATG ACC GCG CTC CCC GGC CCG CTC TGG CTC CTG GGC CTG Met Thr Ala Leu Pro Gly Pro Leu Trp Leu Leu Gly Leu 1 5 10	528
35	GCG CTA TGC GCG CTG GGC GGC GGC CCC GGC CTG CGA CCC CCG CCC Ala Leu Cys Ala Leu Gly Gly Gly Gly Pro Gly Leu Arg Pro Pro Pro 15 20 25	576
40	GGC TGT CCC CAG CGA CGT CTG GGC GCG CGC GAG CGC CGG GAC GTG CAG Gly Cys Pro Gln Arg Arg Leu Gly Ala Arg Glu Arg Arg Asp Val Gln 30 40 45	624
45	CGC GAG ATC CTG GCG GTG CTC GGG CTG CCT GGG CGG C	672
50	GCG CCA CCC GCC GCC TCC CGG CTG CCC GCG TCC GCG CCG C	720
Ju	CTG GAC CTG TAC CAC GCC ATG GCC GGC GAC GAC GAC GAC GAC GGC GCG Leu Asp Leu Tyr His Ala Het Ala Gly Asp Asp Asp Glu Asp Gly Ala 80 85 90	768

5			Glu				Arg					Het			GTT Val	816
J		Het									Gln				TGG Trp 125	864
10						Asp				Pro					GTC Val	912
15															CTC Leu	960
20							ATG Met 165									1008
25							TTT Phe									1056
23							CTG Leu									1104
30							GAC Asp									1152
35							GAT Asp									1200
40							CAG Gln 245									1248
45							ACC Thr									1296
	AGG Arg 270				Lys		AAC Asn		Leu					Arg		1344
	CCA Pro			Phe				Gly					Gln			1392

	CGT Arg	CGG Arg	CAC His	GAG Glu 305	CTC Leu	TAC Tyr	GTC Val	AGC Ser	TTC Phe 310	CAG Gln	GAC Asp	CTC Leu	GGC Gly	TGG Trp 315	CTG Leu	GAC Asp	1440
5	TGG Trp	GTC Val	ATC Ile 320	Ala	CCC Pro	CAA Gln	GGC Gly	TAC Tyr 325	TCG Ser	GCC Ala	TAT Tyr	TAC Tyr	TGT Cys 330	GAG Glu	GGG Gly	GAG Glu	1488
10	TGC Cys	TCC Ser 335	TTC Phe	CCA Pro	CTG Leu	GAC Asp	TCC Ser 340	TGC Cys	ATG Met	AAT Asn	GCC Ala	ACC Thr 345	AAC Asn	CAC His	GCC Ala	ATC Ile	1536
15	CTG Leu 350	CAG Gln	TCC Ser	CTG Leu	GTG Val	CAC His 355	CTG Leu	ATG Het	AAG Lys	CCA Pro	AAC Asn 360	GCA Ala	GTC Val	CCC Pro	AAG Lys	GCG Ala 365	1584
20	TGC Cys	TGT Cys	GCA Ala	CCC Pro	ACC Thr 370	AAG Lys	CTG Leu	AGC Ser	GCC Ala	ACC Thr 375	TCT Ser	GTG Val	CTC Leu	TAC Tyr	TAT Tyr 380	GAC Asp	1632
	AGC Ser	AGC Ser	AAC Asn	AAC Asn 385	GTC Val	ATC Ile	CTG Leu	CGC Arg	AAA Lys 390	GCC Ala	CGC Arg	AAC Asn	Met	GTG Val 395	GTC Val	AAG Lys	1680
25	GCC Ala	TGC Cys	GGC Gly 400	TGC Cys	CAC His	T GA	.GTCA	GCC0	GCC	CAGC	CCT	ACTG	CAG				1723
30	(2)				FOR NCE	-											
35				(A) (B) (D)	LEN TYP TOP	GTH: E: a: OLOG	402 mino Y: 1	ami aci inea	no a d r	cids							
					ULE '		-			ו חד	MO - 2	1.					
40	Met :												Leu A	Ala I	Leu (15	Cys	
45	Ala 1	Leu (Gly (Gly (20	Gly (Gly I	Pro (Gly 1	Leu <i>l</i> 25	Arg I	Pro 1	Pro 1	Pro (30	Cys I	Pro	
	Gln A	Arg A	Arg 1	Leu (Gly A	Ala A	Arg (Slu <i>A</i> 40	Arg A	Arg A	lsp 1	/al (Sln <i>A</i> 45	Arg (Glu 1	[le	
50	Leu A	Ala V 50	al I	Leu (Gly I	Leu I	Pro 0	ly A	Arg F	Pro A	rg H	Pro A	rg A	la E	Pro E	?ro	

	Ala 65		a Se	r Arg	g Leu	70		Ser	: Ala	a Pro	75		Met	t Leu	ı Asp	Leu 80
5	Тут	His	s Ala	a Met	Ala 85		Asp	Asp	Asp	Glu 90		Gly	Ala	Pro	95	Glu
	Arg	, Arg	g Let	1 Gly	_	, Ala	. Asp	Lev	Val 105		Ser	Phe	Va.	Asn 110		Val
10	Glu	L Arg	Asp 115	_	Ala	Leu	Gly	His 120		Glu	Pro	His	Trp 125		Glu	Phe
15	Arg	Phe 130		Leu	Thr	Gln	Ile 135		Ala	Gly	Glu	Ala 140		. Thr	Ala	Ala
15	Glu 145		Arg	Ile	Tyr	Lys 150	Val	Pro	Ser	Ile	His 155	Leu	Leu	Asn	Arg	Thr 160
20	Leu	His	Val	Ser	Met 165	Phe	Gln	Val	Val	Gln 170	Glu	Gln	Ser	Asn	Arg 175	Glu
	Ser	Asp	Leu	Phe 180	Phe	Leu	Asp	Leu	Gln 185	Thr	Leu	Arg	Ala	Gly 190	-	Glu
25	Gly	Trp	Leu 195	Val	Leu	Asp	Val	Thr 200	Ala	Ala	Ser	Asp	Cys 205	Trp	Leu	Leu
20	Lys	Arg 210		Lys	Asp	Leu	Gly 215	Leu	Arg	Leu	Tyr	Val 220	Glu	Thr	Glu	Asp
30	Gly 225	His	Ser	Val	Asp	Pro 230	Gly	Leu	Ala	Gly	Leu 235	Leu	Gly	Gln	Arg	Ala 240
35	Pro	Arg	Ser	Gln	Gln 245	Pro	Phe	Val	Val	Thr 250	Phe	Phe	Arg	Ala	Ser 255	Pro
	Ser	Pro	Ile	Arg 260	Thr	Pro	Arg	Ala	Val 265	Arg	Pro	Leu	Arg	Arg 270	Arg	Gln
40	Pro	Lys	Lys 275	Ser	Asn	Glu	Leu	Pro 280	Gln	Ala	Asn	Arg	Leu 285	Pro	Gly	Ile
45	Phe	Asp 290	Asp	Val	His	Gly	Ser 295	His	Gly	Arg	Gln	Val 300	Cys	Arg	Arg	His
43	Glu 305	Leu	Tyr	Val		Phe 310	Gln	Asp	Leu	Gly	Trp 315	Leu	Asp	Trp	Val	Ile 320
50	Ala	Pro	Gln		Tyr 325	Ser	Ala	Tyr		Cys 330	Glu	Gly	Glu	Cys	Ser 335	Phe
	Pro	Leu	Asp	Ser 340	Cys	Met .	Asn		Thr 345	Asn	His .	Ala	Ile	Leu 350	Gln	Ser

	Leu	Val	His 355		Met	Lys	Pro	Asn 360		Val	Pro	Lys	Ala 365		Cys	Ala		
5	Pro	Thr 370	-	Leu	Ser	Ala	Thr 375	Ser	Val	Leu	Tyr	Tyr 380	Asp	Ser	Ser	Asn		
10	Asn 385	Val	Ile	Leu	Arg	Lys 390	Ala	Arg	, Asn	Met	Val 395		Lys	Ala	Cys	Gly 400		
10	Cys	His																
15	(2)				FOR CE CI													
20		(-,	() () ()	A) LI B) T C) S	ENGTI YPE: TRANI OPOLO	nuc. DEDNI	926 l leic ESS:	acio sin	pai: d	rs								
20		(vi)	OR	IGIN	AL SO	OURCI	E:											
25			Ċ	F) T	RGAN] ISSUI				YO									
30		(ix)	(1 (1	B) L(ME/E CATI CHER /pi	ON: INFO	93	TION:	: /f1 2-PP1		ion=	"OS:	reogi	ENIC	PRO?	rein"		
35	0001	•			CE DE								2000	.cc (CC A	7C A C CCT		<i>e</i> 1
					CCGG											CCAGCT	1	60 13
40	,									Het 1	Ala	Het	Arg	Pro 5	Gly	Pro		
15					GGC Gly												1	L 6 1
. 3	CCG				CAC His												2	209
50	CGC Arg																2	257

											Ala				TCC	305
5					Met					His				Asp	GAC Asp	353
10					CCA Pro										ATG Het	401
15			Val		ATG Het										GAG Glu	449
20					GAA Glu											497
20					GCT Ala 140											545
25					ACA Thr											593
30					AGG Arg											641
35					GAC Asp											689
40					CTG Leu											737
•0			Glu	Thr	GCG Ala 220	Asp	Gly	His	Ser	Met	Asp	Pro	Gly		Gly	785
4 5	CTG Leu															833
50	TTC Phe	Phe					Ser					Pro				881

			Lys	AGG Arg													92:
5		Lys		CCA Pro													97
10	GAG Glu	GTT Val	TGC Cys	CGC Arg	AGG Arg 300	CAT His	GAG Glu	CTC Leu	TAC Tyr	GTC Val 305	AGC Ser	TTC Phe	CGT Arg	GAC Asp	CTT Leu 310	GGC Gly	102
15				TGG Trp 315													1073
20				TGT Cys													112
20				TTG Leu													1169
25				TGC Cys													1217
30				AGC Ser													1265
35				GCC Ala 395					TGAG	GCCC	CG C	CCAG	CATC	C TG	CTTC	TACT	1319
	ACCI	TAC	CAT (CTGGC	CGGC	c co	CTCI	CCAG	AGG	CAGA	AAC	CCTT	CTAT	GT T	ATCA	TAGCT	1379
40	CAGA	CAG	GG (CAATO	GGAG	G CC	CTTC	ACTI	ccc	CTGG	CCA	CTTC	CTGC	TA A	TTAA	CTGGT	1439
40	CTTI	CCCA	GT 1	CCTC	TGTC	C T1	CATG	GGGI	TTC	GGGG	CTA	TCAC	CCCG	cc c	TCTC	CATCC	1499
	TCCI	ACC	CA A	GCAI	AGAC	T GA	ATGO	ACAC	AGC	ATCC	CAG	AGCT	ATGC	TA A	CTGA	GAGGT	1559
45	CTGG	GGT	CAG C	CACTG	AAGG	c cc	ACAT	'GAGG	AAG	ACTG	ATC	CTTG	GCCA	TC C	TCAG	CCCAC	1619
	AATG	GCAA	I TA	CTGG	ATGG	T CT	AAGA	AGGC	CCT	GGAA	TTC	TAAA	CTAG	AT G	ATCT	GGGCT	1679
50	CTCI	GCAC	CA I	TCAT	TGTG	G CA	GTTG	GGAC	ATT	TTTA	GGT	ATAA	CAGA	CA C	ATAC	ACTTA	1739
טט	GATO	AATO	CA I	CGCI	GTAC	T CC	TTGA	AATC	AGA	GCTA	GCT	TGTT.	AGAA	AA A	GAAT	CAGAG	1799
	CCAG	GTAT	'AG C	GGTG	CATG	T CA	TTAA	TCCC	AGC	GCTA	AAG	AGAC.	AGAG.	AC A	GGAG	AATCT	1859

1919

1926

5	GGAATTC															
	(2)	INF	ORMA	TION	FOR	SEQ	ID	NO:2	3:							
10	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 399 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear															
15		(ii) MOLECULE TYPE: protein														
15		(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:														
20	Met 1	Ala	Het	Arg	Pro 5	Gly	Pro	Leu	Trp	Leu 10	Leu	Gly	Leu	Ala	Leu 15	Cys
	Ala	Leu	Gly	Gly 20	Gly	His	Gly	Pro	Arg 25	Pro	Pro	His	Thr	Cys 30	Pro	Gln
25	Arg	Arg	Leu 35	Gly	Ala	Arg	Glu	Arg 40	Arg	Asp	Met	Gln	Arg 45	Glu	Ile	Leu
	Ala	Val 50	Leu	Gly	Leu	Pro	Gly 55	Arg	Pro	Arg	Pro	Arg 60	Ala	Gln	Pro	Ala
30	Ala 65	Ala	Arg	Gln	Pro	Ala 70	Ser	Ala	Pro	Leu	Phe 75	Met	Leu	Asp	Leu	Tyr 80
35	His	Ala	Het	Thr	Asp 85	Asp	Asp	Asp	Gly	Gly 90	Pro	Pro	Gln	Ala	His 95	Leu
	Gly	Arg	Ala	Asp 100	Leu	Val	Met	Ser	Phe 105	Val	Asn	Met	Val	Glu 110	Arg	Asp
40	Arg	Thr	Leu 115	Gly	Tyr	Gln	Glu	Pro 120	His	Trp	Lys	Glu	Phe 125	His	Phe	Asp
	Leu	Thr 130	Gln	Ile	Pro	Ala	Gly 135	Glu	Ala	Val	Thr	Ala 140	Ala	Glu	Phe	Arg
45	Ile 145	Tyr	Lys	Glu	Pro	Ser 150	Thr	His	Pro	Leu	Asn 155	Thr	Thr	Leu	His	Ile 160
50	Ser	Het	Phe	Glu	Val 165	Val	Gln	Glu	His	Ser 170	Asn	Arg	Glu	Ser	Asp 175	Leu
	Phe	Phe	Leu	Asp 180	Leu	Gln	Thr	Leu	Arg 185	Ser	Gly	Asp	Glu	Gly 190	Trp	Leu

WO 94/06399 PCT/US93/08742

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Val Leu Asp Ile Thr Ala Ala Ser Asp Arg Trp Leu Leu Asn His His Lys Asp Leu Gly Leu Arg Leu Tyr Val Glu Thr Ala Asp Gly His Ser 5 Met Asp Pro Gly Leu Ala Gly Leu Leu Gly Arg Gln Ala Pro Arg Ser 10 Arg Gln Pro Phe Met Val Thr Phe Phe Arg Ala Ser Gln Ser Pro Val Arg Ala Pro Arg Ala Ala Arg Pro Leu Lys Arg Arg Gln Pro Lys Lys 15 Thr Asn Glu Leu Pro His Pro Asn Lys Leu Pro Gly Ile Phe Asp Asp Cly His Gly Ser Arg Gly Arg Glu Val Cys Arg Arg His Glu Leu Tyr 20 Val Ser Phe Arg Asp Leu Gly Trp Leu Asp Trp Val Ile Ala Pro Gln Gly Tyr Ser Ala Tyr Tyr Cys Glu Gly Glu Cys Ala Phe Pro Leu Asp Ser Cys Het Asn Ala Thr Asn His Ala Ile Leu Gln Ser Leu Val His 30 Leu Met Lys Pro Asp Val Val Pro Lys Ala Cys Cys Ala Pro Thr Lys Leu Ser Ala Thr Ser Val Leu Tyr Tyr Asp Ser Ser Asn Asn Val Ile 35 Leu Arg Lys His Arg Asn Met Val Val Lys Ala Cys Gly Cys His 390 395

- 40 (2) INFORMATION FOR SEQ ID NO:24:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1368 base pairs
 - (B) TYPE: nucleic acid
- 45 (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA

- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 1..1368

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

		•	•	•													
5	ATG Met 1	Ser		CTG Leu							Val						48
10	CTG Leu	GGA Gly	CTC Leu	GGA Gly 20	ATG Het	GTT Val	CTG Leu	CTC Leu	ATG Met 25	TTC Phe	GTG Val	GCG Ala	ACC Thr	ACG Thr 30	CCG Pro	CCG Pro	96
15	GCC Ala	GTT Val	GAĠ Glu 35	GCC Ala	ACC Thr	CAG Gln	TCG Ser	GGG Gly 40	ATT Ile	TAC Tyr	ATA Ile	GAC Asp	AAC Asn 45	GGC Gly	AAG Lys	GAC Asp	144
				ATG Met													192
20				ATC Ile													240
25				CAC His													288
30				TAC Tyr 100													336
35				GAC Asp													384
40				GAG Glu													432
40				CGG Arg													480
45	AAC Asn			CAC His													528
50	CGC Arg			TTC Phe 180				Asn									576

	AT(G GC	C GAG a Glu 195	ı Lev	CG(C ATO	TAI Tyr	CAG Gln 200	Asr	C GCC	C AAC A Asi	C GAO	G GG(1 Gl ₃ 205	Ly:	G TG	G CTG Leu	624
5	AC(C GC(Ala 21(a Asr	C AGG	GAC Glu	TTC Phe	Thr 215	Ile	ACC	GTA Val	A TAC	GCC Ala 220	ı Ile	GG(C ACC	GGC Gly	672
10	ACC Th: 225	Let	G GGC	CAG Gln	CAC His	Thr 230	Met	GAG Glu	Pro	CTO Lev	S TCC Ser 235	Ser	GTO Val	AA(Asr	C ACC	Thr 240	720
15	GG0 Gly	GAC Asp	TAC Tyr	GTG Val	GGC Gly 245	Trp	TTG Leu	GAG Glu	CTC	AAC Asn 250	Val	ACC Thr	GAG Glu	GGC	CTC Leu 255	CAC His	768
20	GAG Glu	TGG	CTG Leu	GTC Val 260	AAG Lys	TCG Ser	AAG Lys	GAC Asp	AAT Asn 265	CAT	GGC	Ile	TAC	Ile 270	Gly	GCA Ala	816
	CAC His	GCT Ala	GTC Val 275	AAC Asn	CGA Arg	Pro	GAC Asp	CGC Arg 280	GAG Glu	GTG Val	AAG Lys	CTG Leu	GAC Asp 285	GAC Asp	ATT	GGA Gly	864
25	CTG Leu	ATC Ile 290	His	CGC Arg	AAG Lys	GTG Val	GAC Asp 295	GAC Asp	GAG Glu	TTC Phe	CAG Gln	CCC Pro 300	Phe	ATG Net	ATC	GGC Gly	912
30	TTC Phe 305	TTC Phe	CGC Arg	GGA Gly	CCG Pro	GAG Glu 310	CTG Leu	ATC Ile	AAG Lys	GCG Ala	ACG Thr 315	GCC Ala	CAC His	AGC Ser	AGC Ser	CAC His 320	960
35	CAC His	AGG Arg	AGC Ser	AAG Lys	CGA Arg 325	AGC Ser	GCC Ala	AGC Ser	CAT His	CCA Pro 330	CGC Arg	AAG Lys	CGC Arg	AAG Lys	AAG Lys 335	TCG Ser	1008
40	GTG Val	TCG Ser	CCC	AAC Asn 340	AAC Asn	GTG Val	CCG Pro	Leu	CTG Leu 345	GAA Glu	CCG Pro	ATG Met	GAG Glu	AGC Ser 350	ACG Thr	CGC Arg	1056
	AGC Ser	TGC Cys	CAG Gln 355	Met	Gln	ACC Thr	Leu	Tyr	Ile	Asp	Phe	Lys	Asp	Leu	GGC Gly	TGG Trp	1104
45	His	GAC Asp 370	TGG Trp	ATC Ile	ATC Ile	GCA Ala	CCA Pro 375	GAG Glu	GGC Gly	TAT Tyr	GGC Gly	GCC Ala 380	TTC Phe	TAC Tyr	TGC Cys	AGC Ser	1152
50	GGC Gly 385	GAG Glu	TGC Cys	AAT Asn	Phe	CCG Pro 390	CTC . Leu .	AAT (GCG Ala	His	ATG Met 395	AAC Asn	GCC Ala	ACG Thr	Asn	CAT His 400	1200

	GCG Ala	Ile	GTC Val	CAG Gln	ACC Thr 405	CTG Leu	GTC Val	CAC His	CTG Leu	CTG Leu 410	GAG Glu	CCC Pro	AAG Lys	AAG Lys	GTG Val 415	CCC Pro	1248
5			TGC Cys														1296
10			AAC Asn 435														1344
15			TCC Ser					TGA						•			1368
	(2)	INF	ORMA!	rion	FOR	SEQ	ID 1	NO:25	5:								
20			(i) \$	(A) (B)	LEN TYI	NGTH:	45! mine	ERIST 5 ami 5 aci 1 aci	ino a id		5						
25		(:	ii) P	OLEC	CULE	TYPE	2: p1	rotei	in								
		(:	ki) S	EQUE	INCE	DESC	RIP	CION:	SEC	ID	NO:2	25:					
30	Met 1	•	ki) S Gly	_	•								Val	Leu	Ala 15	Ser	
30	1	Ser	•	Leu	Arg 5	Asn	Thr	Ser	Glu	Ala 10	Val	Ala			15		
30 35	1 Leu	Ser	Gly	Leu Gly 20	Arg 5 Met	Asn Val	Thr Leu	Ser Leu	Glu Met 25	Ala 10 Phe	Val Val	Ala Ala	Thr	Thr 30	15 Pro	Pro	
35	1 Leu Ala	Ser Gly Val	Gly Leu Glu	Leu Gly 20 Ala	Arg 5 Met Thr	Asn Val Gln	Thr Leu Ser	Ser Leu Gly 40	Glu Met 25 Ile	Ala 10 Phe Tyr	Val Val Ile	Ala Ala Asp	Thr Asn 45	Thr 30 Gly	15 Pro Lys	Pro Asp	
	1 Leu Ala Gln	Ser Gly Val Thr	Gly Leu Glu 35	Gly 20 Ala Met	Arg 5 Met Thr	Asn Val Gln Arg	Thr Leu Ser Val 55	Leu Gly 40 Leu	Glu Met 25 Ile Ser	Ala 10 Phe Tyr Glu	Val Val Ile Asp	Ala Asp Asp 60	Thr Asn 45 Lys	Thr 30 Gly Leu	Pro Lys Asp	Pro Asp Val	
35	Leu Ala Gln Ser 65	Ser Gly Val Thr 50	Gly Leu Glu 35	Leu Gly 20 Ala Met	Arg 5 Met Thr His	Asn Val Gln Arg Glu 70	Thr Leu Ser Val 55 Phe	Leu Gly 40 Leu	Glu Met 25 Ile Ser Gly	Ala 10 Phe Tyr Glu	Val Val Ile Asp Ala 75	Ala Asp Asp 60 Glu	Thr Asn 45 Lys Arg	Thr 30 Gly Leu Pro	15 Pro Lys Asp	Pro Asp Val His	
35 40	Leu Ala Gln Ser 65 Leu	Ser Gly Val Thr 50 Tyr	Gly Leu Glu 35 Ile Glu Ser	Leu Gly 20 Ala Het Ile	Arg 5 Met Thr His Leu Gln 85	Asn Val Gln Arg Glu 70 Leu	Thr Leu Ser Val 55 Phe	Leu Gly 40 Leu Leu Leu Thr	Glu Met 25 Ile Ser Gly Arg	Ala 10 Phe Tyr Glu Ile Lys 90	Val Ile Asp Ala 75 Ser	Ala Asp Asp 60 Glu Ala	Thr Asn 45 Lys Arg Pro Leu	Thr 30 Gly Leu Pro	15 Pro Lys Asp Thr Phe 95	Pro Asp Val His 80 Leu	

	Asp	130		GIU	ı Asp	GIU	135		GIN	GIN	Lys	140		: 11e	Inr	Asp
5	Leu 145	-	Lys	Arg	, Ala	Ile 150		Glu	Ser	Asp	Ile 155		Het	Thr	Phe	Leu 160
	Asn	Lys	Arg	His	His 165		Val	Asp	Glu	Leu 170		His	Glu	His	Gly 175	Arg
10	Arg	Leu	Trp	Phe 180		Val	Ser	Asn	Val 185		Asn	Asp	Asn	Tyr 190		Val
15	Met	Ala	Glu 195	Leu	Arg	Ile	Tyr	Gln 200	Asn	Ala	Asn	Glu	Gly 205	-	Trp	Leu
1.7	Thr	Ala 210		Arg	Glu	Phe	Thr 215	Ile	Thr	Val	Tyr	Ala 220	Ile	Gly	Thr	Gly
20	Thr 225	Leu	Gly	Gln	His	Thr 230	Het	Glu	Pro	Leu	Ser 235	Ser	Val	Asn	Thr	Thr 240
	Gly	Asp	Tyr	Val	Gly 245	Trp	Leu	Glu	Leu	Asn 250	Val	Thr	Glu	Gly	Leu 255	His
25	Glu	Trp	Leu	Val 260		Ser	Lys	Asp	Asn 265	His	Gly	Ile	Tyr	Ile 270	Gly	Ala
30	His	Ala	Val 275	Asn	Arg	Pro	Asp	Arg 280	Glu	Val	Lys	Leu	Asp 285	Asp	Ile	Gly
,	Leu	Ile 290	His	Arg	Lys	Val	Asp 295	Asp	Glu	Phe	Gln	Pro 300	Phe	Ket	Ile	Gly
35	Phe 305	Phe	Arg	Gly	Pro	Glu 310	Leu	Ile	Lys	Ala	Thr 315	Ala	His	Ser	Ser	His 320
	His	Arg	Ser	Lys	Arg 325	Ser	Ala	Ser	His	Pro 330	Arg	Lys	Arg	Lys	Lys 335	Ser
10	Val	Ser	Pro	Asn 340	Asn	Val	Pro	Leu	Leu 345	Glu	Pro	Met	Glu	Ser 350	Thr	Arg
15	Ser	Cys	Gln 355	Het	Gln	Thr	Leu	Tyr 360	Ile	Asp	Phe	Lys	Asp 365	Leu	Gly	Trp
	His	Asp 370	Trp	Ile	Ile	Ala	Pro 375	Glu	Gly	Tyr	Gly	Ala 380	Phe	Tyr	Cys	Ser
50	Gly 385	Glu	Cys	Asn	Phe	Pro 390	Leu	Asn	Ala	His	Met 395	Asn	Ala	Thr	Asn	His 400
	Ala	Ile	Val	Gln	Thr 405	Leu	Val	His	Leu	Leu 410	Glu	Pro	Lys	Lys	Val 415	Pro

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Lys Pro Cys Cys Ala Pro Thr Arg Leu Gly Ala Leu Pro Val Leu Tyr 420 425 5 His Leu Asn Asp Glu Asn Val Asn Leu Lys Lys Tyr Arg Asn Met Ile 440 Val Lys Ser Cys Gly Cys His 450 (2) INFORMATION FOR SEQ ID NO:26: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 104 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (ix) FEATURE: (A) NAME/KEY: Protein (B) LOCATION: 1..104 (D) OTHER INFORMATION: /note= "BMP3" (xi) SEQUENCE DESCRIPTION: SEQ ID NO:26: Cys Ala Arg Arg Tyr Leu Lys Val Asp Phe Ala Asp Ile Gly Trp Ser Glu Trp Ile Ile Ser Pro Lys Ser Phe Asp Ala Tyr Tyr Cys Ser Gly Ala Cys Gln Phe Pro Het Pro Lys Ser Leu Lys Pro Ser Asn His Ala Thr Ile Gln Ser Ile Val Ala Arg Ala Val Gly Val Val Pro Gly Ile 50

Pro Glu Pro Cys Cys Val Pro Glu Lys Het Ser Ser Leu Ser Ile Leu

Phe Phe Asp Glu Asn Lys Asn Val Val Leu Lys Val Tyr Pro Asn Het

Thr Val Glu Ser Cys Ala Cys Arg 50 100

(2) INFORMATION FOR SEQ ID NO:27:

5		(1)	(B (C) LE 3) TY 3) SI	INGTH PE: RAND	: 10 amin EDNE	TERI 22 am 30 ac SSS: line	ino id sing	acid	s							
		(ii)	HOL	ECUL	E TY	PE:	prot	ein									
10		(∀i)	ORI (A				: номо	SAP	IENS								
15		(ix)	(B) NA) LO	ME/K CATI	ON:	Prot 11 RMAT	02	/no	te=	"BMP	5"					
		(xi)	SEQ	UENC	E DE	SCRI	PTIO	N: S	EQ I	D NO	:27:						
20		Cys 1	Lys	Lys	His	Glu 5	Leu	Tyr	Val	Ser	Phe 10	Arg	Asp	Leu	Gly	Trp 15	Glı
25		Asp	Trp	Ile	Ile 20	Ala	Pro	Glu	Gly	Tyr 25	Ala	Ala	Phe	Tyr	Cys 30	Asp	Gly
2J		Glu	Cys	Ser 35	Phe	Pro	Leu	Asn	Ala 40	His	Met	Asn	Ala	Thr 45	Asn	His	Ala
30		Ile	Val 50	Gln	Thr	Leu	Val	His 55	Leu	Het	Phe	Pro	Asp 60	His	Val	Pro	Lys
		Pro 65	Cys	Cys	Ala	Pro	Thr 70	Lys	Leu	Asn	Ala	Ile 75	Ser	Val	Leu	Tyr	Phe 80
35		Asp	Asp	Ser	Ser	Asn 85	Val	Ile	Leu	Lys	Lys 90	Tyr	Arg	Asn	Met	Val 95	Va]
40		Arg	Ser	Cys	Gly 100	Cys	His										
4 U	(2)	INFO	RMATI	ON I	FOR S	SEQ 1	ED NO	28:	:								
15		(i)	(B) (C)	TYP STF	IGTH: PE: a RANDE	102 mino DNES	TERIS 2 ami 3 aci 5S: s Linea	no a d ingl	cids	•							
50		(ii)	HOLE	CULE	TYP	E: p	rote	in									
- •		(v i)					ОМО	SAPI	ENS								

5		(14)	(B) NA	HE/K CATI HER	ON:	116	02	/no	te=	"BMP	6 "					
		(xi)	SEQ	UENC	E DE	SCRI	PTIO	N: S	EQ I	D NO	:28:						
10		Cys 1	Arg	Lys	His	Glu 5	Leu	Tyr	Val	Ser	Phe 10	Gln	Asp	Leu	Gly	Trp 15	G1
		Asp	Trp	Ile	Ile 20	Ala	Pro	Lys	Gly	Tyr 25	Ala	Ala	Asn	Tyr	Cys 30	Asp	Gl
15		Glu	Cys	Ser 35	Phe	Pro	Leu	Asn	Ala 40	His	Het	Asn	Ala	Thr 45	Asn	His	Al
20		Ile	Val 50	Gln	Thr	Leu	Val	His 55	Leu	Met	Asn	Pro	Glu 60	Tyr	Val	Pro	Ly
20		Pro 65	Cys	Cys	Ala	Pro	Thr 70	Lys	Leu	Asn	Ala	Ile 75	Ser	Val	Leu	Tyr	Pho 80
25		Asp	Asp	Asn	Ser	Asn 85	Val	Ile	Leu	Lys	Lys 90	Tyr	Arg	Trp	Het	Val 95	Va:
		Arg	Ala	Cys	Gly 100	Cys	His										
30	(2)	INFO	RMATI	ON I	FOR S	SEQ 1	D NO	29:	:								
35		(i)	(B)	LEN	CHANGTH:	: 102 umino	ami aci	ino a id		i							
		(ii)	HOLE	CULE	TYP	e: p	rote	ein									
40		(ix)	(A) (B)	NAE LOC	E/KE ATIC	N: 1	10)2	/3 ak	_1	ODA						
45			(u)		FROM	:e= "	WHER ROUP	EIN OF	EACH ONE	OR E	IS ORE	SPEC	CIFIE	D AH	INO	ACID B.2.	S
		(xi)	SEQU	ENCE	DES	CRIP	TION	i: SE	Q ID	NO:	29:						
50		Cys 1	Xaa	Xaa	His	Glu 5	Leu	Tyr	Val	Xaa	Phe 10	Xaa	Asp	Leu	Gly	Trp 15	Xaa

	Asp	Trp	xaa	20 20	Ala	Pro	Xaa	Gly	25	· xaa	АТА	lyr	Туг	30	Glu	GIÀ
5	Glu	ı Cys	Xaa 35	Phe	Pro	Leu	Xaa	Ser 40	Xaa	Het	Asn	Ala	Thr 45	Asn	His	Ala
	Ile	Xaa 50	Gln	Xaa	Leu	Val	His 55	Xaa	Xaa	Xaa	Pro	Xaa 60	Xaa	Val	Pro	Lys
10	Xaa 65	Cys	Cys	Ala	Pro	Thr 70	Xaa	Leu	Xaa	Ala	Xaa 75	Ser	Val	Leu	Tyr	Xa a 80
15	Asp	Xaa	Ser	Xaa	Asn 85	Val	Xaa	Leu	Xaa	Lys 90	Xaa	Arg	Asn	Het	Val 95	Val
1.7	Xaa	Ala	Cys	Gly 100	Cys	His										
20	(2) INFO	RMAT	ION I	FOR S	SEQ J	ID N	0:30	:								
	(i)) LEI	NGTH:	ARACI : 97	amiı	no ac									
25					EDNES Sy: 1			le								
	(ii)	HOLI	ECULI	E TYP	?E: p	rote	ein									
30	(ix)) NAP	IE/KI	EY: F ON: 1											
35		٠,		IER 1 /not FROM	INFOR ce= " I A G	WAT] WHEE ROUL	CON: REIN POF	EACH ONE	I XAI	A IS		PENI	ENT:	LY SE MINO		
40	(xi)	SEQU	JENCE	DES	CRIP	TION	l: SE	Q II	NO:	30:						
	Leu 1	Xaa	Xaa	Xaa	Phe 5	Xaa	Xaa	Xaa	Gly	Trp 10	Xaa	Xaa	Trp	Xaa	Xaa 15	Xaa
15	Pro	Xaa	Xaa	Xaa 20	Xaa	Ala	Xaa	Tyr	Cys 25	Xaa	Gly	Xaa	Cys	Xaa 30	Xaa	Pro
	Xaa	Xaa	Xaa 35	Xaa	Xaa	Xaa	Xaa	Xaa 40	Asn	His	Ala	Xaa	Xaa 45	Xaa	Xaa	Xaa
50	Xaa	Xaa 50	Xaa	Xaa	Xaa :		Xaa 55	Xaa	Xaa	Xaa		Xaa 60	Cys	Cys	Xaa	Pro

Xaa Xaa Xaa Xaa Xaa Xaa Xaa Leu Xaa Xaa Xaa Xaa Xaa Xaa Val Xaa Leu Xaa Xaa Xaa Xaa Xaa Het Xaa Val Xaa Xaa Cys Xaa Cys 5 Xaa 10 (2) INFORMATION FOR SEQ ID NO:31: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 102 amino acids (B) TYPE: amino acid 15 (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein 20 (ix) FEATURE: (A) NAME/KEY: Protein (B) LOCATION: 1..102 (D) OTHER INFORMATION: /label= GENERIC-SEQ6 25 /note= "WHEREIN EACH XAA IS INDEPENDENTLY SELECTED FROM A GROUP OF ONE OR MORE SPECIFIED AMINO ACIDS AS DEFINED IN THE SPECIFICATION. " 30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31: Cys Xaa Xaa Xaa Leu Xaa Xaa Xaa Phe Xaa Xaa Gly Trp Xaa 35 Xaa Trp Xaa Xaa Xaa Pro Xaa Xaa Xaa Xaa Ala Xaa Tyr Cys Xaa Gly Xaa Cys Xaa Xaa Pro Xaa Xaa Xaa Xaa Xaa Xaa Xaa Asn His Ala 40 Xaa Cys Cys Xaa Pro Xaa Xaa Xaa Xaa Xaa Xaa Xaa Leu Xaa Xaa 45 Xaa Xaa Xaa Xaa Xaa Val Xaa Leu Xaa Xaa Xaa Xaa Met Xaa Val 50 Xaa Xaa Cys Xaa Cys Xaa 100 (2) INFORMATION FOR SEQ ID NO:32:

5		(i	· (A) L B) T C) S	CE C ENGT YPE: TRAN OPOL	H: 1 nuc DEDN	247 leic ESS:	base aci sin	pai d	rs								
		(ii) но	LECU	LE T	YPE:	cDN	A										
10		(vi	(.	A) 0	AL S RGAN ISSU	ISM:	HOM	O SA BRAI	PIEN N	S								
15		(ix	(, (,	B) L	AME/1 DCAT: THER	ION:	84. ORMA	TION	: /p:		ct=	"GDF	-1"					
20		(vi	CF	OHEN	CE DI	ECCB.	TPTT	ON•	SEO 1	א מז	n•32	•						
	ccco	•											GCGG.	ACC +	CTGC	GCACTC		60
25	TCTG																	10
•	-0-0														ro C			
30	GGC Gly 10																1	.58
35	CTG Leu	ACC Thr	CGC Arg	GCC Ala	CCC Pro 30	GTG Val	CCC Pro	CCA Pro	GGC Gly	CCA Pro 35	GCC Ala	GCC Ala	GCC Ala	CTG Leu	CTC Leu 40	CAG Gln	2	:06
40	GCT Ala																2	54
40	GTT Val																3	02
45	ACC Thr	AGG Arg 75	TCT Ser	GGC Gly	TCG Ser	CGG Arg	CGG Arg 80	ACG Thr	TCC Ser	CCA Pro	GGG Gly	GTC Val 85	ACC Thr	CTG Leu	CAA Gln	CCG Pro	3	50
50	TGC Cys 90																3	98

	Pro	G GA	C CG(C GG: g Gly	r GCC 7 Ala 110	Pro	ACC Thr	CGC	GCC Ala	Ser 115	r Glu	G CC	r GT(C TC	G GC0 r Ala 120	C GCG A Ala	446
5					Glu					. Phe					a Val	G GAA L Glu	
10	CCC Pro	GCT Ala	GAC Glu 140	ı Arg	CCC Pro	AGC Ser	CGG	GCC Ala 145	Arg	Leu	GAC Glu	CTO Let	G CGT Arg 150	Phe	GCC Ala	G GCG Ala	542
15	GCG Ala	GC0 Ala 155	Ala	GCA Ala	GCC Ala	CCG Pro	GAG Glu 160	Gly	GGC Gly	TGG	GAG Glu	Lev 165	Ser	GTC Val	GCC Ala	CAA Gln	590
20	GCG Ala 170	Gly	CAG Gln	GGC	GCG Ala	GGC Gly 175	GCG Ala	GAC Asp	CCC Pro	GGG Gly	Pro 180	Val	CTG Leu	Leu	CGC Arg	CAG Gln 185	638
	TTG Leu	GTG Val	Pro	GCC Ala	CTG Leu 190	Gly	CCG Pro	CCA Pro	GTG Val	CGC Arg 195	Ala	GAG Glu	CTG Leu	CTG	GGC Gly 200	GCC Ala	686
2 5	GCT Ala	TGG Trp	GCT Ala	CGC Arg 205	AAC Asn	GCC Ala	TCA Ser	TGG Trp	CCG Pro 210	CGC Arg	AGC Ser	CTC Leu	CGC	CTG Leu 215	GCG Ala	CTG Leu	734
30	GCG Ala	CTA Leu	CGC Arg 220	CCC Pro	CGG Arg	GCC Ala	CCT Pro	GCC Ala 225	GCC Ala	TGC Cys	GCG Ala	CGC Arg	CTG Leu 230	GCC Ala	GAG Glu	GCC Ala	782
35	TCG Ser	CTG Leu 235	CTG Leu	CTG Leu	GTG Val	ACC Thr	CTC Leu 240	GAC Asp	CCG Pro	CGC Arg	CTG Leu	TGC Cys 245	CAC His	CCC Pro	CTG Leu	GCC Ala	830
40	CGG Arg 250	CCG Pro	CGG Arg	CGC Arg	GAC Asp	GCC Ala 255	GAA Glu	CCC Pro	GTG Val	TTG Leu	GGC Gly 260	GGC Gly	GGC Gly	CCC Pro	GGG Gly	GGC Gly 265	878
	GCT Ala	TGT Cys	CGC Arg	Ala	CGG Arg 270	Arg	Leu	Tyr	Val	Ser	Phe	Arg	GAG Glu	GTG Val	GGC Gly 280	TGG Trp	926
45	CAC His	CGC Arg	TGG Trp	GTC Val 285	ATC Ile	GCG Ala	CCG Pro	Arg	GGC Gly 290	TTC Phe	CTG Leu	GCC Ala	AAC Asn	TAC Tyr 295	TGC Cys	CAG Gln	974
50	GGT Gly	CAG Gln	TGC Cys 300	GCG Ala	CTG Leu	CCC Pro	Val .	GCG Ala 305	CTG Leu	TCG Ser	GGG Gly	Ser	GGG Gly 310	GGG Gly	CCG Pro	CCG Pro	1022

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			Asn	CAC His									Ala	GCC Ala		
5	GGA Gly 330	Ala	GCC Ala	GAC Asp	CTG Leu	CCC Pro 335	TGC Cys	TGC Cys	GTG Val	CCC Pro	GCG Ala 340	CGC Arg	CTG Leu	TCG Ser	CCC	ATC Ile 345
10				TTC Phe												
15				GTG Val 365								TAA	CCCG	GGG (CGGG	CAGGGA
	CCC	GGGC	CCA A	ACAA!	CAAA'	rg co	CGCG'	rgg								
20	(2)	INF	ORMA!	rion	FOR	SEQ	ID I	10:33	3:							
25			(i) :	(B)	LEI TYI	NGTH: PE: a	: 372 amino	2 ami	ino a id		S					
25		C:	ii) l	•		00109 1977										•
		•	•	SEQUE			-) ID	NO:3	33:				
30	Met 1	(:	ci) S		ence	DESC	CRIPT	CION:	SEC				Leu	Leu	Leu 15	Leu
30 35	1	(: Pro	ci) S Pro	SEQUE	ENCE Gln 5	DES(CRIPT Gly	Pro	SE(Gly 10	His	His			15	
	1 Leu	(? Pro	ri) { Pro Leu	SEQUI Pro Leu	Gln 5 Leu	DESC Gln Pro	Gly Ser	Pro Leu	Cys Pro 25	Gly 10 Leu	His Thr	His Arg	Ala	Pro 30	15 Val	Pro
	Leu Pro	Pro Ala Gly	Pro Leu Pro 35	Pro Leu 20	Gln 5 Leu Ala	DESC Gln Pro Ala	Gly Ser Leu	Pro Leu Leu 40	Cys Pro 25 Gln	Gly 10 Leu Ala	His Thr Leu	His Arg Gly	Ala Leu 45	Pro 30 Arg	15 Val Asp	Pro Glu
35	1 Leu Pro	(: Pro Ala Gly Gln 50	Pro Leu Pro 35	Pro Leu 20	Gln 5 Leu Ala Pro	DESC Gln Pro Ala Arg	Gly Ser Leu 55	Pro Leu Leu 40 Arg	Cys Pro 25 Gln Pro	Gly 10 Leu Ala Val	His Thr Leu Pro	His Arg Gly Pro 60	Ala Leu 45 Val	Pro 30 Arg	15 Val Asp Trp	Pro Glu Arg
35	Leu Pro Pro Leu 65	Pro Ala Gly Gln 50 Phe	Pro Leu Pro 35 Gly Arg	Pro Leu 20 Ala	INCE Gln 5 Leu Ala Pro Arg	DESC Gln Pro Ala Arg Asp 70	Gly Ser Leu Leu 55	Pro Leu Leu 40 Arg	Cys Pro 25 Gln Pro Glu	Gly 10 Leu Ala Val	His Thr Leu Pro Arg 75	His Arg Gly Pro 60 Ser	Ala Leu 45 Val	Pro 30 Arg Het	15 Val Asp Trp Arg	Pro Glu Arg Arg 80
35	Leu Pro Pro Leu 65	Pro Ala Gly Gln 50 Phe	Pro Leu Pro 35 Gly Arg	EQUI Pro Leu 20 Ala Ala	INCE Gln 5 Leu Ala Pro Arg Val 85	DESC Gln Pro Ala Arg Asp 70 Thr	Gly Ser Leu 55 Pro	Pro Leu Leu 40 Arg Gln	Cys Pro 25 Gln Pro Glu Pro	Gly 10 Leu Ala Val Thr	His Thr Leu Pro Arg 75 His	His Arg Gly Pro 60 Ser Val	Ala Leu 45 Val Gly Glu	Pro 30 Arg Met Ser	15 Val Asp Trp Arg	Pro Glu Arg Arg 80

	Val	Val 130	Phe	Asp	Leu	Ser	Ala 135	Val	Glu	Pro	Ala	Glu 140	Arg	Pro	Ser	Ar
5	Ala 145	Arg	Leu	Glu	Leu	Arg 150	Phe	Ala	Ala	Ala	Ala 155	Ala	Ala	Ala	Pro	G11 160
10	Gly	Gly	Trp	Glu	Leu 165	Ser	Val	Ala	Gln	Ala 170	Gly	Gln	Gly	Ala	Gly 175	Ala
10	Asp	Pro	Gly	Pro 180	Val	Leu	Leu	Arg	Gln 185	Leu	Val	Pro	Ala	Leu 190	Gly	Pro
15	Pro	Val	Arg 195	Ala	Glu	Leu	Leu	Gly 200	Ala	Ala	Trp	Ala	Arg 205	Asn	Ala	Ser
	Trp	Pro 210	Arg	Ser	Leu	Arg	Leu 215	Ala	Leu	Ala	Leu	Arg 220	Pro	Arg	Ala	Pro
20	Ala 225	Ala	Cys	Ala	Arg	Leu 230	Ala	Glu	Ala	Ser	Leu 235	Leu	Leu	Val	Thr	Let 240
25	Asp	Pro	Arg	Leu	Cys 245	His	Pro	Leu	Ala	Arg 250	Pro	Arg	Arg	Asp	Ala 255	Glu
	Pro	Val	Leu	Gly 260	Gly	Gly	Pro	Gly	Gly 265	Ala	Cys	Arg	Ala	Arg 270	Arg	Leu
30	Tyr	Val	Ser 275	Phe	Arg	Glu	Val	Gly 280	Trp	His	Arg	Trp	Val 285	Ile	Ala	Pro
	Arg	Gly 290	Phe	Leu	Ala	Asn	Tyr 295	Cys	Gln	Gly	Gln	Cys 300	Ala	Leu	Pro	Val
35	Ala 305	Leu	Ser	Gly	Ser	Gly 310	Gly	Pro	Pro	Ala	Leu 315	Asn	His	Ala	Val	Leu 320
10	Arg	Ala	Leu	Met	His 325	Ala	Ala	Ala	Pro	Gly 330	Ala	Ala	Asp	Leu	Pro 335	Cys
	Cys	Val	Pro	Ala 340	Arg	Leu	Ser	Pro	Ile 345	Ser	Val	Leu	Phe	Phe 350	Asp	Asn
5	Ser	Asp	Asn 355	Val	Val	Leu	Arg	Gln 360	Tyr	Glu	Asp	Het	Val 365	Val	Asp	Glu
	Cys	Gly 370	Cys	Arg												

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What is claimed is:

5 1. A method for enhancing integration of a tooth in a mammalian tooth socket, the method comprising the step of:

providing a therapeutically effective concentration of a morphogen to the tooth socket surface, said concentration being sufficient to induce periodontal tissue morphogenesis in said socket.

- 2. The method of claim 1 wherein said step of providing a therapeutically effective morphogen concentration to said surface comprises the step of administering to said mammal a therapeutically effective concentration of a morphogen.
- 3. The method of claim 1 wherein said step of providing a therapeutically effective morphogen concentration to said surface comprises the step of administering to said mammal an agent that stimulates <u>in vivo</u> a therapeutically effective concentration of an endogenous morphogen.

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4. The method of claim 2 or 3 wherein said morphogen or morphogen-stimulating agent is disposed on the surface of the tooth root prior to implantation of said tooth in said tooth socket.

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5. The method of claim 2 or 3 wherein said morphogen or morphogen-stimulating agent is disposed on the surface of the tooth socket prior to implantation of said tooth in said tooth socket.

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6. The method of claim 4 wherein said tooth root surface is partially demineralized.

- 7. The method of claim 5 wherein said tooth root surface5 is partially demineralized.
 - 8. The method of claim 1 wherein said tooth is an implanted tooth.
- 10 9. The method of claim 1 wherein said tooth is a prosthetic tooth.
 - 10. The method of claim 9 wherein said prosthetic tooth is an allogenic or autologous tooth.

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11. The method of claim 1 wherein said therapeutically effective concentration is sufficient to induce differentiation and proliferation of cementoblasts or periodontoblasts.

- 12. The method of claim 1 wherein said therapeutically effective concentration is sufficient to induce formation of periodontal ligament or cementum.
- 25 13. The method of claim 2 or 3 wherein said morphogen or morphogen stimulating agent is administered to said mammal dispersed in an acellular matrix material.
- 14. The method of claim 13 wherein said matrix material is derived from dentin, periodontal ligament, bone, or cementum tissue.
- 15. A method for regenerating periodontal tissue in a mammalian tooth socket, the method comprising the step of:

providing to the locus of the tooth socket a therapeutically effective concentration of a morphogen sufficient to induce formation of periodontal ligament or cementum.

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- 16. A method for inhibiting the tissue damage associated with periodontal disease, the method comprising the step of:

 providing a therapeutically effective concentration of a morphogen to the periodontal tissue at risk of damage.
- 17. A method for inhibiting periodontal tissue loss in a mammal, the method comprising the step of providing a therapeutically effective concentration of a morphogen to an implanted tooth or tooth socket surface, said concentration being sufficient to induce regeneration of lost or damaged periodontium.
- 20 18. The method of claim 1, 15, 16 or 17 wherein said therapeutic morphogen concentration is less than about $50\mu g$.
- 19. The method of claim 18 wherein said therapeutic morphogen concentration is less than about $25\mu g$.
- 20. The method of claim 1, 15, 16 or 17 wherein said therapeutically effective concentration is sufficient to induce formation of periodontal ligament or cementum.
 - 21. The method of claim 1, 15, 16 or 17 wherein said therapeutic morphogen concentration is sufficient to induce proliferation and differentiation of cementoblasts or periodontoblasts.

22. The method of claim 15, 16 or 17 wherein said morphogen is provided to said tissue by administering to said mammal a therapeutically effective concentration of a morphogen.

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- 23. The method of claim 15, 16 or 17 wherein said morphogen is provided to said tissue by administering to said mammal an agent that stimulates in vivo a therapeutically effective concentration of an endogenous morphogen.
- 24. A method for preparing a tooth for implantation in a mammalian tooth socket, said socket being significantly reduced in viable periodontal tissue, the method comprising the steps of:
 - (a) disposing a therapeutically effective concentration of a morphogen about the exterior surface of a tooth root to be implanted;
 - (b) preparing a tooth socket to receive said tooth; and
- 20 (c) implanting said tooth in said socket.
 - 25. The method of claim 24 comprising the additional step of partially demineralized the tooth root surface before disposing said morphogen on said surface.

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- 26. A method for preparing a tooth socket to receive a tooth, said tooth socket being significantly reduced in viable peridontal tissue, the method comprising the steps of:
- (a) preparing the tooth socket to receive a tooth;
 (b) disposing on the tooth socket surface a therapeutically effective concentration of a morphogen;
 and
 - (c) implanting said tooth in said prepared socket.

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- 27. The method of claim 26 wherein said tooth root surface is partially demineralized before implantation.
- 28. The method of claim 1, 15, 16, 17, 24 or 26 wherein said morphogen comprises an amino acid sequence sharing at least 70% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, Vg1(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).
- The method of claim 28 wherein said morphogen comprises an amino acid sequence sharing a last 80% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, BMP3(fx), BMP5(fx), BMP6(fx), Vg1(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).
- 30. The method of claim 1, 15, 16, 17, 24 or 26 wherein said morphogen comprises an amino acid sequence having greater than 60% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
 - 31. The method of claim 30 wherein said morphogen comprises an amino acid sequence having greater than 65% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
 - 32. The method of claim 31 wherein said morphogen comprises an amino acid sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1), including allelic and species variants thereof.
 - 33. The method of claim 1, 15, 16, 17, 24 or 26 wherein said morphogen comprises an amino acid sequence defined by Generic Sequences 1, 2, 3, 4, 5 or 6 (Seq. ID Nos. 1, 2, 3, 4, 30 or 31).

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- 34. The method of claim 1, 15, 16, 17, 24 or 26 wherein said morphogen comprises an amino acid sequence defined by OPX (Seq. ID No. 29).
- 5 35. A composition for inhibiting periodontal tissue loss in a mammal, said composition comprising a therapeutic concentration of a morphogen in association with a symptom alleviating cofactor.
- 10 36. The composition of claim 35 wherein said therapeutically effective concentration is sufficient to induce periodontal tissue morphogenesis.
- 37. The composition of claim 35 wherein said
 therapeutically effective concentration is sufficient
 to enhance integration of an implanted tooth in a tooth
 socket.
- 38. The composition of claim 35 wherein said cofactor comprises an antibiotic.
 - 39. The composition of claim 35 wherein said cofactor is an antiseptic.
- 25 40. The composition of claim 35 wherein said cofactor comprises an analgesic or anesthetic.
 - 41. The composition of claim 38 wherein said cofactor comprises tetracycline.
 - 42. The composition of claim 35 wherein said morphogen comprises an amino acid sequence sharing at least 70% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, Vg1(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).

- 43. The composition of claim 42 wherein said morphogen comprises an amino acid sequence sharing a last 80% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, BMP3(fx), BMP5(fx), BMP6(fx), Vg1(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).
- 44. The composition of claim 43 wherein said morphogen comprises an amino acid sequence having greater than 60% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
- 45. The composition of claim 44 wherein said morphogen comprises an amino acid sequence having greater than 65% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
- 46. The composition of claim 45 wherein said morphogen comprises an amino acid sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1), including allelic and species variants thereof.
- 47. The composition of claim 46 wherein said morphogen comprises an amino acid sequence defined by Generic Sequences 1, 2, 3, 4, 5 or 6 (Seq. ID Nos. 1, 2, 3, 4, 30 or 31).
- 48. The composition of claim 47 wherein said morphogen comprises an amino acid sequence defined by OPX (Seq. 30 ID No. 29).
 - 49. The composition of claim 35 wherein said morphogen is dispersed in an acellular matrix.

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- 50. The composition of claim 49 wherein said acellular matrix is derived from dentin, bone, periodontal ligament or cementum tissue.
- 5 51. The composition of claim 35 wherein said composition comprises a solution of high viscosity.
 - 52. The method of claim 1, 15, 16 or 17 wherein said morphogen species provided comprises the pro form.

- 53. The method of claim 32 wherein said morphogen species provided comprises the pro form.
- 54. The method of claim 53 wherein said morphogen comprises 15 an amino acid sequence defined by residues 30-431 of Seq. ID No. 16 (hOP1), including allelic and species variants thereof.
- 55. The composition of claim 35 wherein said morphogen species provided comprises the pro form.
 - 56. The composition of claim 46 wherein said morphogen species provided comprises the pro form.
- 25 57. The composition of claim 55 wherein said morphogen comprises an amino acid sequence defined by residues 30-431 of Seq. ID No. 16 (hOP-1), including allelic and species variants thereof.
- 30 58. The method of claim 3 or 23 wherein said agent stimulates expression of a morphogen in a tissue other than periodontal, dentin, or alveolar bone.

59. The use of a morphogen in the manufacture of a pharmaceutical to enhance the integration of a tooth in a tooth socket.

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60. The use of a morphogen in the manufacture of a pharmaceutical to regenerate periodontal tissue or to inhibit periodontal tissue loss or the tissue damage associated with periodontal disease.

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- 61. The use according to 59 or 60 wherein said morphogen comprises an amino acid sequence sharing at least 70% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, Vg1(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).
- 62. The use according to claim 61 wherein said morphogen comprises an amino acid sequence sharing a least 80% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, BMP3(fx), BMP5(fx), BMP6(fx), Vg1(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).
- 63. The use according to claim 59 or 60 wherein said
 25 morphogen comprises an amino acid sequence having
 greater than 60% amino acid identity with the sequence
 defined by residues 43-139 of Seq. ID No. 5 (hOP1).
- 64. The use according to claim 63 wherein said morphogen comprises an amino acid sequence having greater than 65% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).

65. The use according to claim 63 wherein said morphogen comprises an amino acid sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1), including allelic and species variants thereof.

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66. The use according to claim 59 or 60 wherein said morphogen comprises an amino acid sequence defined by Generic Sequences 1, 2, 3, 4, 5 or 6 (Seq. ID Nos. 1, 2, 3, 4, 30 or 31).

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- 67. The use according to claim 59 or 60 wherein said morphogen comprises an amino acid sequence defined by OPX (Seq. ID No. 29).
- 15 68. The invention of claim 1, 15, 16, 17, 35, 59 or 60 wherein said morphogen comprises a polypeptide chain encoded by a nucleic acid that hybridizes under stringent conditions with the DNA sequence defined by nucleotides 1036-1341 of Seq. ID No. 16 or nucleotides 1390-1695 of Seq. ID No. 20.
- 69. The invention of claim 1, 15, 16, 17, 35, 59 or 60 wherein said morphogen comprises a dimeric protein species complexed with a peptide comprising a pro region of a member of the morphogen family, or an allelic, species or other sequence variant thereof.
- 70. The invention of claim 69 wherein said dimeric morphogen species is noncovalently complexed with said peptide.
 - 71. The invention of claim 69 wherein said dimeric morphogen species is complexed with two said peptides.

- 72. The invention of claim 69 wherein said peptide comprises at least the first 18 amino acids of a sequence defining said pro region.
- 5 73. The invention of claim 72 wherein said peptides comprises the full length form of said pro region.
- 74. The invention of claim 69 wherein said peptide comprises a nucleic acid that hybridizes under stringent hybridization conditions with a DNA defined by nucleotides 136-192 of Seq. ID No. 16, or nucleotides 157-211 of Seq. ID No. 20.
- 75. The invention of claim 69 wherein said complex is
 further stabilized by exposure to a basic amino acid, a
 detergent or a carrier protein.

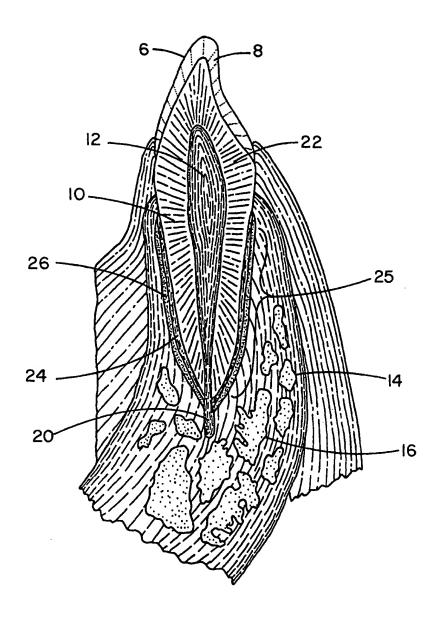
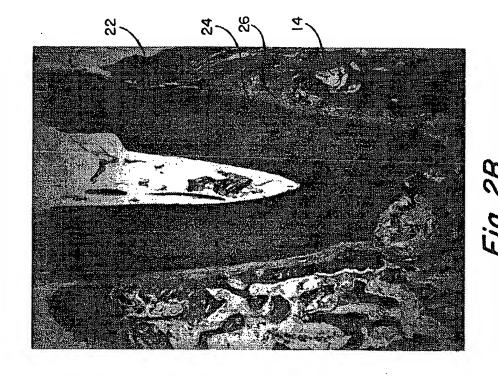
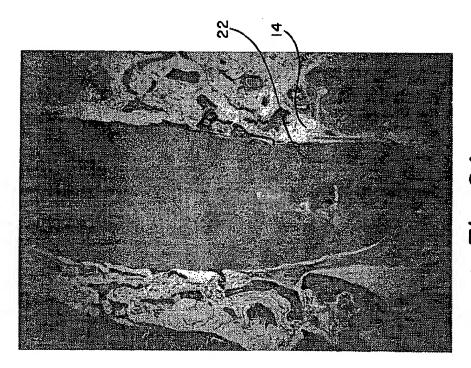


Fig./

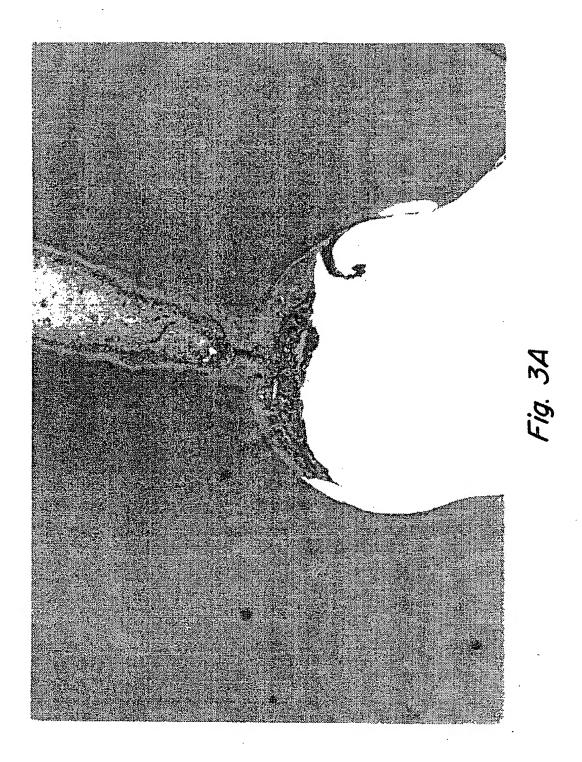
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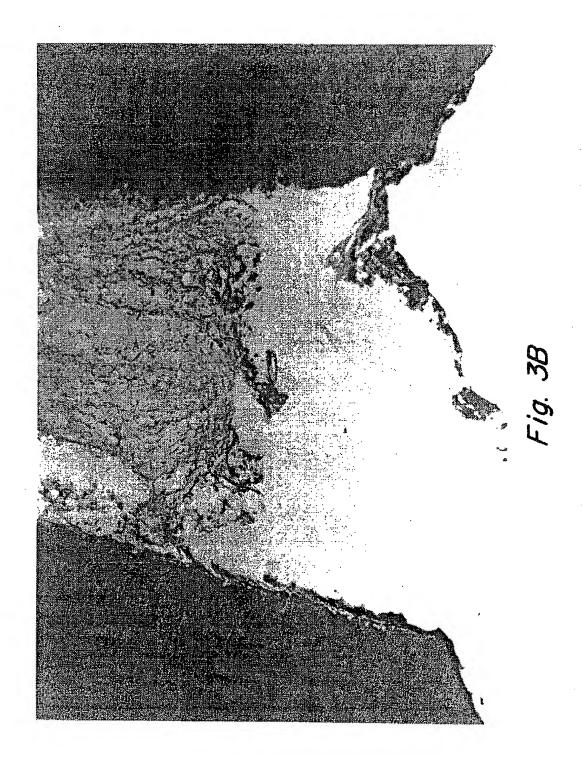


SUBSTITUTE SHEET

Fig. 24



SUBSTITUTE SHEET



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In. ational Application No

PCT/US 93/08742 A. CLASSIFICATION OF SUBJECT MATTER
IPC 5 A61K6/00 A61L27/00 According to International Patent Classification (IPC) or to both national classification and IPC B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) A61K A61L C07K IPC 5 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) C. DOCUMENTS CONSIDERED TO BE RELEVANT Relevant to claim No. Citation of document, with indication, where appropriate, of the relevant passages Category ' 1-75 WO,A,92 15323 (CREATIVE BIOMOLECULES) 17 X,P September 1992 cited in the application see page 7, line 1 - line 8 see page 13, line 5 - page 14, line 11 see claims; tables 1-29, WO,A,88 00205 (GENETICS INSTITUTE) 14 X 35-37, January 1988 49,59-62 see claims; tables see page 9, line 7 see page 9, paragraph 2 see page 10, last paragraph - page 11, paragraph 1 Patent family members are listed in annex. Further documents are listed in the continuation of box C. Special categories of cited documents: "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the "A" document defining the general state of the art which is not considered to be of particular relevance "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such docu-"O" document referring to an oral disclosure, use, exhibition or ments, such combination being obvious to a person skilled document published prior to the international filing date but later than the priority date claimed "&" document member of the same patent family Date of mailing of the international search report Date of the actual completion of the international search 29. 12.93 17 December 1993 Authorized officer Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentiaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016

Form PCT/ISA/210 (second sheet) (July 1992)

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	DOCUMENTS CONCIDENCE TO BE BELLIANT	PC1/03 93	, , , , , ,
C.(Continua Category	citon) DOCUMENTS CONSIDERED TO BE RELEVANT Citation of document, with indication, where appropriate, of the relevant passages		Relevant to claim No.
Y	EP,A,O 495 284 (OSTEOTECH, INC.) 22 July 1992 see claims; examples see column 7, line 25 - column 9, line 42		1-51, 58-67
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International application No.

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Box I	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This inter	national search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
P	Claims Nos.: X ecause they relate to subject matter not required to be searched by this Authority, namely: REMARK: Although claims 1-34,52-54,58 and 68-75(partly) are directed to a
	method of treatment of the human body the search has been carried out and wased on the alleged effects of the composition.
- F	claims Nos.: ecause they relate to parts of the international application that do not comply with the prescribed requirements to such n extent that no meaningful international search can be carried out, specifically:
3.	claims Nos.: ecause they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II C	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This Inter	national Searching Authority found multiple inventions in this international application, as follows:
1. A	s all required additional search fees were timely paid by the applicant, this international search report covers all earchable claims.
2. A	s all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite payment f any additional fee.
	s only some of the required additional search fees were timely paid by the applicant, this international search report overs only those claims for which fees were paid, specifically claims Nos.:
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	To required additional search fees were timely paid by the applicant. Consequently, this international search report is estricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark on	Protest The additional search fees were accompanied by the applicant's protest.
	No protest accompanied the payment of additional search fees.

Information on patent family members

Intern_ _ial Application No
PCT/US 93/08742

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